

Supporting Information

Quality criteria for microplastic effect studies in the context of risk assessment: A critical review.

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Methods continued - Detailed motivation for each criterion used in the quality evaluation.

Particle characterization

Criterion 1. Particle size. Species-specific physiological and behavioral traits can strongly influence the relative size of particles ingested by an organism, including MP.¹⁻⁴ Size selectivity depends on the morphology and feeding strategy of a species, which determines the upper size limit for the food they can ingest, as well as for the ingestible size of MP.⁵⁻⁷ For instance, in a study assessing the ingestion of MP by seven Cladocera species, the maximum size of MP ingested increased proportionally with the body size.⁵ The upper size limit will differ between species at varying trophic levels, but can also show significant variation within species depending on their developmental stage.^{1,8} Based on species traits, size preferences have been demonstrated for a few organisms, being some MP sizes ingested in higher quantities than others.^{2,7,9} Particle shape and polymer identity also affect the probability of MP to be encountered and ingested, thereby affecting the bioavailability of MP.¹ Furthermore, the residence time of MP in the body of the organisms has also been related with the size of the particles.¹⁰ The relative relationship between the ingestion and retention of MP can result in decreased nutritional value and/or physical obstruction in the digestive tract, which have been proposed as two of the mechanisms underlying observed adverse effects for organisms exposed to MP.^{2,3,8,11,12} As the ingestion and effects of MP can be size-dependent, the size distribution of the MP selected in an effect study can directly influence the occurrence and severity of the effects observed and therefore requires analytical characterization. Consequently, studies that report the full particle size distribution of the tested MP are assigned a criterion value of 2. The distribution, however, should be provided with sufficient resolution, ideally with 10 bins or more. If only one size is reported instead of a range, a study receives 2 points when the size reported is supported by analytical characterization and reported with a measurement error. MP sizes should ideally be characterized analytically using dynamic light scattering or laser diffraction methods or alternatively estimated using high resolution microscopy of the MP with a scale in combination with imaging analysis software. When the particle size/sizes are reported but not supported by analytical characterization, based on information provided in material safety data sheets or size separation using sieves, a study is assigned a criterion value of 1. Finally, studies that did not report the size of the MP used in their experiments are assigned a criterion value of 0.

Criterion 2. Particle shape. For several species, selective ingestion, gut retention, and effects of MP have been found to depend on their shape.^{8,10,13} For instance, fibers were more lethal than spheres for the amphipod *Hyaella azteca*.⁸ Authors report that fibers resulted in longer gut retention times, speculating that fibers may have aggregated in the gut.⁸ Additionally, Piarulli et al. (2020) showed that MP analyzed in six different benthic invertebrate species collected from salt marshes, were mostly fibers (98.5%).¹⁴ MP fragments are also reported to be associated with longer gut retention times in the

cladoceran *Daphnia magna* in comparison to spherical MP.¹³ It has been suggested that the rounded shape of spherical MP facilitates their transport through the digestive system of organisms, resulting in less severe effects than for other shapes of MP.⁸ Given several observations reporting on the relative influence of the shape of MP on effect endpoints, the evaluation criterion related to characterizing MP shape is seen as an important factor when interpreting ecotoxicological effects data. The shapes of MP have been defined in many ways, such as e.g., fragment, fiber, film, foam, pellet, sphere, line, bead, flake, sheet and granule.^{15–17} Different shape categories can be found even within these categories; for instance, MP fragments can be further characterized as rounded circular or edgy rectangular shapes.² Further complicating shape characterization is the observation that the dimensions of MP vary along continuous scales and therefore do not lend themselves well to discrete categories of characterization.⁶ Consequently, we consider the term “irregular MP” as an ambiguous definition of the shape, as it includes the potential to reflect several shape categories. Moreover, for a complete characterization of the shape, it is necessary to include at least one high-resolution photo illustrating each of the shapes included in the MP tested. Therefore, studies that provide an image obtained from a high-resolution microscope of the MP tested are assigned a criterion value of 2. Studies that limit the reporting of the shape of MP to the definitions of Rochman et al. (2019)¹⁶ or their synonyms (sphere vs. bead), based on the information obtained from material safety data sheets but without a visual confirmation by the authors are assigned a criterion value of 1. Finally, studies that do not report the shape of the MP used or reported shapes that did not fall within the definitions described by Rochman et al. (2019),¹⁶ are assigned a criterion value of 0.

Criterion 3. Polymer type. The fate, bioavailability, uptake and thus potential effects of MP can be also influenced by the composition of the polymer representing the MP, which determines the density of the particles in aqueous systems.^{18,19} In a sterile system without potential biofouling of the particles and in the absence of agitation, positively buoyant MP will float on the water surface, while negatively buoyant MP will remain in the water column until they sink to the bottom of the system.¹⁸ The fate of the MP in the water column thus influences their bioavailability and therefore the polymer type, as a proxy for density, needs to be characterized and reported. Additionally, knowing the polymer type will allow comparisons with field data on the occurrence, abundance and physical properties of the same polymer type, and possibly linking it with certain products and product emissions. Currently, elaborate techniques for polymer identification are available and widely applied in MP research, such as ATR-FTIR, micro-FTIR, Raman spectroscopy, pyrolysis GC-MS or similar methods.²⁰ For studies that analytically characterize the polymer type using one of these methods, a criterion score of 2 is assigned. When the polymer type is reported following the information given in the material safety data sheet and not confirmed by the authors, the study is assigned a criterion value of 1. Finally, studies that did not report the polymer type of the MP used are assigned a criterion value of 0.

Criterion 4. Source of MP. Reporting the source of where the MP were obtained is essential in order to better interpret the data the MP relate to, and to strengthen data reproducibility in future studies. Some studies, for instance, use in-house manufactured MP, following ad-hoc procedures which may not lend themselves well to reproducibility. In these instances it is imperative that detailed descriptions of the protocol used in producing the MP is provided (e.g., Korez et al., 2019).²¹ Results of effect studies on MP published to date show a wide variety of responses for different organisms.²² Even for the same species, different results can be obtained, which could be attributed to differences in the source(s) of MP.^{2,23} Therefore, when MP are purchased from a commonly available supplier and where specifics of the provider is provided in the main text or in the supporting information, a study is assigned a criterion value of 2, as this scenario lends itself best to reproducibility. For those studies where MP are prepared in-house using commercially available plastic products, we also assign a criterion value of 2 when the name of that plastic product is provided as well as a detailed protocol for the preparation or extraction of the MP. For instance, Jemec Kokalj, Kunej and Skalar (2018),²⁴ extracted MP from a facial cleanser and made MP from a plastic bag. Polymers were characterized using FTIR, particle size distributions were measured by laser diffraction, and images of the MP were taken with a field emission scanning electron microscope. However, they do not provide the name of the facial cleanser nor the precedence of the plastic bag. Consequently, when the information given on a MP source is incomplete and thus not fully reproducible, a criterion value of 1 is assigned. Finally, studies that do not provide any information on the source of the MP are assigned a criterion value of 0.

Criterion 5. Data reporting. It is widely acknowledged that inconsistency in how concentrations are reported make it difficult to compare between effects studies.^{22,25,26} Concentrations of MP can be presented as a particle number concentration, like the number of MP particles per L or per Kg of sediment, food or weight of the organism; or mass concentration, like grams of MP per L or per Kg of sediment, food or weight of the organism.²⁷ Some studies quantify the number of MP in a specific volume or weight using a hemocytometer, a flow cytometer or a coulter counter.²⁸⁻³¹ Other studies estimate the number of MP manually using a stereomicroscope combined with image analysis software, applicable for MP.^{29,31,32} Moreover, some studies convert mass concentrations to number concentrations or vice versa based on assumptions that correlate the size of a particle to its volume, for which MP characteristics such as size distribution, shape and density are required.^{2,33,34} A few other studies make reference to the conversion provided by the supplier of the MP.^{3,35,36} Thus, the reporting and conversion of concentrations between particle number and mass concentration units can be done using a variety of methods, and should be clearly described in the study in order to facilitate comparisons across studies. Since the units of concentration represent a fundamental parameter to assess risk, which compares environmental concentrations to effect threshold concentrations, consistency in units is therefore of paramount importance.^{27,37} Studies that report concentrations in particle number as well as in mass concentrations are thus assigned a criterion value of 2, as they provide the greatest opportunity to

compare between studies and for use in assessing environmental risk. Studies that limit the reporting of concentrations to only either particle number or mass concentrations, are assigned a criterion value of 1. Finally, studies where concentrations of MP are not reported receive a criterion value of 0.

Experimental design

Criterion 6. Chemical purity. Studies that aimed to investigate the interactive effects of MP and chemicals are not included in this study but are reviewed elsewhere.^{22,38,39} Persistent organic pollutants (POPs), such as polychlorinated biphenyls (PCBs) and organochlorine pesticides are ubiquitous in the environment and will partition into any organic carbon, including MP.³⁷ Experiments measuring the partitioning behavior between MP and organic chemicals are relevant for determining sorption/desorption coefficients and/or sorption kinetics. However, from the perspective of assessing risk it is more relevant to evaluate the toxicity of plastic-associated chemicals in the absence of MP.³⁷ Assessing the adverse effects of the chemical stressor in the absence of the MP individually first, can provide an effective strategy for developing more complex test systems aimed at assessing multiple chemical and non-chemical stressors, and help address the immediate challenges of assessing the environmental risks of MP themselves.³⁷ This reasoning also applies for the diversity of chemical additives and plasticizers commonly associated with plastic.⁴⁰ Moreover, disentangling the effect assessment associated with chemical stressors from the non-chemical particle stressor can strengthen overall understanding of the mechanisms that influence MP toxicity. For instance, studies by Martínez-Gómez et al., (2017) and Pikuda et al., (2019) have shown that the toxicity from leachates derived from additives are more harmful than the inert polymer material, highlighting the importance of washing MP before the start of an experiment, if insight regarding the effects associated with the particles themselves is the main objective.^{41,42} Otherwise, the chemical stressor overwhelms the effects that might be associated with the particles, preventing the ability to distinguish between the two. Additionally, the artificiality of an exposing test organisms to MP containing chemical additives within a closed system represents a worst-case scenario that is not representative of an environmentally relevant exposure. In the environment, organic chemicals, including POPs, chemical additives and plasticizers are widely dispersed as a consequence of their use in manufacturing and consumer products, and partition into all environmental media, resulting in various exposure pathways to exist. Consequently, assessing chemical exposure requires an understanding of the multimedia behavior of organic chemicals, whereby exposure via MP likely represents a negligible pathway as compared to other sources.^{38,43} Therefore, in order to disentangle the effects associated with the particle stressor from confounding chemical effects, the toxicity of background chemicals should be minimized. This includes minimizing exposure to chemical additives and plasticizer that might be present in MP, but also chemicals associated with food particle surfactants (e.g., Tween) and markers (fluorescence). Minimizing chemical exposures in MP effects studies, however, represents a major challenge. For instance, a recent study by Cole et al. (2019) extensively measured chemicals in MP, and reports that a wide variety of unknown chemicals are used

in MP, making it nearly impossible to confirm conclusively that all relevant chemicals have been assessed.⁴⁴ Therefore, it is preferred to repeatedly wash the particles with an organic solvent(s) in an effort to minimize effects associated with a chemical-associated contaminant. It is notable, however, that this could have the undesired effect of altering the properties of the particles themselves, consequently care is required with respect to which organic solvents are used as well as the conditions of cleaning. Alternatively, several studies have demonstrated that it is possible to minimize the influence of the chemical stressor by providing evidence that the mass of chemical in the test system is at an exposure that remains below a chemical toxicity (e.g., Bellingeri et al., 2019; van Weert et al., 2019; Redondo-Hasselerharm et al., 2020).⁴⁵⁻⁴⁷ In summary, studies that report the inclusion of methods to thoroughly clean MP by washing with an organic solvent are assigned a criterion value of 2, since the observations of adverse effects could be more confidently allocated to a particle-associated effect. If a certificate from the manufacturer was used or measurements were taken to subsequently use a control for the chemicals or the toxicity of chemicals was calculated based on L_{50} or EC_{50} from literature, the study is assigned a criterion value of 1. Finally, studies that did not address the potential influence of a chemical stressor on observed adverse effects when testing MP are assigned a criterion value of 0.

Criterion 7. Laboratory preparation. The importance of preventing contamination when testing MP is emphasized in several recent papers and critical reviews.^{15,48-52} Catarino et al. (2018), for instance, quantified atmospheric fall-out within households, which, when rescaled to the surface area of a representative experimental test system of e.g. 20 x 25 cm², would imply a flux of 8333 particles per test system per day.⁵³ The amount of natural fall-out of MP likely differs between locations within a laboratory and among laboratories. Catarino et al. (2018), emphasized the need to account for atmospheric deposition during experiments, even in instances where relatively high concentrations are tested.⁵³ We argue, therefore, that the uncertainty related to contamination with MP during MP effects studies, also requires care in mitigating the potential for deposition and with respect to characterizing and quantifying the nature of the contaminants. This is because the nature of the MP-contaminants may be significantly different than those used in the test system, in that they may contain chemical additives that can strongly influence observed effects, negating test results. This is particularly relevant to the control test-system, meant to have zero MP concentration, or very low dosed systems, for which greater sensitivity would be anticipated due to the influence of MP-contaminants. Some studies thoroughly report measures taken to prevent MP-contaminates, such as wearing cotton lab coats, rinsing of equipment, covering the test systems or avoiding the use of plastic materials during the experiment.⁵⁴⁻⁵⁶ Consequently, a criterion value of 2 is assigned for those studies adopting measures aimed at avoiding contamination from air, water and all materials used during the experiment. Studies adopting limited measures are assigned a criterion value of 1. Finally, studies that do not report the use of any measure to prevent contamination are assigned a criterion value of 0.

Criterion 8. Verification of background contamination. Whereas the previous criterion focuses on the measures taken to mitigate background MP-contaminants, the present criterion evaluates the extent to which studies verify that such measures are successful or alternatively that the adoption of taking no action to reduce background contamination is needed because the potential for MP-contaminants is demonstrated to be minimal. In this case, verification implies the use of methods that characterize and analytically measure MP concentrations in exposure systems. A study by Welden and Cowie, 2016, for instance, observed a fiber in the foregut of one of their control animals, underlining the importance of including method verification in MP effects test studies⁵⁷. A few studies, on the other hand, have limited verification of background contamination to the reporting of visual observations.^{54,55,58} Visual inspection, however, is generally considered inaccurate, as there is a high probability of missing small and transparent MP.^{15,51} Moreover, reliance on the use of visual observations is susceptible to false positives.²⁰ Based on these considerations, a criterion value of 2 is assigned to studies measuring background contamination with analytical detection methods, such as by FTIR or Raman. For studies that limit the verification of background MP-contaminants to a visual inspection, a criterion value of 1 is assigned. Finally, for studies that do not report on background contamination of MP, a criterion value of 0 is assigned.

Criterion 9. Verification of exposure. In order to obtain accurate dose-effect relationships, exposure concentrations in the test systems must be quantified. Test concentrations are typically prepared by adding particles to the test medium, occasionally followed by dilution and homogenization steps. There are several reasons why the actual exposure concentration can deviate from the nominal concentration estimated from the initial preparation. First, human error can occur in the initial calculations or laboratory manipulations of glass ware and equipment can lead to deviations in the concentration. Secondly, the test system itself can influence exposure, whereby particles can stick to container walls and/or become unevenly distributed across test systems when homogenization is insufficient. Actual concentrations can also be higher than nominal concentrations due to background MP-contaminants, as discussed in the previous criterion.^{53,59} These factors can propagate and substantially influence initial estimates of test concentrations. Furthermore, the dynamic behavior of the particles themselves can cause significant changes in exposure during the test. While less important for sediment-test systems, the behavior of particles in aqueous test systems can result in settling, floating or aggregation of the particles, changing the actual exposure conditions over time.⁶⁰ Fundamentally, the exposure of the stressor in an ecotoxicity test system should be constant over time and reproducible for each test. Demonstrating consistency in the exposure concentrations for the duration of the test is thus important to develop accurate dose-effect relationships, and the quantification of the exposure concentration should therefore be verified. A criterion value of 2 is assigned to studies that verify the exposure concentration of MP and ensure that at least 80% of the nominal concentration is maintained throughout the test.^{61,62} Studies that measure the exposure concentration, but without verifying that at least 80% of

the nominal concentration is maintained throughout the test are assigned a criterion value of 1. Studies that only report the nominal concentration or limit the verification of the concentration to the stock solution are assigned a criterion value of 0.

Criterion 10. Homogeneity of exposure. The previous criterion evaluates the extent to which the exposure concentration is verified. However, unlike the fate of dissolved chemicals in ecotoxicological effect testing, solid particles are prone to inhomogeneity of exposure as they tend to settle or float depending on a variety of factors, such as the difference in their density compared to that of the medium they are dispersed in.^{63–70} Therefore, especially for aqueous test systems, MP that have a higher density than water may settle when the dispersion is not well mixed, whereas buoyant particles may tend to reside at the surface of the test system only. Presence of air pockets or biofilm layers may change over time and influence exposure as a result of settling or causing differences in particle-particle interactions and settling velocities as a function of time, thus questioning the assumption of exposure homogeneity. These inhomogeneities can strongly influence the bioavailability and thus the exposure of the particles, resulting in a lack of control and reproducibility of test results. Methods for addressing heterogeneity in test systems assessing particle stressors include the use of ultrasonic agitation, and other physical mixing techniques (circular, wrist action shaking, plankton wheels) prior or during exposure, or by simply reporting the absence of such problems based on visual observations.^{33,44,71–74}

Natural sediments are comprised of a mixture of particles with densities spanning a wide range, as compared to that of the solid polymeric particles that have been tested. MP mixed in sediment are ‘held’ in the sediment matrix and progressively encapsulated when biofilms form and test particles form hetero-aggregates and – agglomerates with the natural particles in the sediment matrix. This implies that exposure in effects test systems of MP mixed in sediment are homogeneously distributed. Many studies have recognized the need for homogeneity and have described in detail how MP were mixed in the exposure medium and sometimes also how homogeneity of exposure was verified.^{47,75} For aqueous exposures, a criterion value of 2 is assigned to studies that verify that MP were homogeneously distributed through the use of microscopy photos and/or apply analytical tools to demonstrate that the MP were well mixed or dispersed in the solution. In instances where the method used to generate a homogeneous exposure is described but not verified, a criterion value of 1 is assigned. Effect testing of MP in sediment test systems, for which the verification of homogeneity is deemed to be not crucial, results in a criterion score of 2 for all studies that describe the method by which the MP are homogeneously mixed with the sediment, in detail. Studies that do not address the issue of homogeneity, or that observed an inhomogeneous exposure, are assigned a criterion value of 0.

Criterion 11. Exposure assessment of organisms. To be able to understand and interpret effect data, it is important to be able to causally link an observed effect to actual exposure data. The question ‘what is

an organism exposed to?’ however can have different answers for different organisms, particles and/or test conditions. The metric used to quantify the effect should be ecologically relevant and should be the same as the one used to quantify exposure.³⁷ Microplastics can have multiple of such environmentally relevant metrics (ERMs). They can be characterized on the basis of known species- and particle-specific effect mechanisms. Hence, it is the actual effect mechanism which defines how microplastic particles and test organisms interact and how actual exposure should be assessed. Exposure then can be seen as accumulation at the receptor site, i.e. where the interaction takes place, and which is considered as the target for the microplastic effect under consideration. We illustrate the principle with three examples. For instance, one of the more well understood effect mechanisms, is the deterioration of food quality due to the dilution of nutritious food particles caused by an elevated exposure to low-caloric, non-digestible MP that are co-ingested with food (Redondo-Hasselerharm et al., 2018; Wright et al., 2013).^{2,12} Therefore, for a study that would ascribe observed effects to this mechanism, demonstrating ingestion would be a crucial criterion. Instead, studies that ascribe suborganismal effects to damage at the cell level^{73,76–78} should ideally demonstrate systemic uptake and/or penetration of MPs and should demonstrate that these cells are reached. As a final example, studies that explain growth inhibition in algal cultures from a decrease in photosynthesis, should verify the presence of MPs at or in between algal cells in the culture.^{79,80} A detailed overview and analysis of such reported effect mechanisms is provided in section 3.3 of his review. In the majority of instances, effects related to the ingestion of MP are reported as the most relevant exposure pathway, implying that the quality criteria to detect and quantify MP ingested by biota are of critical importance. Exposure due to translocation and cell penetration also requires detection and quantification of MP in biota tissue and is thus also important in defining the quality criteria. These criteria have been reported in a previous study,⁵¹ for which criteria related to tissue digestion, particle detection and polymer identification are all applicable. For adverse effects influenced by external exposure of MP, i.e. from MP just being present in water or sediment, as in the example for algae, criteria for the analysis and quantification of MP in water are most relevant. It is widely understood that visual sorting of MP is insufficient to detect the small and often light-colored MP against a background of e.g., animal tissue. Therefore, following the QA/QC criteria suggested by Hermsen et al. (2018), a criterion value of 2 is assigned to studies that report the detection of MP quantitatively using e.g., FTIR or Raman imaging, to support statements of MP ingestion and/or penetration into cells of biological tissues that have been appropriately digested and filtered.⁵¹ Studies demonstrating exposure of organisms to MP based on qualitative or visual observation, or citing results from a separate experiment, or in the absence of a digestion step, are assigned a criterion value of 1. Studies that do not report data on exposure, are assigned a criterion value of 0.

Criterion 12. Replication. In every effect assessment, an adequate experimental design requires a sufficient number of replicates in order to ensure statistically reliable results.^{81,82} Studies should therefore clearly explain the degree of replication of each treatment.⁸¹ Some studies, however, fail to

report on the use of replicates in their experimental design^{24,57} while other studies report the use of replicates, but which are not actual replicates but better characterized as pseudo-replicates.^{70,83,84} For instance, Jovanović et al. (2018) considered as replicates the 15 fish exposed to MP in the same tank.⁸⁵ As each replicate should be an independent experimental unit, with the experimental unit here being the tank, the exposure of all fish via the same tank should thus be better defined as multiple measurements taken one experimental unit.⁸⁶ In contrast to soluble chemicals, which can be homogeneously distributed in the test system, the severity of the effects detected in MP studies can be attributed to the relative extent of bioavailability of the particles and the probability of encountering them in the test system. Therefore, in the case of MP, it is especially important to have several replicates to compensate for the uncertainties associated with the potential for inhomogeneous exposure associated with the test system. Studies were assigned a value of 2 when they included results from a minimum of three replicates. A criterion value of 1 is assigned to studies using only two replicates. Finally, studies that do not include any replicates or do not report the number of replicates used are assigned a criterion value of 0.

Applicable to risk assessment

Criterion 13. Endpoints. Effect studies with MP use a wide variety of endpoints, sometimes even within studies. We argue that when data from such studies are to be used in ecological risk assessment, the ecological relevance of the selected endpoint represents an important criterion to consider. From a risk assessment perspective, endpoints such as survival, growth and reproduction are considered ecologically relevant, because these endpoints directly relate to a population-level effect. These endpoints are preferred over e.g. suborganismal or behavioral endpoints, which are generally less relevant in assessing population-level responses, unless there is a clear demonstrated causal relationship between these responses and a higher level effect e.g. population effect.^{25,87} For instance, de Sá et al., (2015) speculated that reduced food intake caused by the ingestion of MPs adversely affects both the individual and population-level fitness of a species.⁸⁸ The endpoints studied, however, are attributed to the predatory performance and efficiency of the species, which does not necessarily translate to an ecologically relevant population level effect. Whereas it has been suggested that suborganismal endpoints such as biomarkers can be representative of early warning signals and are thus more sensitive indicators than the traditional endpoints used in risk assessment^{55,89}, they can also be perceived as being susceptible to type I and II error, due to under-replication and pseudo-replication in ecotoxicological bioassays, which could lead to false alarms or undetected effects.^{89,90} Moreover, there is no evidence that suborganismal endpoints are more sensitive than endpoints taken at higher organismal level responses, particularly for MP effects studies. Additionally it is possible that effects seen at the suborganismal level merely resemble reaction to decreased nutritional intake.⁹¹ Furthermore, endpoints at these suborganismal levels are not likely to be useful predictors since they have complicated time- or dose-dependent responses, which makes it difficult to extrapolate correlations to higher levels of biological organization.⁸⁹ Still, in carefully controlled studies e.g. biomarkers can be useful for elucidating mechanisms of toxic action.⁸⁹

In summary, a criterion value of 2 is assigned to studies where endpoints at either community or individual level of biological organization (e.g. survival, growth, development or reproduction) are used. If suborganismal endpoints are used, for which a causal relationship with effects on higher levels of biological organization is demonstrated, a criterion value of 1 is assigned to the study. Finally, studies that use endpoints that cannot be unambiguously linked to a threat at the individual or population level are assigned a criterion value of 0.

Criterion 14. Presence of natural (food) particles. It is important to note that the natural environment is not free of particles and that organisms have adapted various species-specific traits in relation to strategies for interacting with particles. While MP are ubiquitous in the aquatic environment, the amount of natural particles is typically greater than the concentrations of MP that have been reported in the environment.^{38,92} Therefore, when designing an experiment meant to simulate natural conditions it is important to consider the response of organisms to both naturally occurring particles as well as a MP-stressor exposure.^{25,91} Exposure to naturally occurring particulates, for instance, can represent an important food source to an organism or may otherwise form part of their natural habitat, such as sediment or suspended solids.^{2,47} The inclusion of food and other particulates is needed because ecotoxicological effects of MP on organisms has been demonstrated to be influenced by the presence of naturally occurring particulates.^{1,70,93,94} Observations that the co-exposure of both naturally occurring particulates and MP can mitigate toxicity implies the relative importance of a species ability to selectively feed and therefore reduce the risks associated with MP under environmentally relevant conditions.⁹¹ We argue that without taking natural (food) particles into account, the observed adverse effects represent a system-dependent artefact that does not lend itself to risk assessment purposes. An exception, however, is made for algal studies, as their food source are nutrients and light, and therefore the addition of other naturally occurring particles is less likely to influence adverse effects.⁹⁵

It is further noted that there are several studies that adopt standard test protocol guidelines for acute toxicity testing, which are applicable to soluble chemicals.^{24,34,96,97} In such experiments the test guidance is not to feed the organisms, which is logical when testing soluble chemicals as the food particles may influence the bioavailability of the test chemical and the presence of food does not represent a limiting factor due to the short duration of the acute study. However, this guidance is not applicable to experiments aimed at assessing the acute response of MP, because the adverse effects can also potentially be influenced by the presence of food particles.⁹¹ Therefore, when natural particles (at least food) are added to avoid an exposure that might be perceived as analogous to ‘force feeding’ the organisms with MP, a criterion value of 2 is assigned to the study. Studies that add food, but in which the food is not optimally available to the organisms are assigned a criterion value of 1. Finally, studies that do not include any naturally occurring or food particles are assigned a criterion value of 0.

Criterion 15. Reporting of effect thresholds. To date, the majority of effect studies report adverse effects for MP at a single or limited number of test concentrations.^{11,88,98–100} These observations are beneficial in demonstrating the potential adverse effects that MP can have on biota. It remains unclear, however, the threshold concentration above which the adverse effect initiates. For the purposes of risk assessment, where the ratio of exposure concentrations to that of effect threshold concentrations are derived, accurate estimates of effect threshold concentrations, such as derived from dose-response relationships in the form of L(E)C_x, (or the generally less preferred NOEC or LOEC),^{101–103} are required. Given the paucity of dose-response threshold effects data for MP, the need for effect threshold concentrations to help inform the risk assessment process has been widely recognized.^{22,27,104} Therefore, given the relative importance of this criterion regarding applicability in risk assessment, effect studies aiming at reporting effect thresholds are assigned the greatest value. To be effective it is notable that effect threshold concentrations must be accompanied with estimates of error or uncertainty, in order to evaluate that differences in exposure concentrations are statistically meaningful. Based on this reasoning, we assign a criterion value of 2 to studies that report threshold effects data using L(E)C_x derived from dose-response relationship modelling, with error data (95% confidence interval, standard error or standard deviation). If other metrics like NOEC or LOEC are used, or when no error data are provided, the data are still considered useful and a criterion value of 1 is assigned. Studies that do not explicitly provide data on threshold concentrations for the reported effects are assigned a criterion value of 0.

Criterion 16. Quality of the dose-response relationship - Effect threshold concentrations, such as EC₅₀ or LC₅₀, are typically obtained by fitting a logit or probit model to dose-response data,¹⁰⁵ in which EC₅₀ or LC₅₀ is a model parameter. This implies that the statistical significance of the resulting EC₅₀ or LC₅₀ value depends on the quality of the fit to the data, and on the number of parameters fitted, compared to the number of data points in the dose-response relationship. In standard ecotoxicity test systems it is generally suggested to assess effects using a minimum of six different exposure dose concentrations, including the control, to obtain an accurate EC₅₀ or LC₅₀ value.¹⁰⁵ Ideally, the exposure concentrations used are representative of the full range of effects, i.e. from low effect to near-maximum effect, such that an EC₅₀ or LC₅₀ value can be derived without extrapolation. Intuitively, replication of test results at each exposure concentration will also contribute to more accurate EC₅₀ or LC₅₀ values. Since replication is already covered by criterion 12, only the number of exposure concentrations used in an effect study is evaluated under this criterion. Studies that use the recommended minimum of six exposure dose concentrations or more, including a treatment control (zero microplastic concentration), are assigned a criterion value of 2, and a criterion value of 1 if five different concentrations are used. For studies reporting dose-response relationships using less than five concentrations, a criterion value of 0 is assigned.

Criterion 17. Concentration range tested. Recent studies have drawn attention to the need to better define ecologically relevant concentration ranges for effect testing of MP.^{26,106} As previously discussed, studies reporting adverse effects for MP often use unrealistically high exposure concentrations, which has resulted in suggestions for future studies to assess effects using lower, more environmentally relevant, concentrations.^{37,106} However, if studies limit assessing effects to low concentrations, it is possible that derivation of effect threshold concentrations may not be possible. Consequently, we argue that studies must follow standard principles adopted in assessing the risks of chemicals, such as through the use of quantitative dose-effect relationships to obtain an assessment of effect threshold endpoints typical of ecotoxicology (i.e., EC₅₀ or LC₅₀) with sufficient quality. To meet this requirement, effect testing can include both high and low concentrations, as long as the results are used to quantitatively derive the appropriate threshold values. For example, if an effect observed in an ecotoxicity test system occurs only at concentrations that exceed environmentally relevant exposure concentrations by several orders of magnitude, the end result would be supportive of demonstrating low risk. Nevertheless, there can also be strong arguments that support the use of environmentally realistic, low concentrations in ecotoxicity effects tests. This is because the reported effects occurring at high concentrations may be linked to an effect associated with a decrease in food quality, resulting from either the ingestion of inert non-digestible particles or due to an overwhelming number of particles in the test system that results in a decreased potential for the test organisms to find food particles. This type of effects occurs with any type of particle of low nutritional value and may be perceived as an artefact of the test system design, not an effect that is intrinsic to the MP themselves^{25,33} and is therefore better understood as a non-specific particle effect. This exposure scenario typically results in the test organisms suffering from starvation prior to any other modes of action that the MP may cause – effects that might occur at lower concentrations following a chronic exposure.^{47,107} In other words, at environmentally relevant concentrations, it is unlikely that food dilution represents a mechanism of ecological significance, but that more subtle effect mechanisms (related to behavior, avoidance, reproduction, particle toxicity) are likely of greater relevance to assess and for which long term chronic effects testing would be beneficial. For this reason, some studies intentionally assess the effects associated with lower test concentrations.^{2,32} In summary, environmentally relevant concentrations should be given priority for effects testing of MP, which forms the basis of a legitimate criterion for the ecological relevance associated with chronic ecotoxicity test system design. Note that exposure duration is evaluated below, in a separate criterion, and only the ecological relevance of the concentration is evaluated under this criterion. Thus, studies that use two or more environmentally realistic concentrations in the exposure concentration doses tested, supported by credible literature data, are assigned a criterion value of 2. If the test system uses only a single environmentally relevant concentration, supported by credible literature data, a criterion value of 1 is assigned. Studies that acknowledge that concentrations are far above environmentally relevant concentrations, or that do not evaluate their exposure concentrations with environmental monitoring data, are assigned a criterion value of 0.

Criterion 18. Aging and biofouling. Under environmentally relevant conditions, MP undergo abiotic and biotic processes that alter their shape, size, structure and eventually their bioavailability.¹⁰⁸ Vroom et al. (2017) demonstrate that the aging of MP promotes their ingestion by marine zooplankton.¹⁰⁹ As the surface of MP functions as a substrate for biofilm to grow, ingestion of biofouled MP potentially represents an additional energy source for test organisms.¹¹⁰ This implies that ecotoxicity tests that assess pristine particles may potentially underestimate the ingestion rates that may occur in the environment, whereby the potential to ingest aged and biofouled particles may be higher. Since MP undergo both aging and biofouling in the environment, it would thus be beneficial to consider how such processes influence ecotoxicity results and would further strengthen aims directed at ecological relevance. Consequently, studies that include aging of MP to make them more environmentally realistic and also characterized the MP for aging and biofouling, for instance by scanning electron microscope (SEM), are assigned a criterion value of 2. Studies that have only aged the MP but do not characterize them (e.g., Zettler et al. (2013))¹¹¹ are assigned a criterion value of 1. Finally, studies that limit testing to only the use of pristine MP and/or conditions that prevent the formation of a biofilm are assigned a criterion value of 0.

Criterion 19. Diversity of MP tested. To date, most studies assessing the effects of MP limit observations to a relatively small sub-set of all possible characteristics. For instance, studies testing MP based on a single or limited range of particle sizes, shapes and polymeric type may provide valuable information on how specific particle characteristics influence uptake and effects, but under ecologically relevant conditions, organisms will encounter a wide variety of characteristics, of which size, shape and density often are considered the most important properties influence the transport, fate and bioavailability of MP.^{6,104,112} Species-specific biological and behavioral traits can also play an important factor in determining which properties of MP found in the environment will most likely result in an exposure for the individuals of a species.^{2,104} The ecotoxicological effects related to the properties of the relevant fraction of MP for a species, may also be influenced by the presence of either other MP or of naturally occurring particles. Simulating species-specific responses to exposures of environmentally relevant heterogeneous mixtures of both MP and naturally occurring particles represents a significant challenge in MP effects testing. Recently, Kooi and Koelmans (2019) reviewed the ranges and distributions of the characteristics of environmentally relevant MP and observed relative similarity across datasets taken from different locations, with respect to their physicochemical characteristics.⁶ Given the recent awareness associated with this criterion, we suggest that future studies adopt the use of distributions in physicochemical properties of MP as a standard approach to enable better environmental realism in MP effects testing. Consequently, studies that use MP with a range of sizes, shapes and densities in one mixture exposure, and which attempts to simulate the diversity of environmental MP, are assigned a criterion value of 2. If the diversity related to only one or two of the physicochemical characteristics

and/or a limited distribution, a criterion value of 1 is assigned. Studies that limited effect testing to a single type of MP were assigned a criterion value of 0.

Criterion 20. Exposure time. Standard test protocol guidelines for the ecotoxicity testing of chemicals recommend the application of defined exposure times for each of the endpoints assessed. While these guidelines are also routinely adopted in the effects testing of MP, some studies highlight the need for longer exposure times, due to the detection of time-dependent effects.^{34,47,84,107,113–115} For instance, the effects of MP on the freshwater coral *Lophelia pertusa* differed between exposure times of 7, 20 and 47 days. While the coral growth rate decreased over time, effects on capture prey and polyp activity disappeared after 47 days, revealing that both positive and adverse effects of MP can differ with time.⁸⁴ Furthermore, observations for the marine mussel *Mytilus edulis*, report the formation of granulocytomas and the destabilization of the lysosomal membrane increased significantly with longer exposure times when exposed to MP.¹¹⁴ Moreover, adverse effects of MP on the growth of the cladoceran *Daphnia magna* were only found after 25-31 days of exposure.¹¹³ For *D. magna*, another study demonstrated that their immobilization increased over time when exposed to MP.³⁴ Generational effects following exposure to MP have also been reported, as in the case of the copepod *Tigriopus japonicus*.¹¹⁵ Therefore, the importance of exposure duration, which can influence the detection of adverse effects that might differ between chemicals and MP is emphasized within this evaluation criterion. Exposure duration is of particular importance for endpoints that seem to be time-dependent, such as growth, reproduction and long term community effects.^{47,116} Additionally, increasing the exposure time can be perceived as adding greater environmental relevance to the effect study, explaining the logic for why this criterion is in the ecological relevance category. Thus, for studies that include a minimum exposure time of 7 days for bacteria or phytoplankton, 21 days for zooplankton, 28 days for benthic invertebrates, macrophytes or fish larvae and 3 months for adult fish, the study is assigned a criterion value of 2. For studies that use an exposure time between 1 and 7 days for bacteria or phytoplankton, between 4 and 21 days for zooplankton, between 7 and 28 days for benthic invertebrates, macrophytes or fish larvae and between 1 and 3 months for adult fish, a criterion value of 1 is assigned. Finally, studies that use substantially shorter exposure times, specifically <1d for bacteria and phytoplankton, 4 days for zooplankton, 7 days for benthic invertebrates, macrophytes or fish larvae and 1 month for adult fish, are assigned a criterion value of 0, except in instances where multigenerational studies are performed, where a criterion value of 1 is assigned

Table S1. Study characteristics.

| Author | Year | System | Plant/Algae/ Invertebrate /Fish | Class studied | Species | Polymer type | Shape | Size (µm) | Exposure duration (h) | Endpoints studied | Endpoints affected | Effect threshold |
|------------------------------------|------|--------|---------------------------------------|---------------|----------------------------------|-----------------|----------|--------------|-----------------------------|--|--------------------------------------|---------------------|
| <i>Aljaibachi et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PS | Sphere | 2 | 504 | Survival, Growth & Reproduction | Survival, Growth | |
| <i>Au et al.</i> | 2015 | Fresh | invertebrate | Malacostraca | <i>Hyalella azteca</i> | PE | Sphere | 10 - 27 | 240 | Survival | Survival | LC ₅₀ |
| <i>Au et al.</i> | 2015 | Fresh | invertebrate | Malacostraca | <i>Hyalella azteca</i> | PE | Sphere | 10 - 27 | 672 | Growth, Reproduction | Growth, Reproduction | |
| <i>Au et al.</i> | 2015 | Fresh | invertebrate | Malacostraca | <i>Hyalella azteca</i> | PP | Fiber | 75-20 (x 20) | 240 | Survival | Survival | LC ₅₀ |
| <i>Au et al.</i> | 2015 | Fresh | invertebrate | Malacostraca | <i>Hyalella azteca</i> | PP | Fiber | 75-20 (x 20) | 240 | Growth, Egestion time | Growth, Egestion time | |
| <i>Blarer & Burkhardt-Holm</i> | 2016 | Fresh | invertebrate | Malacostraca | <i>Gammarus fossarum</i> | PS | Sphere | 1,6 | 672 | Feeding rate, Assimilation efficiency, Wet weight change | Assimilation efficiency | |
| <i>Blarer & Burkhardt-Holm</i> | 2016 | Fresh | invertebrate | Malacostraca | <i>Gammarus fossarum</i> | PA | Fiber | 500 x 20 | 672 | Feeding rate, Assimilation efficiency, Wet weight change | Assimilation efficiency | |
| <i>Bosker et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PS | Sphere | 1 - 5 | 504 | Population abundance, biomass | Population abundance, biomass | |
| <i>Bour et al.</i> | 2018 | Marine | invertebrate | Bivalve | <i>Ennucula tenuis</i> | PE | Fragment | 4 - 6 | 672 | Survival, Condition Index, Burrowing behavior | | |
| <i>Bour et al.</i> | 2018 | Marine | invertebrate | Bivalve | <i>Ennucula tenuis</i> | PE | Fragment | 20 - 25 | 672 | Survival, Condition Index, Burrowing behavior | Total energy | |
| <i>Bour et al.</i> | 2018 | Marine | invertebrate | Bivalve | <i>Ennucula tenuis</i> | PE | Fragment | 125 - 500 | 672 | Survival, Condition Index, Burrowing behavior, Total energy, Protein content | Total energy | |
| <i>Bour et al.</i> | 2018 | Marine | invertebrate | Bivalve | <i>Abra nitida</i> | PE | Fragment | 4 - 6 | 672 | Survival, Condition Index, Burrowing behavior | | |
| <i>Bour et al.</i> | 2018 | Marine | invertebrate | Bivalve | <i>Abra nitida</i> | PE | Fragment | 20 - 25 | 672 | Survival, Condition Index, Burrowing behavior | | |
| <i>Bour et al.</i> | 2018 | Marine | invertebrate | Bivalve | <i>Abra nitida</i> | PE | Fragment | 125 - 500 | 672 | Survival, Condition Index, Burrowing behavior, Total energy, Protein content | Protein content | |
| <i>Browne et al.</i> | 2008 | Marine | invertebrate | Bivalve | <i>Mytilus edulis</i> | PS | Sphere | 3 | 3 | Feeding behavior, Phagocytic activity, ROS production | | |
| <i>Browne et al.</i> | 2008 | Marine | invertebrate | Bivalve | <i>Mytilus edulis</i> | PS | Sphere | 9,6 | 3 | Feeding behavior, Phagocytic activity, ROS production | | |
| <i>Bruck & Ford</i> | 2018 | Marine | invertebrate | Malacostraca | <i>Echinogammarus marinus</i> | PS | Sphere | 8 | 840 | Food consumption, Growth & Moulting | | |
| <i>Caniff & Hoang</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Sphere | 63 - 75 | 504 | Survival, Reproduction rate | | |
| <i>Capolupo</i> | 2018 | Marine | invertebrate | Bivalve | <i>Mytilus galloprovincialis</i> | PS | Sphere | 3 | 48 - 916 | Feeding efficiency, Morphological development, Gene transcription | Gene transcription | |
| <i>Chae et al.</i> | 2019 | Marine | algae | Chlorophyceae | <i>Dunaliella salina</i> | PE | Sphere | 180 - 212 | 144 | Cell growth, Photosynthetic activity, Cell morphology | Cell growth, Photosynthetic activity | |
| <i>Chapron et al.</i> | 2018 | Marine | invertebrate | Anthozoa | <i>Lophelia pertusa</i> | LDPE | Sphere | 500 | 168 | Capture rates | Capture rates | |
| <i>Chapron et al.</i> | 2018 | Marine | invertebrate | Anthozoa | <i>Lophelia pertusa</i> | LDPE | Sphere | 500 | 480 | Polyp activity | Polyp activity | |

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|-----------------------------|------|-----------|--------------|----------------------|------------------------------------|-------|-----------|-------------|------|--|---|
| Chapron et al. | 2018 | Marine | invertebrate | Anthozoa | <i>Lophelia pertusa</i> | LDPE | Sphere | 500 | 1656 | Skeleton growth rate | Skeleton growth rate |
| Choi et al. | 2018 | Marine | fish | Actinopterygii | <i>Cyprinodon variegatus</i> | PE | Sphere | 150 - 180 | 96 | Survival, Malformation, Swimming behavior, Oxydative stress, Gene transcription, Enzyme activity | Oxidative stress, Gene expression, Enzymatic activity |
| Choi et al. | 2018 | Marine | fish | Actinopterygii | <i>Cyprinodon variegatus</i> | PE | Irregular | 6 - 350 | 96 | Survival, Malformation, Swimming behavior, Oxydative stress, Gene transcription, Enzyme activity | Swimming behavior, Oxydative stress, Gene expression |
| Chritchell & Hoogenboom | 2018 | Marine | fish | Actinopterygii | <i>Acanthochromis polyacanthus</i> | PET | Irregular | 1000 - 2000 | 1008 | Growth, Body condition & Behavior | |
| Cole & Galloway | 2015 | Marine | invertebrate | Bivalve | <i>Crassostrea gigas</i> | PS | Sphere | 1 | 192 | Feeding rate, growth | Feeding rate |
| Cole & Galloway | 2015 | Marine | invertebrate | Bivalve | <i>Crassostrea gigas</i> | PS | Sphere | 10 | 192 | Feeding rate, growth | |
| Cole et al. | 2013 | Marine | invertebrate | Hexanauplia | <i>Centropages typicus</i> | PS | Sphere | 7,3 | 24 | Feeding rate | Feeding rate |
| Cole et al. | 2013 | Marine | invertebrate | Hexanauplia | <i>Centropages typicus</i> | PS | Sphere | 20,6 | 24 | Feeding rate | |
| Cole et al. | 2015 | Marine | invertebrate | Hexanauplia | <i>Calanus helgolandicus</i> | PS | Sphere | 20 | 216 | Survival, Reproductive output, Egg production rates, Respiration rate | Reproductive output |
| Cole et al. | 2019 | Marine | invertebrate | Hexanauplia | <i>Calanus finmarchicus</i> | Nylon | Fiber | 10 x 30 | 144 | Feeding, Prosome length, Moulting, Lipid accumulation | Feeding selectivity, Moulting |
| Cole et al. | 2019 | Marine | invertebrate | Hexanauplia | <i>Calanus finmarchicus</i> | Nylon | Granule | 10 - 50 | 144 | Feeding, Prosome length, Moulting, Lipid accumulation | Moulting |
| Cong et al. | 2019 | Marine | fish | Actinopterygii | <i>Oryzias melastigma</i> | PS | Sphere | 10 - 11 | 336 | Survival, Growth, Reproduction | Survival, Growth, Reproduction |
| de Sá et al. | 2015 | Estuarine | fish | Actinopterygii | <i>Pomatoschistus microps</i> | PE | Sphere | 420 - 500 | 0,05 | Predatory performance & efficiency | Predatory performance & efficiency |
| Detree & Gallardo-Escarrate | 2017 | Marine | invertebrate | Bivalve | <i>Mytilus galloprovincialis</i> | PE | Sphere | 1 - 50 | 24 | Gene expression | Gene expression |
| Espinosa et al. | 2017 | Marine | fish | Actinopterygii | <i>Sparus aurata</i> | PVC | N/A | 40 - 150 | 720 | Growth, immune status, liver damage | Immune status, liver damage |
| Franzellitti et al. | 2019 | Marine | invertebrate | Bivalve | <i>Mytilus galloprovincialis</i> | PS | Sphere | 3 | 48 | Multixenobiotic resistance | Multixenobiotic resistance |
| Franzellitti et al. | 2019 | Marine | invertebrate | Bivalve | <i>Mytilus galloprovincialis</i> | PS | Sphere | 3 | 96 | Multixenobiotic resistance | Multixenobiotic resistance |
| Franzellitti et al. | 2019 | Marine | invertebrate | Bivalve | <i>Mytilus galloprovincialis</i> | PS | Sphere | 45 | 96 | Multixenobiotic resistance | Multixenobiotic resistance |
| Gambardella et al. | 2019 | Fresh | bacteria | Gammaproteo bacteria | <i>Vibrio fischeri</i> | PE | Irregular | 4.25 | 0,5 | Bioluminescence | NOEC |
| Gambardella et al. | 2019 | Fresh | bacteria | Gammaproteo bacteria | <i>Vibrio fischeri</i> | PE | Irregular | 9.03 | 0,5 | Bioluminescence | NOEC |
| Gambardella et al. | 2019 | Fresh | bacteria | Gammaproteo bacteria | <i>Vibrio fischeri</i> | PE | Irregular | 14.07 | 0,5 | Bioluminescence | NOEC |
| Gambardella et al. | 2019 | Fresh | bacteria | Gammaproteo bacteria | <i>Vibrio fischeri</i> | PE | Irregular | 24.46 | 0,5 | Bioluminescence | NOEC |
| Gambardella et al. | 2019 | Fresh | bacteria | Gammaproteo bacteria | <i>Vibrio fischeri</i> | PE | Irregular | 125 - 500 | 0,5 | Bioluminescence | NOEC |
| Gambardella et al. | 2019 | Fresh | bacteria | Gammaproteo bacteria | <i>Vibrio fischeri</i> | PE | Irregular | 10.14 | 0,5 | Bioluminescence | NOEC |

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|-----------------------------|------|--------|--------------|----------------------|----------------------------------|-----|-----------|-----------|------|---|---|
| <i>Gambardella et al.</i> | 2019 | Fresh | bacteria | Gammaproteo bacteria | <i>Vibrio fischeri</i> | PE | Irregular | 14.73 | 0,5 | Bioluminescence | NOEC |
| <i>Gambardella et al.</i> | 2019 | Fresh | bacteria | Gammaproteo bacteria | <i>Vibrio fischeri</i> | PE | Irregular | 62.14 | 0,5 | Bioluminescence | NOEC |
| <i>Gambardella et al.</i> | 2019 | Fresh | bacteria | Gammaproteo bacteria | <i>Vibrio fischeri</i> | PE | Irregular | 90.60 | 0,5 | Bioluminescence | NOEC |
| <i>Gambardella et al.</i> | 2019 | Fresh | algae | Bacillariophyceae | <i>Phaeodactylum tricornutum</i> | PE | Irregular | 4.25 | 72 | Growth | NOEC |
| <i>Gambardella et al.</i> | 2019 | Fresh | algae | Bacillariophyceae | <i>Phaeodactylum tricornutum</i> | PE | Irregular | 9.03 | 72 | Growth | NOEC |
| <i>Gambardella et al.</i> | 2019 | Fresh | algae | Bacillariophyceae | <i>Phaeodactylum tricornutum</i> | PE | Irregular | 14.07 | 72 | Growth | NOEC |
| <i>Gambardella et al.</i> | 2019 | Fresh | algae | Bacillariophyceae | <i>Phaeodactylum tricornutum</i> | PE | Irregular | 24.46 | 72 | Growth | NOEC |
| <i>Gambardella et al.</i> | 2019 | Fresh | algae | Bacillariophyceae | <i>Phaeodactylum tricornutum</i> | PE | Irregular | 125 - 500 | 72 | Growth | NOEC |
| <i>Gardon et al.</i> | 2018 | Marine | invertebrate | Bivalve | <i>Pinctada margaritifera</i> | PS | Sphere | 6, 10 | 720 | Feeding activity, Oxygen consumption, Assimilation efficiency, Scope for growth | Assimilation efficiency, Scope for growth |
| <i>Gardon et al.</i> | 2018 | Marine | invertebrate | Bivalve | <i>Pinctada margaritifera</i> | PS | Sphere | 6, 10 | 1440 | Growth, Reproductive effort, Regression of gametogenesis | Regression of gametogenesis |
| <i>Gerdes et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PET | Fragment | 5 | 96 | Immobilization | LC ₅₀ |
| <i>Goncalves et al.</i> | 2019 | Marine | invertebrate | Bivalve | <i>Mytilus galloprovincialis</i> | PS | Sphere | 2, 6, 10 | 48 | Oxidative stress | |
| <i>Goncalves et al.</i> | 2019 | Marine | invertebrate | Bivalve | <i>Mytilus galloprovincialis</i> | PS | Sphere | 5, 10 | 504 | Hystopathology | |
| <i>Gorokhova et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | N/A | Sphere | 1 - 5 | 12 | Swimming, Filtering behavior | Swimming, Filtering behavior |
| <i>Gray & Weinstein</i> | 2017 | Marine | invertebrate | Malacostraca | <i>Palaemonetes pugio</i> | PS | Sphere | 30 | 3 | Survival | No statistical comparison made with control |
| <i>Gray & Weinstein</i> | 2017 | Marine | invertebrate | Malacostraca | <i>Palaemonetes pugio</i> | PP | Fragment | 34 | 3 | Survival | No statistical comparison made with control |
| <i>Gray & Weinstein</i> | 2017 | Marine | invertebrate | Malacostraca | <i>Palaemonetes pugio</i> | PP | Fiber | 34 | 3 | Survival | No statistical comparison made with control |
| <i>Gray & Weinstein</i> | 2017 | Marine | invertebrate | Malacostraca | <i>Palaemonetes pugio</i> | PE | Sphere | 35 | 3 | Survival | No statistical comparison made with control |
| <i>Gray & Weinstein</i> | 2017 | Marine | invertebrate | Malacostraca | <i>Palaemonetes pugio</i> | PE | Sphere | 59 | 3 | Survival | No statistical comparison made with control |
| <i>Gray & Weinstein</i> | 2017 | Marine | invertebrate | Malacostraca | <i>Palaemonetes pugio</i> | PS | Sphere | 75 | 3 | Survival | No statistical comparison made with control |
| <i>Gray & Weinstein</i> | 2017 | Marine | invertebrate | Malacostraca | <i>Palaemonetes pugio</i> | PE | Sphere | 83 | 3 | Survival | No statistical comparison made with control |

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|------------------|------|--------|--------------|--------------|----------------------------|------|----------|------------|------|--|---|
| Gray & Weinstein | 2017 | Marine | invertebrate | Malacostraca | <i>Palaemonetes pugio</i> | PP | Fragment | 93 | 3 | Survival | No statistical comparison made with control |
| Gray & Weinstein | 2017 | Marine | invertebrate | Malacostraca | <i>Palaemonetes pugio</i> | PP | Fiber | 93 | 3 | Survival | No statistical comparison made with control |
| Gray & Weinstein | 2017 | Marine | invertebrate | Malacostraca | <i>Palaemonetes pugio</i> | PE | Sphere | 116 | 3 | Survival | No statistical comparison made with control |
| Gray & Weinstein | 2017 | Marine | invertebrate | Malacostraca | <i>Palaemonetes pugio</i> | PE | Sphere | 165 | 3 | Survival | No statistical comparison made with control |
| Green | 2016 | Marine | invertebrate | Bivalve | <i>Ostrea edulis</i> | HDPE | N/A | 0.48 - 316 | 1440 | Respiration, Filtration rate, Shell growth | |
| Green | 2016 | Marine | invertebrate | Bivalve | <i>Ostrea edulis</i> | PLA | N/A | 0.6 - 363 | 1440 | Respiration, Filtration rate, Shell growth | |
| Green | 2016 | Marine | invertebrate | Gastropoda | <i>Littorina sp.</i> | HDPE | N/A | 0.48 - 316 | 1440 | Abundance, Biomass | Abundance |
| Green | 2016 | Marine | invertebrate | Gastropoda | <i>Littorina sp.</i> | PLA | N/A | 0.6 - 363 | 1440 | Abundance, Biomass | Abundance |
| Green | 2016 | Marine | invertebrate | Malacostraca | <i>Idotea balthica</i> | HDPE | N/A | 0.48 - 316 | 1440 | Abundance, Biomass | Abundance |
| Green | 2016 | Marine | invertebrate | Malacostraca | <i>Idotea balthica</i> | PLA | N/A | 0.6 - 363 | 1440 | Abundance, Biomass | Abundance |
| Green | 2016 | Marine | invertebrate | Bivalve | <i>Scrobicularia plana</i> | HDPE | N/A | 0.48 - 316 | 1440 | Abundance, Biomass | Biomass |
| Green | 2016 | Marine | invertebrate | Bivalve | <i>Scrobicularia plana</i> | PLA | N/A | 0.6 - 363 | 1440 | Abundance, Biomass | Biomass |
| Green et al. | 2016 | Marine | invertebrate | Polychaeta | <i>Arenicola marina</i> | PLA | N/A | 1.4 - 707 | 744 | Growth, metabolic rate, cast production | Metabolic rate |
| Green et al. | 2016 | Marine | invertebrate | Polychaeta | <i>Arenicola marina</i> | HDPE | N/A | 2.5 - 316 | 744 | Growth, metabolic rate, cast production | Metabolic rate |
| Green et al. | 2016 | Marine | invertebrate | Polychaeta | <i>Arenicola marina</i> | PVC | N/A | 8.7 - 478 | 744 | Growth, metabolic rate, cast production | Metabolic rate, cast production |
| Green et al. | 2016 | Marine | algae | N/A | N/A | PLA | N/A | 1.4 - 707 | 744 | Biomass | Biomass |
| Green et al. | 2016 | Marine | algae | N/A | N/A | HDPE | N/A | 2.5 - 316 | 744 | Biomass | Biomass |
| Green et al. | 2016 | Marine | algae | N/A | N/A | PVC | N/A | 8.7 - 478 | 744 | Biomass | Biomass |
| Green et al. | 2016 | Marine | bacteria | N/A | N/A | PLA | N/A | 1.4 - 707 | 744 | Biomass, chlorophyll-a content | Chlorophyll-a content |
| Green et al. | 2016 | Marine | bacteria | N/A | N/A | HDPE | N/A | 2.5 - 316 | 744 | Biomass, chlorophyll-a content | Chlorophyll-a content |
| Green et al. | 2016 | Marine | bacteria | N/A | N/A | PVC | N/A | 8.7 - 478 | 744 | Biomass, chlorophyll-a content | Chlorophyll-a content |
| Green et al. | 2017 | Marine | invertebrate | Bivalve | <i>Ostrea edulis</i> | HDPE | N/A | 0.48 - 316 | 1200 | Filtration rate | Filtration rate |
| Green et al. | 2017 | Marine | invertebrate | Bivalve | <i>Mytilus edulis</i> | HDPE | N/A | 0.48 - 316 | 1200 | Filtration rate | Filtration rate |
| Green et al. | 2017 | Marine | invertebrate | Bivalve | <i>Ostrea edulis</i> | PLA | N/A | 0.6 - 363 | 1200 | Filtration rate | Filtration rate |
| Green et al. | 2017 | Marine | invertebrate | Bivalve | <i>Mytilus edulis</i> | PLA | N/A | 0.6 - 363 | 1200 | Filtration rate | Filtration rate |
| Green et al. | 2017 | Marine | algae | N/A | N/A | HDPE | N/A | 0.48 - 316 | 1200 | Biomass | |

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|-----------------------------|------|--------|--------------|----------------|---------------------------------|----------------------|-----------|-------------|------|--|---|
| <i>Green et al.</i> | 2017 | Marine | algae | N/A | <i>N/A</i> | PLA | N/A | 0.6 - 363 | 1200 | Biomass | |
| <i>Green et al.</i> | 2017 | Marine | bacteria | N/A | <i>N/A</i> | HDPE | N/A | 0.48 - 316 | 1200 | Biomass | Biomass |
| <i>Green et al.</i> | 2017 | Marine | bacteria | N/A | <i>N/A</i> | PLA | N/A | 0.6 - 363 | 1200 | Biomass | Biomass |
| <i>Green et al.</i> | 2017 | Marine | invertebrate | Polychaeta | <i>Eteone picta</i> | HDPE | N/A | 0.48 - 316 | 1200 | Abundance, Biomass | Abundance |
| <i>Green et al.</i> | 2017 | Marine | invertebrate | Polychaeta | <i>Eteone picta</i> | PLA | N/A | 0.6 - 363 | 1200 | Abundance, Biomass | Abundance |
| <i>Green et al.</i> | 2017 | Marine | invertebrate | Anopla | <i>Lineus longissimus</i> | HDPE | N/A | 0.48 - 316 | 1200 | Abundance, Biomass | Abundance |
| <i>Green et al.</i> | 2017 | Marine | invertebrate | Anopla | <i>Lineus longissimus</i> | PLA | N/A | 0.6 - 363 | 1200 | Abundance, Biomass | Abundance |
| <i>Green et al.</i> | 2019 | Marine | invertebrate | Bivalve | <i>Mytilus edulis</i> | PLA | Fragment | 0.6-363 | 1248 | Tenacity, Number of byssal threads, Haemolymph proteome | Tenacity, Number of byssal threads, Haemolymph proteome |
| <i>Green et al.</i> | 2019 | Marine | invertebrate | Bivalve | <i>Mytilus edulis</i> | HDPE | Fragment | 0.48-316 | 1248 | Tenacity, Number of byssal threads, Haemolymph proteome | Tenacity, Number of byssal threads, Haemolymph proteome |
| <i>Hämer et al.</i> | 2014 | Marine | invertebrate | Malacostraca | <i>Idotea emarginata</i> | PS | Sphere | 10 | 1008 | Survival, Growth, Intermolt duration, Ingestion rate | |
| <i>Hämer et al.</i> | 2014 | Marine | invertebrate | Malacostraca | <i>Idotea emarginata</i> | PS | Fragment | 1 - 100 | 1008 | Survival, growth, intermolt duration, ingestion rate | |
| <i>Hämer et al.</i> | 2014 | Marine | invertebrate | Malacostraca | <i>Idotea emarginata</i> | PA | Fiber | 20 - 2500 | 1008 | Survival, growth, intermolt duration, ingestion rate | |
| <i>Hankins et al.</i> | 2018 | Marine | invertebrate | Anthozoa | <i>Montastraea cavernosa</i> | PE | Sphere | 90 - 106 | 48 | Calcification | |
| <i>Hankins et al.</i> | 2018 | Marine | invertebrate | Anthozoa | <i>Orbicella faveolata</i> | PE | Sphere | 425 - 500 | 48 | Calcification | |
| <i>Hankins et al.</i> | 2018 | Marine | invertebrate | Anthozoa | <i>Montastraea cavernosa</i> | PE | Sphere | 850 - 1000 | 48 | Calcification | |
| <i>Hankins et al.</i> | 2018 | Marine | invertebrate | Anthozoa | <i>Orbicella faveolata</i> | PE | Sphere | 90 - 106 | 48 | Calcification | |
| <i>Hankins et al.</i> | 2018 | Marine | invertebrate | Anthozoa | <i>Montastraea cavernosa</i> | PE | Sphere | 425 - 500 | 48 | Calcification | |
| <i>Hankins et al.</i> | 2018 | Marine | invertebrate | Anthozoa | <i>Orbicella faveolata</i> | PE | Sphere | 850 - 1000 | 48 | Calcification | |
| <i>Imhof & Laforsch</i> | 2016 | Fresh | invertebrate | Gastropoda | <i>Potamopyrgus antipodarum</i> | PA, PET, PC, PS, PVC | Irregular | 4.64 - 602 | 1344 | Growth, reproduction | |
| <i>Imhof et al.</i> | 2017 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PA, PC, PET, PVC | Irregular | 27.5 - 72.5 | 48 | Survival, Growth, Reproduction, Gene expression | Growth |
| <i>Imhof et al.</i> | 2017 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | ABS, PVC, POM, SAN | Irregular | 22.3 - 48.5 | 48 | Survival, Growth, Reproduction, Gene expression | Growth |
| <i>Jabeen et al.</i> | 2018 | Fresh | fish | Actinopterygii | <i>Carassius auratus</i> | EVA | Fiber | 700 - 5000 | 1008 | Survival, Condition factor, Length, Weight, Hyspathology | Condition factor, Weight, Hyspathology |
| <i>Jabeen et al.</i> | 2018 | Fresh | fish | Actinopterygii | <i>Carassius auratus</i> | PS | Fragment | 2500 - 3000 | 1008 | Survival, Condition factor, Length, Weight, Hyspathology | Condition factor, Weight, Hyspathology |

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|------------------------|------|--------|--------------|----------------|------------------------------|-----------------------|-----------|-------------|------|--|------------------------|------------------|
| <i>Jabeen et al.</i> | 2018 | Fresh | fish | Actinopterygii | <i>Carassius auratus</i> | Polyethylene acrylate | Pellet | 4900 - 5000 | 1008 | Survival, Condition factor, Length, Weight, Hyspathology | Weight, Hystopathology | |
| <i>Jacob et al.</i> | 2019 | Marine | fish | Actinopterygii | <i>Acanthurus triostegus</i> | PS | Sphere | 91.26 | 192 | Survival, Foraging activity, Predation | | |
| <i>Jaikumar et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | N/A | Sphere | 1 - 5 | 48 | Survival | Survival | LC ₅₀ |
| <i>Jaikumar et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | N/A | Sphere | 1 - 5 | 96 | Survival | Survival | LC ₅₀ |
| <i>Jaikumar et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Irregular | 1 - 10 | 48 | Survival | Survival | LC ₅₀ |
| <i>Jaikumar et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Irregular | 1 - 10 | 96 | Survival | Survival | LC ₅₀ |
| <i>Jaikumar et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | N/A | Sphere | 1 - 5 | 48 | Survival | Survival | LC ₅₀ |
| <i>Jaikumar et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | N/A | Sphere | 1 - 5 | 96 | Survival | Survival | LC ₅₀ |
| <i>Jaikumar et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | PE | Irregular | 1 - 10 | 48 | Survival | Survival | LC ₅₀ |
| <i>Jaikumar et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | PE | Irregular | 1 - 10 | 96 | Survival | Survival | LC ₅₀ |
| <i>Jaikumar et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | N/A | Sphere | 1 - 5 | 48 | Survival | Survival | LC ₅₀ |
| <i>Jaikumar et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | N/A | Sphere | 1 - 5 | 96 | Survival | Survival | LC ₅₀ |
| <i>Jaikumar et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | PE | Irregular | 1 - 10 | 48 | Survival | Survival | LC ₅₀ |
| <i>Jaikumar et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | PE | irregular | 1 - 10 | 96 | Survival | Survival | LC ₅₀ |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | N/A | Sphere | 1 - 5 | 504 | Day of first brood | | NOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | N/A | Sphere | 1 - 5 | 504 | Day of first brood | | NOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | N/A | Sphere | 1 - 5 | 168 | Day of first brood | | NOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Irregular | 1 - 10 | 504 | Day of first brood | | NOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | PE | Irregular | 1 - 10 | 504 | Day of first brood | Day of first brood | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | PE | Irregular | 1 - 10 | 168 | Day of first brood | Day of first brood | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | N/A | Sphere | 1 - 5 | 504 | Size of first brood | Size of first brood | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | N/A | Sphere | 1 - 5 | 504 | Size of first brood | Size of first brood | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | N/A | Sphere | 1 - 5 | 168 | Size of first brood | Size of first brood | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Irregular | 1 - 10 | 504 | Size of first brood | Size of first brood | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | PE | Irregular | 1 - 10 | 504 | Size of first brood | | NOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | PE | Irregular | 1 - 10 | 168 | Size of first brood | Size of first brood | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | N/A | Sphere | 1 - 5 | 504 | Total # of broods | | NOEC |

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|-------------------------|------|--------|--------------|----------------|---------------------------|------------|-----------|--------|------|--|--------------------------|------|
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | N/A | Sphere | 1 - 5 | 504 | Total # of broods | Total # of broods | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | N/A | Sphere | 1 - 5 | 168 | Total # of broods | Total # of broods | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Irregular | 1 - 10 | 504 | Total # of broods | | NOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | PE | Irregular | 1 - 10 | 504 | Total # of broods | | NOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | PE | Irregular | 1 - 10 | 168 | Total # of broods | | NOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | N/A | Sphere | 1 - 5 | 504 | Size of first 3 broods | Size of first 3 broods | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | N/A | Sphere | 1 - 5 | 504 | Size of first 3 broods | Size of first 3 broods | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | N/A | Sphere | 1 - 5 | 168 | Size of first 3 broods | Size of first 3 broods | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Irregular | 1 - 10 | 504 | Size of first 3 broods | Size of first 3 broods | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | PE | Irregular | 1 - 10 | 504 | Size of first 3 broods | Size of first 3 broods | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | PE | Irregular | 1 - 10 | 168 | Size of first 3 broods | Size of first 3 broods | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | N/A | Sphere | 1 - 5 | 504 | Cumulative # of neonates | Cumulative # of neonates | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | N/A | Sphere | 1 - 5 | 504 | Cumulative # of neonates | Cumulative # of neonates | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | N/A | Sphere | 1 - 5 | 168 | Cumulative # of neonates | Cumulative # of neonates | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Irregular | 1 - 10 | 504 | Cumulative # of neonates | Cumulative # of neonates | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | PE | Irregular | 1 - 10 | 504 | Cumulative # of neonates | Cumulative # of neonates | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | PE | Irregular | 1 - 10 | 168 | Cumulative # of neonates | Cumulative # of neonates | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | N/A | Sphere | 1 - 5 | 504 | Terminal length | Terminal length | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | N/A | Sphere | 1 - 5 | 504 | Terminal length | | NOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | N/A | Sphere | 1 - 5 | 168 | Terminal length | Terminal length | LOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Irregular | 1 - 10 | 504 | Terminal length | | NOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Daphnia pulex</i> | PE | Irregular | 1 - 10 | 504 | Terminal length | | NOEC |
| <i>Jaikumar et al.</i> | 2019 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | PE | Irregular | 1 - 10 | 168 | Terminal length | | NOEC |
| <i>Jovanović et al.</i> | 2018 | Marine | fish | Actinopterygii | <i>Sparus aurata</i> | PVCHM W | Irregular | 75,6 | 1080 | Growth, Hypsathology, Levels of glucose, AST, ALT, LDH, & GGT in serum | | |
| <i>Jovanović et al.</i> | 2018 | Marine | fish | Actinopterygii | <i>Sparus aurata</i> | PA | Irregular | 111,7 | 1080 | Growth, Hypsathology, Levels of glucose, AST, ALT, LDH, & GGT in serum | | |
| <i>Jovanović et al.</i> | 2018 | Marine | fish | Actinopterygii | <i>Sparus aurata</i> | UHMWP E | Irregular | 23,4 | 1080 | Growth, Hypsathology, Levels of glucose, AST, ALT, LDH, & GGT in serum | | |

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| <i>Jovanović et al.</i> | 2018 | Marine | fish | Actinopterygii | <i>Sparus aurata</i> | PS | Irregular | 51 | 1080 | Growth, Hyspathology, Levels of glucose, AST, ALT, LDH, & GGT in serum | |
| <i>Jovanović et al.</i> | 2018 | Marine | fish | Actinopterygii | <i>Sparus aurata</i> | MDPE | Irregular | 54,5 | 1080 | Growth, Hyspathology, Levels of glucose, AST, ALT, LDH, & GGT in serum | |
| <i>Jovanović et al.</i> | 2018 | Marine | fish | Actinopterygii | <i>Sparus aurata</i> | PWCLM W | Irregular | 87,6 | 1080 | Growth, Hyspathology, Levels of glucose, AST, ALT, LDH, & GGT in serum | |
| <i>Jemec et al.</i> | 2016 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PET | Fiber | 60-1400 x 30-530 µm | 48 | Survival, Growth | Survival |
| <i>Jemec et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Fragment | 183.1 | 48 | Survival, Growth, Immobility | |
| <i>Jemec et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Fragment | 102.9 | 48 | Survival, Growth, Immobility | |
| <i>Jemec et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Fragment | 63.05 | 48 | Survival, Growth, Immobility | |
| <i>Jemec et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Fragment | 264 | 48 | Survival, Growth, Immobility | |
| <i>Jemec et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Fragment | 247.9 | 48 | Survival, Growth, Immobility | |
| <i>Jemec et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Fragment | 136.8 | 48 | Survival, Growth, Immobility | |
| <i>Jemec et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PET | Fiber | 22.8 | 48 | Survival, Growth, Immobility | |
| <i>Jemec et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Artemia franciscana</i> | PE | Fragment | 183.1 | 48 | Growth | Growth |
| <i>Jemec et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Artemia franciscana</i> | PE | Fragment | 102.9 | 48 | Growth | Growth |
| <i>Jemec et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Artemia franciscana</i> | PE | Fragment | 63.05 | 48 | Growth | Growth |
| <i>Jemec et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Artemia franciscana</i> | PE | Fragment | 264 | 48 | Growth | Growth |
| <i>Jemec et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Artemia franciscana</i> | PE | Fragment | 247.9 | 48 | Growth | Growth |
| <i>Jemec et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Artemia franciscana</i> | PE | Fragment | 136.8 | 48 | Growth | Growth |
| <i>Jemec et al.</i> | 2018 | Fresh | invertebrate | Branchiopoda | <i>Artemia franciscana</i> | PET | Fiber | 22.8 | 48 | Growth | Growth |
| <i>Jeong et al.</i> | 2016 | Marine | invertebrate | Monogononta | <i>Brachionus koreanus</i> | PS | Sphere | 6 | 288 | Survival, Growth, Reproduction | |
| <i>Jeong et al.</i> | 2016 | Marine | invertebrate | Monogononta | <i>Brachionus koreanus</i> | PS | Sphere | 6 | 24 | ROS production, enzyme activity | ROS production, enzyme activity |
| <i>Jeong et al.</i> | 2017 | Marine | invertebrate | Hexanauplia | <i>Paracyclopsina nana</i> | PS | Sphere | 6 | 24 | Growth rate, Fecundity, Development | Development |
| <i>Jin et al.</i> | 2018 | fresh | fish | Actinopterygii | <i>Danio rerio</i> | PS | Sphere | 50 | 336 | Microbiota dysbiosis, Inflammation | Microbiota dysbiosis |
| <i>Kalcikova et al.</i> | 2017 | Fresh | plant | Liliopsida | <i>Lemna minor</i> | PE | Irregular/s harp surface | 30 - 600 | 168 | Specific leaf growth rate, Photosynthetic pigments content, Root growth, Root cell viability | Root growth |
| <i>Kalcikova et al.</i> | 2017 | Fresh | plant | Liliopsida | <i>Lemna minor</i> | PE | Irregular/s mooth surface | 40 - 400 | 168 | Specific leaf growth rate, Photosynthetic pigments content, Root growth | Root growth |

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| <i>Kaposi et al.</i> | 2014 | Marine | invertebrate | Echinoidea | <i>Tripneustes gratilla larvae</i> | PE | Sphere | 10 - 45 | 120 | Survival, Growth | Growth |
| <i>Karami et al.</i> | 2017 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | LDPE | Fragments | 4.64 - 17.6 | 240 | Condition factor, Growth, Gene expression | |
| <i>Karami et al.</i> | 2017 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | LDPE | Fragments | 4.64 - 17.6 | 480 | Condition factor, Growth, Gene expression | Gene expression |
| <i>Korez et al.</i> | 2019 | Marine | invertebrate | Malacostraca | <i>Idotea emarginata</i> | PMMA | Fragment | 10 - 100 | 192 | Feeding rate, Enzymatic activity | Feeding rate, Enzymatic activity |
| <i>Kratina et al.</i> | 2019 | Fresh | invertebrate | Malacostraca | <i>Gammarus pulex</i> | PMMA | Sphere | 40.2 | 24 | Feeding rate, Metabolic rate | Metabolic rate |
| <i>Lee et al.</i> | 2013 | Estuarine | invertebrate | Hexanauplia | <i>Tigriopus japonicus F0 generation</i> | PS | Sphere | 6 | 336 | Survival, developmental time & fecundity | Fecundity EC ₅₀ |
| <i>Lee et al.</i> | 2013 | Estuarine | invertebrate | Hexanauplia | <i>Tigriopus japonicus F1 generation</i> | PS | Sphere | 6 | 336 | Survival, developmental time & fecundity | Fecundity EC ₅₀ |
| <i>Lei et al.</i> | 2018 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | PA | Irregular | 20 - 180 | 240 | Survival, Hypsathology | Hypsathology |
| <i>Lei et al.</i> | 2018 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | PE | Irregular | 20 - 120 | 240 | Survival, Hypsathology | Hypsathology |
| <i>Lei et al.</i> | 2018 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | PP | Irregular | 40 - 180 | 240 | Survival, Hypsathology | Survival, Hypsathology |
| <i>Lei et al.</i> | 2018 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | PVC | Irregular | 50 - 170 | 240 | Survival, Hypsathology | Hypsathology |
| <i>Lei et al.</i> | 2018 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | PS | Sphere | 5 | 240 | Survival, Hypsathology | |
| <i>Lei et al.</i> | 2018 | Fresh | invertebrate | Chromadorea | <i>Caenorhabditis elegans</i> | PA | Irregular | 20 - 180 | 48 | Survival, Body length, Reproduction, Calcium levels, Gene expression | Survival, Body length, Reproduction, Calcium levels, Gene expression |
| <i>Lei et al.</i> | 2018 | Fresh | invertebrate | Chromadorea | <i>Caenorhabditis elegans</i> | PE | Irregular | 20 - 120 | 48 | Survival, Body length, Reproduction, Calcium levels, Gene expression | Survival, Body length, Reproduction, Calcium levels, Gene expression |
| <i>Lei et al.</i> | 2018 | Fresh | invertebrate | Chromadorea | <i>Caenorhabditis elegans</i> | PP | Irregular | 40 - 180 | 48 | Survival, Body length, Reproduction, Calcium levels, Gene expression | Survival, Body length, Reproduction, Calcium levels, Gene expression |
| <i>Lei et al.</i> | 2018 | Fresh | invertebrate | Chromadorea | <i>Caenorhabditis elegans</i> | PVC | Irregular | 50 - 170 | 48 | Survival, Body length, Reproduction, Calcium levels, Gene expression | Survival, Body length, Reproduction, Calcium levels, Gene expression |
| <i>Lei et al.</i> | 2018 | Fresh | invertebrate | Chromadorea | <i>Caenorhabditis elegans</i> | PS | Sphere | 5 | 48 | Survival, Body length, Reproduction, Calcium levels, Gene expression | Survival, Body length, Reproduction, Gene expression |
| <i>LeMoine et al.</i> | 2018 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | PE | Sphere | 10 - 45 | 336 | Survival, Growth, Hatching, Oxygen consumption, Gene expression | Gene expression |
| <i>Leung & Chan</i> | 2018 | Marine | invertebrate | Polychaeta | <i>Perinereis aibuhitensis</i> | PS | Sphere | 8 - 12 | 672 | Survival, Reduction of posterior segment regeneration | Mortality, Reduction of posterior segment regeneration |
| <i>Leung & Chan</i> | 2018 | Marine | invertebrate | Polychaeta | <i>Perinereis aibuhitensis</i> | PS | Sphere | 32 - 38 | 672 | Survival, Reduction of posterior segment regeneration | Mortality, Reduction of posterior segment regeneration |
| <i>Lo & Chan</i> | 2018 | Marine | invertebrate | Gastropoda | <i>Crepidula onyx</i> | PS | Sphere | 2 - 5 | 336 | Survival, Growth, Feeding, Larval settlement | Feeding, Larval settlement |
| <i>Lu et al.</i> | 2016 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | PS | Sphere | 5 | 504 | Oxidative stress, inflammation & lipid accumulation | Oxidative stress, inflammation & lipid accumulation |

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|-----------------------------|------|--------|--------------|------------------|------------------------------|-----|-----------|-----------|------|--|--|
| Lu et al. | 2018 | Fresh | fish | Actinopterygii | <i>Oncorhynchus mykiss</i> | PS | Sphere | 1 | 2 | Gene expression | |
| Lu et al. | 2018 | Fresh | fish | Actinopterygii | <i>Oncorhynchus mykiss</i> | PS | Sphere | 20 | 2 | Gene expression | |
| Lu et al. | 2018 | Fresh | fish | Actinopterygii | <i>Oncorhynchus mykiss</i> | PS | Sphere | 40 | 2 | Gene expression | Gene expression |
| Lu et al. | 2018 | Fresh | fish | Actinopterygii | <i>Oncorhynchus mykiss</i> | PS | Sphere | 90 | 2 | Gene expression | Gene expression |
| Lu et al. | 2018 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | PS | Sphere | 1 | 2 | Gene expression | Gene expression |
| Lu et al. | 2018 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | PS | Sphere | 20 | 2 | Gene expression | |
| Lu et al. | 2018 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | PS | Sphere | 40 | 2 | Gene expression | |
| Lu et al. | 2018 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | PS | Sphere | 90 | 2 | Gene expression | Gene expression |
| Magni et al. | 2018 | Fresh | invertebrate | Bivalve | <i>Dreissena polymorpha</i> | PS | Sphere | 1, 10 | 144 | Cellular stress, Oxidative damage, Neuro-genotoxicity | Cellular stress |
| Magni et al. | 2019 | Fresh | invertebrate | Bivalve | <i>Dreissena polymorpha</i> | PS | Sphere | 1, 10 | 144 | Protein modulation | Protein modulation |
| Manlinich et al. | 2018 | Fresh | fish | Actinopterygii | <i>Pimephales promelas</i> | PE | Sphere | 180 - 212 | 720 | Growth | |
| Mao et al. | 2018 | Fresh | algae | Trebouxiophyceae | <i>Chlorella pyrenoidosa</i> | PS | Sphere | 1 | 720 | Growth, Photosynthesis, Morphology | Growth, Photosynthesis, Morphology |
| Mateos-Cárdenas | 2019 | Fresh | plant | Liliopsida | <i>Lemna minor</i> | PE | Sphere | 10 - 45 | 240 | Growth, Photosynthesis | |
| Mateos-Cárdenas | 2019 | Fresh | plant | Liliopsida | <i>Lemna minor</i> | PE | Sphere | 10 - 45 | 720 | Growth | |
| Mateos-Cárdenas | 2019 | Fresh | invertebrate | Malacostraca | <i>Gammarus duebeni</i> | PE | Sphere | 10 - 45 | 48 | Survival, Mobility | |
| Mazurais et al. | 2015 | Marine | fish | Actinopterygii | <i>Dicentrarchus labrax</i> | PE | Sphere | 10 - 45 | 1032 | Survival, Growth, Gene expression | Survival |
| Murphy & Quinn | 2018 | Fresh | invertebrate | Hydrozoa | <i>Hydra attenuata</i> | PE | Irregular | < 400 | 1 | Feeding rate | Feeding rate |
| Murphy & Quinn | 2018 | Fresh | invertebrate | Hydrozoa | <i>Hydra attenuata</i> | PE | Irregular | < 400 | 1 | Morphological score, Hydranth numbers | Morphological score, Hydranth numbers |
| Ogonowski et al. | 2016 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | N/A | Sphere | 1 - 5 | 504 | Number of produced offspring (NID) | NID |
| Ogonowski et al. | 2016 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Irregular | 2,6 | 504 | Number of produced offspring (NID) | NID |
| Ogonowski et al. | 2016 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | N/A | Sphere | 1 - 5 | 504 | Survival, Growth, Number of broods (BID) | |
| Ogonowski et al. | 2016 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Irregular | 2,6 | 504 | Survival, Growth, Number of broods (BID) | Survival, BID |
| Peixoto et al. | 2019 | Marine | invertebrate | Branchiopoda | <i>Artemia franciscana</i> | N/A | Sphere | 1 - 5 | 1056 | Survival, Growth, Feeding, Reproduction | Reproduction |
| Qiao et al. | 2019 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | PS | Sphere | 5 | 504 | Inflammation, oxidative stress, Metabolism, Gut microbiome | Inflammation, oxidative stress, Metabolism, Gut microbiome |
| Redondo-Hasselerharm et al. | 2018 | Fresh | invertebrate | Malacostraca | <i>Gammarus pulex</i> | PS | Irregular | 20 - 500 | 672 | Growth | Growth |

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|------------------------------------|------|-----------|--------------|----------------|-------------------------------|--------|-----------|-----------|-------|--|---|------------------|
| <i>Redondo-Hasselerharm et al.</i> | 2018 | Fresh | invertebrate | Malacostraca | <i>Gammarus pulex</i> | PS | Irregular | 20 - 500 | 672 | Survival, Feeding activity | | |
| <i>Redondo-Hasselerharm et al.</i> | 2018 | Fresh | invertebrate | Malacostraca | <i>Hyalella azteca</i> | PS | Irregular | 20 - 500 | 672 | Survival, Growth, Feeding activity | | |
| <i>Redondo-Hasselerharm et al.</i> | 2018 | Fresh | invertebrate | Clitellata | <i>Lumbriculus variegatus</i> | PS | Irregular | 20 - 500 | 672 | Reproduction | | |
| <i>Redondo-Hasselerharm et al.</i> | 2018 | Fresh | invertebrate | Clitellata | <i>Tubifex sp.</i> | PS | Irregular | 20 - 500 | 672 | Survival, Growth | | |
| <i>Redondo-Hasselerharm et al.</i> | 2018 | Fresh | invertebrate | Bivalve | <i>Sphaerium corneum</i> | PS | Irregular | 20 - 500 | 672 | Survival, Growth | | |
| <i>Redondo-Hasselerharm et al.</i> | 2018 | Fresh | invertebrate | Malacostraca | <i>Asellus aquaticus</i> | PS | Irregular | 20 - 500 | 672 | Survival, Growth | | |
| <i>Redondo-Hasselerharm et al.</i> | 2020 | Fresh | invertebrate | Clitellata | <i>Tubificidae</i> | PS | Fragment | 20 - 516 | 10800 | Population abundance | Population abundance | |
| <i>Rehse et al.</i> | 2016 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Sphere | 1 - 4 | 96 | Immobilization | Immobilization | EC ₅₀ |
| <i>Rehse et al.</i> | 2016 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PE | Sphere | 90 - 106 | 96 | Immobilization | Immobilization | |
| <i>Reichert et al.</i> | 2018 | Marine | invertebrate | Anthozoa | <i>Acropora humilis</i> | PE | Irregular | 37 - 163 | 672 | Tissue necrosis, Bleaching | Bleaching | |
| <i>Reichert et al.</i> | 2018 | Marine | invertebrate | Anthozoa | <i>Acropora millepora</i> | PE | Irregular | 37 - 163 | 672 | Tissue necrosis, Bleaching | Bleaching | |
| <i>Reichert et al.</i> | 2018 | Marine | invertebrate | Anthozoa | <i>Pocillopora verrucosa</i> | PE | Irregular | 37 - 163 | 672 | Tissue necrosis, Bleaching | Tissue necrosis | |
| <i>Reichert et al.</i> | 2018 | Marine | invertebrate | Anthozoa | <i>Pocillopora damicornis</i> | PE | Irregular | 37 - 163 | 672 | Tissue necrosis, Bleaching | Tissue necrosis | |
| <i>Reichert et al.</i> | 2018 | Marine | invertebrate | Anthozoa | <i>Porites lutea</i> | PE | Irregular | 37 - 163 | 672 | Tissue necrosis, Bleaching | | |
| <i>Reichert et al.</i> | 2018 | Marine | invertebrate | Anthozoa | <i>Porites cylindrica</i> | PE | Irregular | 37 - 163 | 672 | Tissue necrosis, Bleaching | Bleaching | |
| <i>Revel et al.</i> | 2018 | Estuarine | invertebrate | Polychaeta | <i>Hediste diversicolor</i> | PE, PP | N/A | 0.4 - 400 | 240 | Cell viability, Phagocytosis activity, Phenoloxdase, Acid phosphatase | Cell viability | |
| <i>Revel et al.</i> | 2019 | Marine | invertebrate | Bivalve | <i>Mytilus spp.</i> | PE, PP | N/A | 0.4 - 950 | 240 | Condition index, clearance rate, Oxydative stress, Enzyme activity, Genotoxicity | Oxydative stress, Genotoxicity | |
| <i>Revel et al.</i> | 2020 | Marine | invertebrate | Bivalve | <i>Crassostrea gigas</i> | PE, PP | Fragment | 0.4 - 500 | 240 | Clearance rate, Tissue alteration, Oxidative stress, Enzyme activity, Genotoxicity | | |
| <i>Ribeiro et al.</i> | 2017 | Marine | invertebrate | Bivalve | <i>Scrobicularia plana</i> | PS | Sphere | 20 | 336 | Condition index, Oxidative stress, Genotoxicity | Oxidative stress, Genotoxicity | |
| <i>Rist et al.</i> | 2017 | Fresh | invertebrate | Branchiopoda | <i>Daphnia magna</i> | PS | Sphere | 2 | 504 | Survival, Growth, time to first offspring, number of broods, number of neonates, total number of molts | | |
| <i>Romano et al.</i> | 2018 | Fresh | fish | Actinopterygii | <i>Barbodes gonionotus</i> | PVC | Fragment | 40 - 310 | 96 | Trypsin activity, Chymotrypsin activity, Hyspathology | Trypsin activity, Chymotrypsin activity | |

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|-------------------------------|------|-----------|--------------|----------------------|---------------------------------|------|-----------|------------|------|--|--|-------------------|
| <i>Seoane et al.</i> | 2019 | Marine | algae | Bacillariophyceae | <i>Chaetoceros neogracile</i> | PS | Sphere | 2 | 72 | Growth, Photosynthetic efficiency, Oxidative stress, Membrane potential, Enzyme activity, Lipid content | Enzyme activity, Lipid content | |
| <i>Silva et al.</i> | 2019 | Fresh | invertebrate | Insecta | <i>Chironomus riparius</i> | PE | Irregular | 32 - 63 | 240 | Growth | Growth | LOEC |
| <i>Silva et al.</i> | 2019 | Fresh | invertebrate | Insecta | <i>Chironomus riparius</i> | PE | Irregular | 63 - 250 | 240 | Growth | Growth | LOEC |
| <i>Silva et al.</i> | 2019 | Fresh | invertebrate | Insecta | <i>Chironomus riparius</i> | PE | Irregular | 125 - 500 | 240 | Growth | Growth | LOEC |
| <i>Silva et al.</i> | 2019 | Fresh | invertebrate | Insecta | <i>Chironomus riparius</i> | PE | Irregular | 32 - 63 | 672 | Emergence | Emergence | EmT ₅₀ |
| <i>Silva et al.</i> | 2019 | Fresh | invertebrate | Insecta | <i>Chironomus riparius</i> | PE | Irregular | 63 - 250 | 672 | Emergence | Emergence | EmT ₅₀ |
| <i>Silva et al.</i> | 2019 | Fresh | invertebrate | Insecta | <i>Chironomus riparius</i> | PE | Irregular | 125 - 500 | 672 | Emergence | Emergence | EmT ₅₀ |
| <i>Sjollema et al.</i> | 2016 | Marine | algae | Chlorophyceae | <i>Dunaliella tertiolecta</i> | PS | Sphere | 6 | 72 | Growth, Photosynthesis | | |
| <i>Straub et al.</i> | 2017 | Fresh | invertebrate | Malacostraca | <i>Gammarus fossarum</i> | PMMA | Irregular | 32-63 | 672 | Feeding rate, WW change, Assimilation efficiency | WW change, Assimilation efficiency | |
| <i>Straub et al.</i> | 2017 | Fresh | invertebrate | Malacostraca | <i>Gammarus fossarum</i> | PHB | Irregular | 32-63 | 672 | Feeding rate, WW change, Assimilation efficiency | WW change | |
| <i>Sun et al.</i> | 2018 | Marine | bacteria | Gamma proteobacteria | <i>Halomonas alkaliphila</i> | PS | Sphere | 1.6 | 2 | Cell viability, Oxidative stress, Cell membrane composition, Conversion efficiencies for inorganic N | Cell viability, Oxidative stress, Cell membrane composition, Conversion efficiencies for inorganic N | |
| <i>Sussarellu et al.</i> | 2016 | Marine | invertebrate | Bivalve | <i>Crassostrea gigas</i> | PS | Sphere | 2, 6 | 1440 | Feeding, Fecundity, Offspring development | Feeding, Fecundity, Offspring development | |
| <i>Tang et al.</i> | 2018 | Marine | invertebrate | Anthozoa | <i>Pocillopora damicornis</i> | PS | Smooth | 1 | 24 | Symbiotic density, Chlorophyll content, Enzyme activity | Chlorophyll content, Enzyme activity | |
| <i>Van Cauwenbergh et al.</i> | 2015 | Marine | invertebrate | Bivalve | <i>Mytilus edulis</i> | PS | Sphere | 10, 30, 90 | 336 | Energy budget | | |
| <i>Van Cauwenbergh et al.</i> | 2015 | Marine | invertebrate | Polychaeta | <i>Arenicola marina</i> | PS | Sphere | 10, 30, 90 | 336 | Energy budget | | |
| <i>Von Moos et al.</i> | 2012 | Marine | invertebrate | Bivalve | <i>Mytilus edulis</i> | HDPE | Sphere | 0 - 80 | 96 | Condition index, Granulocytoma formation, Lysosomal membrane stability, Lipofuscin accumulation, Neutral lipid content | Granulocytoma formation, Lysosomal membrane stability | |
| <i>Vroom et al.</i> | 2017 | Marine | invertebrate | Hexanauplia | <i>Calanus finmarchicus</i> | PS | Sphere | 15 | 264 | Survival | | |
| <i>Wan et al.</i> | 2019 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | PS | Sphere | 5 | 168 | Gut microbiome, Metabolism, Oxidative stress, Gene transcription | Gut microbiome, Metabolism, Oxidative stress, Gene transcription | |
| <i>Wan et al.</i> | 2019 | Fresh | fish | Actinopterygii | <i>Danio rerio</i> | PS | Sphere | 50 | 168 | Gut microbiome, Metabolism, Oxidative stress, Gene transcription | Gut microbiome, Metabolism, Oxidative stress, Gene transcription | |
| <i>Wang et al.</i> | 2019 | Estuarine | invertebrate | Branchiopoda | <i>Artemia parthenogenetica</i> | PS | Sphere | 10 - 11 | 24 | Survival | | NOEC |

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|----------------|------|-----------|--------------|------------------|---|------|-----------|-------------|------|--|--|
| Wang et al. | 2019 | Estuarine | invertebrate | Branchiopoda | <i>Artemia parthenogenetica</i> | PS | Sphere | 10 - 11 | 336 | Growth, Instar development time | NOEC |
| Wang et al. | 2019 | Estuarine | invertebrate | Branchiopoda | <i>Artemia parthenogenetica</i> | PS | Sphere | 10 - 11 | 24 | Epithelial cell morphology | Epithelial cell morphology |
| Watts et al. | 2016 | Marine | invertebrate | Hexanauplia | <i>Carcinus maenas</i> | PS | Sphere | 8 | 1 | Oxygen consumption | Oxygen consumption |
| Watts et al. | 2016 | Marine | invertebrate | Hexanauplia | <i>Carcinus maenas</i> | PS | Sphere | 8 | 24 | Oxygen consumption, Plasma Na+ & Ca2+ ion concentrations | Plasma Na+ & Ca2+ ion concentrations |
| Weber et al. | 2018 | Fresh | invertebrate | Malacostraca | <i>Gammarus pulex</i> | PET | Fragment | 10 - 150 | 1152 | Survival, molting, metabolism, feeding activity | |
| Welden & Cowie | 2016 | Marine | invertebrate | Malacostraca | <i>Nephrops norvegicus</i> | PP | Fiber | 3000 - 5000 | 5760 | Growth, Feeding rate, Metabolic depression | Growth |
| Wen et al. | 2018 | Fresh | fish | Actinopterygii | <i>Symphysodon aequifasciatus</i> | PE | Sphere | 70 - 88 | 720 | Survival, Growth, Predatory performance, Enzyme activity | Predatory performance, Enzyme activity |
| Wright et al. | 2013 | Marine | invertebrate | Polychaeta | <i>Arenicola marina</i> | UPVC | Irregular | 130 | 672 | Feeding activity, Phagocytic activity, Energy reserves | Feeding activity, Phagocytic activity, Energy reserves |
| Wright et al. | 2013 | Marine | invertebrate | Polychaeta | <i>Arenicola marina</i> | UPVC | Irregular | 130 | 48 | Gut residence time | Gut residence time |
| Wu et al. | 2019 | Fresh | algae | Trebouxiophyceae | <i>Chlorella pyrenoidosa</i> | PP | N/A | 157 | 264 | Chlorophyll a content, Photosynthetic efficiency | Chlorophyll a content, Photosynthetic efficiency |
| Wu et al. | 2019 | Fresh | bacteria | Cyanophyceae | <i>Microcystis flos-aqua</i> | PP | N/A | 157 | 168 | Chlorophyll a content, Photosynthetic efficiency | Chlorophyll a content, Photosynthetic efficiency |
| Wu et al. | 2019 | Fresh | algae | Trebouxiophyceae | <i>Chlorella pyrenoidosa</i> | PVC | N/A | 172 | 264 | Chlorophyll a content, Photosynthetic efficiency | Chlorophyll a content, Photosynthetic efficiency |
| Wu et al. | 2019 | Fresh | bacteria | Cyanophyceae | <i>Microcystis flos-aqua</i> | PVC | N/A | 172 | 168 | Chlorophyll a content, Photosynthetic efficiency | Chlorophyll a content, Photosynthetic efficiency |
| Xu et al. | 2017 | Marine | invertebrate | Bivalve | <i>Atactodea striata</i> | PS | Granule | 63 - 250 | 240 | Clearance rate, Absorption efficiency, Respiration rate | Clearance rate |
| Yin et al. | 2018 | Marine | fish | Actinopterygii | <i>Sebastes schlegelii</i> | PS | Sphere | 15 | 336 | Feeding activity, Growth, Swimming speed, Hyspathology | Feeding activity, Growth, Swimming speed, Hyspathology |
| Yin et al. | 2019 | Marine | fish | Actinopterygii | <i>Sebastes schlegelii</i> | PS | Sphere | 15 | 336 | Survival, Growth, Condition factor, Behavior, Oxygen consumption, Ammonia excretion, Nutritional composition | Behavior, Oxygen consumption, Ammonia excretion, Nutritional composition |
| Yu et al. | 2018 | Estuarine | invertebrate | Malacostraca | <i>Eriocheir sinensis</i> | PS | Sphere | 5 | 504 | Survival, Growth, Oxydative stress, Inflammation | Growth, Oxydative stress, Inflammation |
| Zhang et al. | 2017 | Marine | algae | Mediophyceae | <i>Skeletonema costatum</i> | PVC | Sphere | 1 | 96 | Growth, Photosynthesis | Growth, Photosynthesis |
| Zhang et al. | 2019 | Fresh | invertebrate | Hexanauplia | <i>Tigriopus japonicus F0, F1 generations</i> | PS | Sphere | 5,84 | 48 | Survival, sex ratio, developmental time of nauplius phase, Developmental time to maturation, Number of clutches, Number of nauplii/clutch, Fecundity, Proteomic analysis | Survival, number of nauplii/clutch, Fecundity, Proteomic analysis |
| Zhang et al. | 2019 | Fresh | invertebrate | Hexanauplia | <i>Tigriopus japonicus F2 generation</i> | PS | Sphere | 5,84 | 48 | Survival, sex ratio, developmental time of nauplius phase, Developmental time to maturation, Number of clutches, | |

| | | | | | | | | | | Number of nauplii/clutch, Fecundity, Proteomic analysis | | |
|--------------------------|------|--------|--------------|--------------|---------------------------|-----------|--------|-----------|-----|--|--|------------------|
| <i>Zhao et al.</i> | 2019 | Marine | algae | Dinophyceae | <i>Karenia mikimotoi</i> | PVC | Sphere | 1 | 96 | Growth, Chlorophyll content, Photosynthetic efficiency | Growth, Chlorophyll content, Photosynthetic efficiency | |
| <i>Ziajahromi et al.</i> | 2018 | Fresh | invertebrate | Insecta | <i>Chironomus tepperi</i> | PE | Sphere | 1 - 4 | 120 | Survival, Growth, Emergence | Survival, Growth, Emergence | |
| <i>Ziajahromi et al.</i> | 2018 | Fresh | invertebrate | Insecta | <i>Chironomus tepperi</i> | PE | Sphere | 10 - 27 | 120 | Survival, Growth, Emergence | Survival, Growth, Emergence | |
| <i>Ziajahromi et al.</i> | 2018 | Fresh | invertebrate | Insecta | <i>Chironomus tepperi</i> | PE | Sphere | 43 - 54 | 240 | Survival, Growth, Emergence | Survival, Growth, Emergence | |
| <i>Ziajahromi et al.</i> | 2018 | Fresh | invertebrate | Insecta | <i>Chironomus tepperi</i> | PE | Sphere | 100 - 126 | 240 | Survival, Growth, Emergence | Emergence | |
| <i>Ziajahromi et al.</i> | 2017 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | PE | Sphere | 1 - 4 | 48 | Survival | Survival | LC ₅₀ |
| <i>Ziajahromi et al.</i> | 2017 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | Polyester | Fiber | 0 - 1000 | 48 | Survival | Survival | LC ₅₀ |
| <i>Ziajahromi et al.</i> | 2017 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | PE | Sphere | 1 - 4 | 192 | Survival | Survival | |
| <i>Ziajahromi et al.</i> | 2017 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | Polyester | Fiber | 0 - 1000 | 192 | Survival | Survival | |
| <i>Ziajahromi et al.</i> | 2017 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | PE | Sphere | 1 - 4 | 192 | Growth, Reproduction | Reproduction | EC ₅₀ |
| <i>Ziajahromi et al.</i> | 2017 | Fresh | invertebrate | Branchiopoda | <i>Ceriodaphnia dubia</i> | Polyester | Fiber | 0 - 1000 | 192 | Growth, Reproduction | Reproduction | EC ₅₀ |

Table S2. Explanation of the quantitative scoring system proposed to evaluate the studies testing the effects of MP on aquatic biota using the (QA/QC) criteria. The purpose of the quantitative scoring system criteria is to assess the quality of the papers and to give guidance for appropriate methods for MP particle studies in the future. The first subset of criteria (criteria 1 – 12) relates to the general technical quality of effect tests. The second set of criteria (criteria 13-20) relates to relevance of the papers to be used in risk assessment. For each criterion a score of either 2 (adequate), 1 (adequate with restrictions) or 0 (inadequate) points were assigned, which are explained below.

| CRITERIA RELATING TO THE TECHNICAL QUALITY OF EFFECT TESTS (1 – 12) | | | | |
|--|--|--|--|---|
| Criterion | Explanation | Score 2 | Score 1 | Score 0 |
| Particle characterization | | | | |
| 1. Particle size | Size is a crucial factor explaining effects of MP and thus should be reported. | - If a range of sizes is used; a full (i.e. ≥ 10 bins) size distribution is measured and reported. - If a single size is used, that size is measured with an indication of measurement error and reported. | If particle size/sizes are reported but not measured. | No information on size reported. |
| 2. Particle shape | Shape is a crucial factor explaining effects of MP and thus should be measured and reported. | Shapes are measured with high resolution picture and reported. | Particle shapes are reported but not measured. | No information on particle shape is reported. |
| 3. Polymer type | Polymer type can be a factor explaining effects of MP and thus should be reported. | Polymer identity confirmed with e.g. FTIR, Raman spectroscopy or similar methods. | Polymer type provided with certificate or as provided by manufacturer. | No information on polymer identity is reported. |

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|----------------------------------|---|---|--|---------------------------------------|
| 4. Source of MP | Specification on where MP stock or solution is bought and/or how it is self-made maximizes reproducibility and thus should be reported. | The origin and/or production of MP in own laboratory is reported in detail. | The information given on MP source is incomplete and hence not fully reproducible. | No information on MP source reported. |
| 5. Data reporting | Unambiguous units are required to ensure reproducibility of the experiment and to make it possible to compare data across experiments. | MP concentrations are reported as mass as well as number concentration. | MP concentrations are reported as mass or as number concentration. | MP concentrations are not reported. |
| Experimental design | | | | |
| 6. Chemical purity | In order to test particle toxicity, the toxicity of other chemicals in solution or mixture should be ruled out. This includes additives present in MP, chemicals associated with food particles and surfactants (e.g. Tween). | Chemical effects other than from the polymer or solution/mixtures are ruled out. MP are cleaned with organic solvent. | - Chemicals are analyzed or studies relied on manufacturer certificate. - Controls are used or calculations are made with values from literature (i.e., LC ₅₀ or EC ₅₀) to rule out toxicity of chemical impurities. | Not mentioned. |
| 7. Laboratory preparation | MP contamination arising from the laboratory (air, water and materials) should be minimized. All materials used (equipment, tools, work surfaces and clothing) should be free of MP. | - All materials used are thoroughly washed with high quality water (e.g. Milli-Q water). - Measures are taken to prevent MP contamination from air. - Cotton lab coats were used to avoid microfiber contamination. | Only part of the measures under 2 are taken to avoid MP contamination. | Not mentioned. |

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|--|---|---|--|---|
| 8. Verification of background contamination | Assessment of MP contamination of the exposure systems in the laboratory. | Level of contamination evaluated and quantified, e.g. with FTIR, Raman or similar method. | Contamination visually inspected. | No verification of contamination. |
| 9. Verification of exposure | Not only the nominal concentration should be mentioned. The exposure concentration should be measured. | Measurement of exposure concentration and evidence that at least 80% of the nominal concentration throughout the test is maintained. | Measurement of exposure concentration, however no evidence that at least 80% of the nominal concentration throughout the test is maintained. | No verification of exposure concentration. |
| 10. Homogeneity of exposure | Verification of homogeneity throughout the entire exposure system is crucial for the MP characterization and the assessment of bioavailability. | Water as medium: Picture or measurement of MP in water that demonstrated well mixed or dispersion in solution. Sediment as medium: Description of method used to obtain homogenous exposure. | Water as medium: Description of the method used to obtain homogeneous exposure. Sediment as medium: | Not mentioned or exposure is not homogenous. |
| 11. Exposure assessment | Exposure of the organism to MP should be verified by measurement. | Exposure of the organism to MP is measured quantitatively with e.g. FTIR or Raman. In case MP are ingested additionally an digestion step is included (see Hermsen et al., 2018). ⁵¹ | Exposure of the organism to MP is demonstrated qualitatively, visually, in a separate experiment or without digestion step. | No measurement of exposure of MP to organism. |
| 12. Replication | For statistical rigor in detecting effect thresholds (e.g., EC ₅₀ or EC ₁₀), sufficient replicates should be tested. | 3 or more replicates. | 2 replicates. | No replicates. |

CRITERIA APPLICABLE TO ECOLOGICAL RISK ASSESSMENT (13-20)

| Criterion | Explanation | Score 2 | Score 1 | Score 0 |
|--|--|--|--|---|
| Applicable to Risk assessment | | | | |
| 13. Endpoints | Endpoints should be considered that inform ecologically relevant population level risk assessment and clearly reported. | Endpoints taken at the community (e.g. bacteria and algae) or individual level (e.g. survival, mortality, growth, development, reproduction). | Suborganismal responses as long as a causal relationship with the endpoints mentioned under '2' has been demonstrated. | Endpoints cannot be unambiguously linked to a threat on the population or individual level. |
| 14. Presence of natural (food) particles | The exposure conditions should be environmentally relevant. | Natural particles (at least food) are added to avoid force feeding of MP. Criterion not applicable to algae or bacteria and hence these studies receive 2 points. | Food is not optimally available. | No food or natural particles are added to avoid force feeding of MP. |
| 15. Reporting of effect thresholds | To enable PEC/PNEC types of comparisons, the effect threshold should be assessed with error of uncertainty using dose- response relationships. | Effect thresholds are reported as L(E)Cx with error or uncertainty intervals. | Effect thresholds are reported as LOEC or NOEC, or as L(E)Cx value without error or confidence interval. | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC or not possible to derive effect threshold. |
| 16. Quality of dose-response relationship | For statistical rigor in detecting effect thresholds (e.g., EC ₅₀ , EC ₁₀), sufficient doses should be tested, including a treatment control, covering the full shape of the effect curve and emphasizing the slope for parameter estimation. | Multiple doses, at least 6, including a treatment control. | Multiple doses, at least 5, including a treatment control. | Less than 5 doses. |

| Ecological relevance | | | | |
|---------------------------------------|--|---|---|---|
| 17. Concentration range tested | Concentrations should be motivated (with a reference in the appropriate unit) from measured environmental concentrations (MEC). | More than 1 environmentally relevant concentration was used within the range tested. | At least 1 environmentally relevant concentration was used within the range tested. | - No relevant concentrations were used. - No comparison with MEC. |
| 18. Aging and biofouling | Aging, weathering and biofouling is what occurs in the environment and could affect the uptake of MP; therefore, it is crucial to consider this for an ecological relevant experiment. | MP particles have undergone process to make them environmentally realistic by accounting for biofouling. In addition, pictures of altered particles are provided. | MP particles have undergone process to make them environmentally realistic, accounting for aging, weathering and/or biofouling, however they have not been characterized. | Pristine MP used and/or conditions were as such that it was not possible to form a biofilm during the exposure time. |
| 19. Diversity of MP tested | In the environment, MP have a wide variety of shapes and sizes. This needs to be taken into account for environmentally relevant effect assessment. | A wide range of sizes (order of magnitude), shapes and densities are used, thereby approaching the diversity of environmental MP. | Diversity relates to only 1 or 2 of the characteristics (e.g. only a wide size range) and/or spans a part of the characteristics range only. | Only a single type of particles is tested (i.e. single size, shape and density). |
| 20. Exposure time | It is crucial to use appropriate exposure times to allow for the detection of adverse effects. | Bacteria and phytoplankton \geq 1 week Zooplankton \geq 21 d Benthic invertebrates \geq 28 d Fish \geq 3 months Macrophytes \geq 28 d | Bacteria, phytoplankton: 1 – 7 d Zooplankton: 4 – 21 d Benthic invertebrates: 7 - 28 d Fish: 1 - 3 months Macrophytes: 7 - 28 d | Bacteria, phytoplankton < 1 d Zooplankton < 4 d Benthic invertebrates < 7 d Fish < 1 month Macrophytes: < 7 d |

Table S3. Scores and mechanisms per author. For each criterion a score of either 2 (adequate), 1 (adequate with restrictions) or 0 (inadequate) points is assigned to the study under consideration. Maximum possible score is 40 points. Texts on the demonstrated mechanism often are literal quotes from the respective articles. These quotes are identified by quotation marks (“x”).

Paper: Aljaibachi and Callaghan (2018)⁹³

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Average size given (2 µm) but not measured | 1 |
| 2 | Particle shape | Shape mentioned (bead) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Sigma-Aldrich) | 2 |
| 5 | Data reporting | Data reported in mg/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Authors state that there was no evidence for aggregation, however, was not demonstrated | 1 |
| 11 | Exposure assessment of organisms | Ingestion quantified in separate experiment | 1 |
| 12 | Replication | 5 replicates used | 2 |
| <u>Subtotal</u> | | | <u>10</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, growth and reproduction | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Detection of effect thresholds (LT ₁₀ , LT ₅₀ and LT ₉₀) with standard error | 2 |
| 16 | Quality of dose-response relationship | 6 concentrations including control | 2 |
| 17 | Concentration range tested | No comparison made with MEC | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 21 days | 2 |
| <u>Total</u> | | | <u>21</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effect | Where ample food is present, MPs have little effect on adults. | |

Paper: Au et al. (2015)⁸

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Min/max size values (10 - 27 µm for spheres & 75-20 x 20 µm for fibers) given and confirmed with microscope. No PSD or average with error given | 1 |
| 2 | Particle shape | Shapes given (sphere and fiber), but no pictures shown | 1 |
| 3 | Polymer type | Polymer type given and checked for PP fibers, not for PE spheres | 2 |
| 4 | Source MP | Source given for spheres (Cospheric); fibers are self-made | 2 |
| 5 | Data reporting | Data reported in particles/ml | 1 |
| 6 | Chemical purity | Spheres were not cleaned, fibers cleaned with water | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Spheres in ethanol quantified twice using a hemocytometer prior to experiment, fibers counted individually | 1 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified in the same experiment | 1 |
| 12 | Replication | At least 3 replicates | 2 |
| Subtotal | | | 11 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, growth, reproduction and egestion time | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | LC ₅₀ for survival. For the rest, effect thresholds are reported without the use of L(E)Cx or NOEC value | 2 |
| 16 | Quality of dose-response relationship | 6 concentrations including control for survival, 4 for other endpoints | 2 |
| 17 | Concentration range tested | They do not include environmentally realistic concentrations for spheres (5000 MP/ml is the lowest conc.) | 0 |
| 18 | Aging and biofouling | Fibers were aged (no pictures), while spheres were pristine MP | 1 |
| 19 | Diversity of MP tested | 2 polymer types separately tested, 2 shapes separately tested, a small range of sizes | 0 |
| 20 | Exposure time | Up to 42 days | 2 |
| Total | | | 22 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Reduced growth and reproduction | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “MP ingestion <i>may cause</i> a mechanical hazard , resulting in blocking the food passage (1) or <i>may cause</i> the organism to feel satiated , indirectly resulting in a reduction in food intake (2) . Increased toxicity due to PP MP fiber exposures <i>may be</i> attributable to the difference in shape.” “It is possible that some of the differences seen between the acute exposures to PE and PP MP <i>could be</i> due to the oxygen-containing functional groups (3) present on the surface of PP MP fibers due to the various aging processes the marine rope endured.” | |

Paper: Blarer and Burkhardt-Holm (2016)¹¹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Average size given (1.6 µm for spheres & 500 x 20 µm for fibers) but not confirmed | 1 |
| 2 | Particle shape | Shapes given (bead and fiber), no pictures shown | 1 |
| 3 | Polymer type | Polymer types given (PS for spheres, PA for fibers) but not confirmed | 1 |
| 4 | Source MP | Particles are made at Nuertingen-Geislingen University. Information given is incomplete and not fully reproducible. | 1 |
| 5 | Data reporting | Data reported in particles/cm for fibers, particles/ml for spheres | 1 |
| 6 | Chemical purity | It is stated that both MP were produced without additives | 1 |
| 7 | Lab preparation | Only mentioned for digestion of samples (after exposure experiment) | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Quantified at the start of the experiment | 1 |
| 10 | Homogeneity of exposure | Fibers were not homogeneously distributed; however, spheres were measured throughout water column. Concentrations were stable through time (SI). | 2 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified for fibers and qualitatively assessed for spheres | 1 |
| 12 | Replication | 15 replicates | 2 |
| <u>Subtotal</u> | | | <u>12</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Feeding rate, assimilation efficiency, wet weight change | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | They do not include environmentally relevant concentrations | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 28 days | 2 |
| <u>Total</u> | | | <u>19</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effect | “Microplastics affect assimilation efficiency in the freshwater amphipod <i>Gammarus fossarum</i> ” | |
| * | Demonstrated mechanism | “The presence of PA fibers in the digestive tract inhibited food assimilation.” | |
| * | Speculated mechanism | “The long PA fibers <i>may physically damage the digestive tract (1)</i> and thereby <i>impair assimilation efficiency (2)</i> .” The sharp-edged fibers, however, <i>could</i> have caused more pronounced mechanical injuries of the delicate gut epithelium than the round and smooth-surfaced beads, but different surface properties (3) <i>could</i> have contributed as well.” | |

Paper: Bour et al. (2018)⁷⁵

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Min/max size values given (4-6 µm; 20-25 µm and 125-500 µm) and confirmed with microscope only for the biggest size class | 2 |
| 2 | Particle shape | Shape mentioned (Irregular fragment) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PE) but not confirmed | 1 |
| 4 | Source MP | Source given (Micro Powders) | 2 |
| 5 | Data reporting | Data reported in mg/kg and particles/kg of sediment | 2 |
| 6 | Chemical purity | Analysis of three pollutants in MP. Not all chemical effects can be ruled out. | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | MP visually quantified in sediment only at the end of the experiment | 1 |
| 10 | Homogeneity of exposure | Homogenization mentioned and checked at the end of the experiment | 2 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified in the same experiment | 1 |
| 12 | Replication | 4 replicates | 2 |
| Subtotal | | | 15 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, Condition index, burrowing behavior and surborganismal responses | 2 |
| 14 | Presence of natural (food) particles | Use of sediment for benthic invertebrate, so natural particles present | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 4 concentrations including control | 0 |
| 17 | Concentration range tested | Environmentally relevant concentrations used only for bigger particles | 1 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, a variety of sizes separately tested | 0 |
| 20 | Exposure time | 28 days | 2 |
| Total | | | 23 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Decreased energy reserve in <i>E. tenuis</i> . Decrease protein content in <i>A. nitida</i> | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “When passing through the digestive tracts, MP <i>could</i> physically damage gut tissues (1) , resulting in decreased nutrient uptake (2) . Physical damage <i>can</i> lead to inflammatory responses. We <i>postulate</i> that long-term exposure of benthic bivalves would lead to depletion of energy reserves (2) and potentially result in effects at the organism level.” | |

Paper: Bosker et al. (2019)¹⁰⁷

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Min/max and average sizes given (1- 5, 4.1 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (sphere) but not confirmed | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | Data provided in particles/ml | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Concentration verified with a hemocytometer prior to exposure | 1 |
| 10 | Homogeneity of exposure | Vortexing prior to exposure, daily resuspension manually (pipette), plus aeration | 1 |
| 11 | Exposure of assessment of organisms | Not assessed | 0 |
| 12 | Replication | 4 replicates | 2 |
| <u>Subtotal</u> | | | <u>10</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Population size and population biomass | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 5 concentrations including control | 1 |
| 17 | Concentration range tested | High concentrations of MP, no comparison with MEC | 0 |
| 18 | Aging and biofouling | Exposure time of experiment is long enough for biofilm to develop | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 21 days | 1 |
| <u>Total</u> | | | <u>17</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Reduction in total biomass population | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “ First, the accumulation of MP in the gut <i>might</i> reduce the uptake efficiency of the food or reduce assimilation of food (2) . After uptake MP can form aggregates in the gut of organisms, and as a result can cause a blockage in the gut (1) which could reduce food uptake.” “ changes in the energy translocation to cope with elimination of the MP (9) .” | |

Paper: Browne et al. (2008)⁹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Average size given (3 and 9.6 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape mentioned (sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Molecular probes) | 2 |
| 5 | Data reporting | Number of particles/treatment (no concentration per ml or L given) | 0 |
| 6 | Chemical purity | No leaching observed of fluorescent chemical label. Not all chemical effects can be ruled out | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Verified by Coulter Counter analysis at the start of the experiment | 1 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified in the same experiment | 1 |
| 12 | Replication | At least 3 replicates | 2 |
| <u>Subtotal</u> | | | <u>10</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Feeding activity and suborganismal responses | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | Environmentally relevant concentration not included | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, 2 sizes separately tested | 0 |
| 20 | Exposure time | 48 days | 2 |
| <u>Total</u> | | | <u>17</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects found | |

Paper: Bruck and Ford (2018)¹¹⁷

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Average size given (8 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Thermo Scientific) | 2 |
| 5 | Data reporting | Only in MP/g of food. No info on how many grams of food were added. | 0 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified | 1 |
| 12 | Replication | 3 replicates | 2 |
| <u>Subtotal</u> | | | <u>8</u> |
| <i>Applicable for risk assessment</i> | | | |
| 13 | Endpoints | Feeding, growth and moulting | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 4 concentrations including control | 0 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 35 days | 2 |
| <u>Total</u> | | | <u>15</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects found. | |

Paper: Canniff and Hoang (2018)¹¹⁰

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Min/max size values given (63-75 µm) but distribution not shown | 1 |
| 2 | Particle shape | Shape given (bead) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) and confirmed with FTIR | 2 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | Data reported in mg/L and particles/L | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified | 1 |
| 12 | Replication | 4 replicates | 2 |
| <u>Subtotal</u> | | | <u>11</u> |
| <i>Applicable for risk assessment</i> | | | |
| 13 | Endpoints | Survival and reproduction | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 4 concentrations including control | 0 |
| 17 | Concentration range tested | No comparison MEC | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, small size range | 0 |
| 20 | Exposure time | 21 days | 2 |
| <u>Total</u> | | | <u>18</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects found. | |

Paper: Capolupo et al. (2018)¹¹⁸

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Average size given (3 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Polyscience) | 2 |
| 5 | Data reporting | Data reported in MP/mL and µg/L | 2 |
| 6 | Chemical purity | Stated that there are no surfactants added or present in MP suspension | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Lack of aggregation verified with epifluorescence microscopy at the start of the experiment | 2 |
| 11 | Exposure assessment of organisms | MP visually quantified | 1 |
| 12 | Replication | 4 replicates | 2 |
| Subtotal | | | 13 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Feeding efficiency, Morphological/development and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 7 concentrations including control | 2 |
| 17 | Concentration range tested | MP concentrations far exceed those detected in marine environment | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 48-h Embryo-larval development | 0 |
| Total | | | 19 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects found on consumption or development. However, MP induced physical and transcriptional impairments in mussel larvae, indicating sub-lethal impacts. | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “Transcriptional data obtained in this study outline the <i>potential</i> impacts of MP on shell biogenesis, immune and lysosomal systems (8). ” | |

Paper: Chae, Kim and An (2019)¹¹⁹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Particle size distribution measured with microscope and CellSens software. Min/max size values (180-212 µm) and average size with standard deviation provided (203.84 ± 13.76 µm) | 2 |
| 2 | Particle shape | Shape given (sphere) and confirmed with microscopic images | 2 |
| 3 | Polymer type | Polymer type given (PE) but not confirmed | 1 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | In mg/L and particles/L | 2 |
| 6 | Chemical purity | Chemicals from MP were analyzed qualitatively using liquid chromatography | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Verification of exposure at beginning of experiment | 1 |
| 10 | Homogeneity of exposure | MP floated on the surface of the solution and the algae were exposed to MP at the surface of the medium only | 0 |
| 11 | Exposure of assessment | Not addressed | 0 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 13 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Cell growth, photosynthetic activity and cell morphology | 2 |
| 14 | Presence of natural (food) particles | NA | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly. | 0 |
| 16 | Quality of dose-response relationship | 8 concentrations including control | 2 |
| 17 | Concentration range tested | Authors state that concentration range tested includes environmentally relevant concentrations, however no reference is provided | 0 |
| 18 | Aging and biofouling | Not mentioned | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, small size range | 0 |
| 20 | Exposure time | 6 days | 1 |
| Total | | | 20 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Cell growth and photosynthetic activity enhanced. Cell morphology was not affected. | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “This phenomenon <i>might</i> be explained by trace concentrations of additive chemicals (4) endocrine disruptors, phthalates, stabilizers) that possibly leached from MP promoting the growth and photosynthetic activity of <i>D. salina</i> .” | |

Paper: Chapron et al. (2018)⁸⁴

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Average size given (500 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (bead) but not shown | 1 |
| 3 | Polymer type | Polymer type given (LDPE) but not confirmed | 1 |
| 4 | Source MP | Source given (Santa Cruz Biotechnology) | 2 |
| 5 | Data reporting | Data reported in beads/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Not addressed | 0 |
| 12 | Replication | No replicates | 0 |
| | | <u>Subtotal</u> | <u>6</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Growth, feeding and behavior | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | No comparison MEC | 0 |
| 18 | Aging and biofouling | MP were incubated for 2 months in filtered seawater | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 69 days | 2 |
| | | <u>Total</u> | <u>13</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effect on behavior or feeding. Reduced growth/calcification | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “MP did not impact polyp behavior or prey capture rates; however, calcification was still reduced compared to control and in situ conditions. The exact causes are <i>still unclear</i> , but they <i>might</i> involve possible physical damages (5) or energy storage alteration (2) .” “ Thus, the fitness of the organism <i>could</i> be affected by an increased energy cost caused by the egestion of plastics. ” | |

Paper: Choi et al. (2018)¹²⁰

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | PSD measured but only max/min values given without error (150-180 µm for spheres, 6-350 µm for irregular MP) | 1 |
| 2 | Particle shape | Shapes given (sphere, irregular) and shown | 2 |
| 3 | Polymer type | Polymer type given (PE) but not confirmed | 1 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | Data reported in mg/L and particles/L | 2 |
| 6 | Chemical purity | Control used for surfactant Tween-80 | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Tween-80 solution added to obtain dispersibility, not verified | 1 |
| 11 | Exposure assessment of organisms | Ingestion qualitatively assessed | 1 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 13 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, malformations, swimming behavior, and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | No addition of food | 0 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations including control | 0 |
| 17 | Concentration range tested | Only unrealistic concentrations used | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | 2 polymers separately tested, various shapes and wide size ranges for irregular MP (more than 1 order of magnitude) | 1 |
| 20 | Exposure time | 4 days | 0 |
| Total | | | 16 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Decreased swimming behavior, ROS generated, ROS-related molecular changes | |
| * | Demonstrated mechanism | “The <u>mechanisms</u> underlying the responses to MP are <u>still unclear</u> .” | |
| * | Speculated mechanism | “The generation of intracellular ROS , followed by the differential regulation of genes related to ROS, <i>suggest</i> that ROS-related toxic effects (10) occur in organisms exposed to MP.” | |

Paper: Cole and Galloway (2015)³⁰

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Average size values given (1 and 10 µm) but not confirmed for effect experiment (only for uptake experiment) | 2 |
| 2 | Particle shape | Shape given (bead) but not confirmed for effect experiment (only for uptake experiment) | 2 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 2 |
| 4 | Source MP | Source given (Spherotech) | 2 |
| 5 | Data reporting | Data reported in particles/l and ug/ml | 2 |
| 6 | Chemical purity | Preservatives removed by centrifuging, removing the supernatant, washing in Milli-Q and centrifuging again | 2 |
| 7 | Lab preparation | Bioassays were covered to limit airborne contamination | 1 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Quantified at the start of the experiment using a coulter counter | 1 |
| 10 | Homogeneity of exposure | MP beads added to seawater and sonicated. Postexposure subsamples were viewed under microscope and aggregation observed | 0 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified in separate experiment | 1 |
| 12 | Replication | At least 4 replicates | 2 |
| <u>Subtotal</u> | | | <u>17</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Feeding and growth | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control for growth; 5 concentration including control for feeding | 1 |
| 17 | Concentration range tested | They do not include environmentally realistic concentrations | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, 2 sizes separately tested | 0 |
| 20 | Exposure time | 8 days | 1 |
| <u>Total</u> | | | <u>23</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects found | |

Paper: Cole et al. (2013)⁴

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Average sizes given (7.3 and 20.6 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape given (bead) but not confirmed | 1 |
| 3 | Polymer type | Polymer type given (PS) and confirmed with Coherent anti-Stokes Raman scattering (CARS) | 2 |
| 4 | Source MP | Source given (Spherotech) | 2 |
| 5 | Data reporting | Data reported in particles/ml | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Ingestion quantified however in separate experiment. Found MP trapped between external appendages. | 1 |
| 12 | Replication | At least 5 replicates | 2 |
| Subtotal | | | 10 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Feeding | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 5 concentrations including control | 1 |
| 17 | Concentration range tested | They do not include environmentally realistic concentrations | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, 2 sizes separately tested | 0 |
| 20 | Exposure time | 24 h | 0 |
| Total | | | 15 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Reduced feeding | |
| * | Demonstrated mechanism | Reduced algal ingestion rate | |
| * | Speculated mechanism | “ During our studies, we also found MP were becoming trapped between the external appendages and carapace segments of live copepods. We found that very small MP (0.4–3.8 µm) became lodged between the filamental hairs and setae of the antennules, furca, and the swimming legs. As these appendages have key roles in copepod function and behavior, this <i>may</i> have repercussions for locomotion, ingestion, mating, and mechanoreception, that may limit their ability to detect prey, feed, reproduce, and evade predators.” | |

Paper: Cole et al. (2015)⁹⁸

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average size given (20 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape mentioned (sphere) but not confirmed | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Sigma-Aldrich) | 2 |
| 5 | Data reporting | Data reported in particles/ml and % of food | 1 |
| 6 | Chemical purity | Toxicity ruled out using references | 1 |
| 7 | Lab preparation | Chambers covered with loosely fitting lids to prevent airborne contamination | 1 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Quantified at the start of the experiment using a coulter counter | 1 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | In the ingestion rate experiment MP are quantitatively assessed. In the long exposure MP uptake verified by visually checking fecal pellets. | 1 |
| 12 | Replication | At least 5 replicates | 2 |
| Subtotal | | | 12 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, Reproductive output, Egg production rates, Respiration rate | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | They say the concentration is not extreme as used in recent ecotoxicological papers, however, do not compare to MEC | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | Up to 9 days | 1 |
| Total | | | 17 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Significant reduction in egg volume on days 7 and 9 resulted from reduced ingested carbon biomass (owing to MP exposure) of the adult copepods. | |
| * | Demonstrated mechanism | “ Here we demonstrate that ingestion of MP can significantly alter the feeding capacity of the pelagic copepod <i>Calanus helgolandicus</i> .” “ The budget helps identify that MP exposed copepods will have much higher energetic deficiencies (2) than controls, predominantly owing to the 40% reduction in ingested carbon biomass. ” | |
| * | Speculated mechanism | - | |

Paper: Cole et al. (2019)⁴⁴

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average size given (10-30 µm for granules, 30 µm for fibers) and pictures shown. No indication of measurement error. | 1 |
| 2 | Particle shape | Shape given and shown in pictures SI | 2 |
| 3 | Polymer type | Polymer type given (nylon) but not confirmed | 1 |
| 4 | Source MP | Source given (Goodfellow) | 2 |
| 5 | Data reporting | Data reported in particles/ml | 1 |
| 6 | Chemical purity | MP were rinsed with ethanol and water prior to their use. Additionally, chemicals were analyzed. | 2 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Quantified using a Sedgewick rafter chamber prior to the experiment | 1 |
| 10 | Homogeneity of exposure | Use of a rotating plankton wheel, no verification of homogeneity | 1 |
| 11 | Exposure of assessment of organisms | Ingestion quantified in separate experiment | 1 |
| 12 | Replication | 10 replicates | 2 |
| Subtotal | | | 14 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Feeding, Prosome length, Moulting, Lipid accumulation | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | Higher than relevant | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 6 days | 1 |
| Total | | | 19 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Shift in prey selectivity and premature moulting, reduction in ingestion rate for <i>T. rotula</i> and <i>S. trochoidea</i> | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “When consumed, fibers are more prone to causing physical damage owing to their sharp edges (1).” Limited assimilation efficiency (2), additives (4) ” Reduced feeding (2) and stymied lipid accumulation <i>may</i> both have contributed to earlier moulting; however endocrine disruption <i>might</i> also have played a role.” | |

Paper: Cong et al. (2019)¹²¹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Min/max sizes given (10 - 11 µm) and measured with coulter counter, data not shown. No indication of measurement error | 1 |
| 2 | Particle shape | Shape given (sphere) but not checked | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Name of provider given, Thermo Fisher Scientific corporation, USA | 2 |
| 5 | Data reporting | Data reported in particles/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Verification of concentration in sock solution | 1 |
| 10 | Homogeneity of exposure | Not assessed | 0 |
| 11 | Exposure of assessment of organisms | Ingestion quantitatively assessed in separate experiment | 0 |
| 12 | Replication | 3 replicates | 2 |
| | | <u>Subtotal</u> | <u>9</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, Growth, Reproduction | 2 |
| 14 | Presence of natural (food) particles | Feeding included in one of the treatments | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | Higher than natural levels | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 14 days | 2 |
| | | <u>Total</u> | <u>15</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Increased mortality and decreased growth and reproduction | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | - | |

Paper: de Sá, Luís and Guilhermino, (2015)⁸⁸

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|-----------------|
| 1 | Particle size | Min/max size values given (420 - 500 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape given (sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PE) but not confirmed | 1 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | Data reported in particles/treatment (no concentration per ml or L given) | 0 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | The relative percentage of MP ingested was calculated relatively to the total number of preys ingested | 1 |
| 12 | Replication | No replication | 0 |
| | | <u>Subtotal</u> | <u>6</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Predatory performance and efficiency | 0 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | They do not include environmentally realistic concentrations/no comparison to MEC | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, small range of sizes | 0 |
| 20 | Exposure time | 3 minutes | 0 |
| | | <u>Total</u> | <u>8</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Reduction of the predatory performance and efficiency | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | - | |

Paper: Critchell and Hoogenboom (2018)⁹⁴

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Min/max size values given (1-2 mm) but not measured | 1 |
| 2 | Particle shape | Shapes given (fragment, sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PET) but not confirmed | 1 |
| 4 | Source MP | Source given (Visy Plastics) | 2 |
| 5 | Data reporting | Data reported in mg/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified | 1 |
| 12 | Replication | 6 replicates | 2 |
| <u>Subtotal</u> | | | <u>9</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Growth, body condition and behavior | 2 |
| 14 | Presence of natural (food) particles | Addition of food in one experiment, in other experiment food is replaced by MP | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 5 concentrations including control | 1 |
| 17 | Concentration range tested | Reported that highest concentration is higher than MEC, authors did not mention if other concentrations were environmentally relevant | 0 |
| 18 | Aging and biofouling | MP were soaked for at least 2 weeks, however not characterized | 1 |
| 19 | Diversity of MP tested | One polymer type, various shapes for fragments, small size range (less than one order of magnitude) | 1 |
| 20 | Exposure time | 1 week acute exposure and 6 weeks chronic exposure | 1 |
| <u>Total</u> | | | <u>17</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | When food is replaced by MP it negatively affects growth and body condition due to limited food availability | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | - | |

Paper: Détrée and Gallardo-Escárate (2017)⁷⁸

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Min/max size values given (1 - 50 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (bead) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PE) but not confirmed | 1 |
| 4 | Source MP | Source given (Abifor) | 2 |
| 5 | Data reporting | Data reported in microbeads/L | 1 |
| 6 | Chemical purity | Report that MP is free of any additive | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Aeration systems added to optimize the distribution and suspension of MP | 1 |
| 11 | Exposure assessment of organisms | Not addressed | 0 |
| 12 | Replication | 2 replicates | 1 |
| <u>Subtotal</u> | | | <u>9</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Suborganismal, no causal link with endpoints taken higher | 0 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, small size range | 0 |
| 20 | Exposure time | 24 hours | 0 |
| <u>Total</u> | | | <u>11</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “Up-regulation of genes relative to carbon metabolism, oxidative stress, immune response and apoptosis in the mantle and digestive gland, down-regulation of genes involved in carbon metabolism in the hemolymph and gills.” | |
| * | Demonstrated mechanism | “Key enzymes of metabolism <i>are</i> impacted.” “ Disruptive and tissue dependent effect of a short MP exposure on major biological processes (8) in <i>M. galloprovincialis</i> .” | |
| * | Speculated mechanism | “Ingestion of MP <i>may</i> provoke adverse effects that require an augmentation of energy production to maintain the organism’s homeostasis .” “MP <i>may</i> modulate the mussel's oxidative system (10) .” “The damage provoked by the ingestion of MP and the inflammation that follows, <i>may</i> require the production of stress, immune and antioxidant proteins to maintain the global homeostasis of the organism and demand increased production and utilization of energy.” | |

Paper: Espinosa, Cuesta and Esteban (2017)¹²²

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Min/max size values given (40 - 150 µm) and confirmed with microscope | 2 |
| 2 | Particle shape | Shape not given | 0 |
| 3 | Polymer type | Polymer type given (PVC) but not confirmed | 1 |
| 4 | Source MP | Source given (Centro Tecnológico del Calzado y del Plástico), but not commercial and not reproducible | 1 |
| 5 | Data reporting | Data reported in mg/kg of feed | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Not assessed | 0 |
| 12 | Replication | No replicates | 0 |
| | | Subtotal | 5 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Growth and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations tested including control | 0 |
| 17 | Concentration range tested | Includes one environmentally relevant concentration | 1 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, small range of sizes | 0 |
| 20 | Exposure time | Up to 30 days | 1 |
| | | Total | 12 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “Our data <u>suggest low damage</u> to muscle, liver and kidney, but not to gut, as a result of PVC-MP intake.” “ Our results indicated that PVC-MP intake did not have a significant impact on the HK leucocyte cellular activities (phagocytosis, respiratory burst activity and peroxidase content) or on the humoral response (peroxidase activity and IgM levels) in the serum or skin mucus.” | |
| * | Speculated mechanism | “These findings <i>could indicate</i> that MP ingestion caused cellular stress (6) in the liver .” | |
| * | Demonstrated mechanism | - | |

Paper: Franzellitti et al. (2019)⁷⁷

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Average sizes reported (3 and 45 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) but not measured | 1 |
| 4 | Source MP | Source given (Polyscience) | 2 |
| 5 | Data reporting | Data reported in particles/mL | 1 |
| 6 | Chemical purity | Authors state that no surfactants were present or added, however did not assure this by cleaning | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Before spiking, lack of aggregation was verified by epifluorescence microscopy | 2 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified | 1 |
| 12 | Replication | At least 3 replicates | 2 |
| <u>Subtotal</u> | | | <u>12</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Only suborganismal endpoints used, cannot be unambiguously linked to a threat on the population level. | 0 |
| 14 | Presence of natural (food) particles | No addition of food | 0 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations including control | 0 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | Not mentioned | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, 2 sizes separately tested | 0 |
| 20 | Exposure time | 4 days | 0 |
| <u>Total</u> | | | <u>12</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “Reduction of MXR activity and down-regulation of ABCB and ABCC transcripts” | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “Data reported in this study <i>suggest</i> that modulation of the MXR system may be a part of a generalized response triggered by particle ingestion and stimulation of digestive and immune functions both in larval and adult stages of mussels.” | |

Paper: Gambardella et al. (2019)⁷¹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | PSD measured by electronic counter. D10, D50 (2.92, 8.27, 13.79, 23.65, 9.04, 14.64, 59.22, 93.43 µm) and D90 provided with error | 2 |
| 2 | Particle shape | Shapes mentioned (irregular) but not measured | 0 |
| 3 | Polymer type | Polymer type (PE) given but not measured | 1 |
| 4 | Source MP | Source given (Micro Powders, Cospheric, Rotogal) | 2 |
| 5 | Data reporting | Data reported in mg/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Actual exposure checked but only in separate experiment | 1 |
| 10 | Homogeneity of exposure | Shaker, rotating wheel, air point mentioned | 1 |
| 11 | Exposure of assessment of organisms | For non-ingesting organisms (algae), assessment of external exposure suffices | 2 |
| 12 | Replication | 3 replicates | 2 |
| <u>Subtotal</u> | | | <u>12</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Algal growth and bacterial bioluminescence | 2 |
| 14 | Presence of natural (food) particles | NA for algae and bacteria | 2 |
| 15 | Detection of effect thresholds | Effect thresholds provided (although higher than highest dose) | 2 |
| 16 | Quality of dose-response relationship | Number of concentrations not mentioned, but a wide range was used | 0 |
| 17 | Concentration range tested | Range of doses spanned environmentally relevant concentrations | 2 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, various shapes, wide range of sizes used, however separately tested | 0 |
| 20 | Exposure time | 30 minutes for bacteria, 72 h for algal growth | 1 |
| <u>Total</u> | | | <u>21</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects found | |

Paper: Gardon et al. (2018)¹²³

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Average sizes given (6 and 10 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (bead) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) and confirmed with Raman | 2 |
| 4 | Source MP | Source given (Polyscience) | 2 |
| 5 | Data reporting | Data reported in µg/L and in particles/L | 2 |
| 6 | Chemical purity | Tween-20 added to control tank | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Tween-20 added to avoid agglutination | 1 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified | 1 |
| 12 | Replication | 4 replicates | 2 |
| Subtotal | | | 13 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Growth, Reproductive effort, Regression of gametogenesis, Feeding activity, Oxygen consumption, Assimilation efficiency, Scope for growth | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 4 concentrations including control | 0 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, small size range | 0 |
| 20 | Exposure time | 2 months | 2 |
| Total | | | 20 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | MP have negative impact on Assimilation Efficiency (AE) and mean energy balance (SFG). In exposed oyster's gametogenesis was strongly impacted | |
| * | Demonstrated mechanism | “This study <i>highlights</i> the impact of ingesting a diet containing polystyrene microbeads on the assimilation efficiency (2) of the pearl oyster which directly influences its energy balance . The dose-dependent decrease in AE and SFG supports these results and <i>demonstrates</i> an immutable effect of micro-PS on the oyster physiology .” | |
| * | Speculated mechanism | “Thus, we <u>cannot distinguish</u> between the possible mechanisms explaining the toxicity of MP-PS, to which direct particle toxicity and effects of MP-PS associated chemicals (4) such as DVB, may contribute.” | |

Paper: Gerdes et al. (2019)³³

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | PSD given and confirmed with DLS (5 µm) | 2 |
| 2 | Particle shape | Shape not mentioned, but picture provided | 2 |
| 3 | Polymer type | Polymer type (PET) given and confirmed with FTIR | 2 |
| 4 | Source MP | Source given (Goodfellow) for original material. Pellets were milled to powder. Information is sufficient for the MP to be reproducible. | 2 |
| 5 | Data reporting | Data reported in mg/L and particles/L (Table S1) | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Use of a plankton wheel. No sedimentation observed. Continuous particle ingestion by the test animals was confirmed for both materials through observation of the exposed animals by bright-field microscopy | 2 |
| 11 | Exposure of assessment of organisms | Ingestion qualitatively by bright field microscopy | 1 |
| 12 | Replication | At least four replicates | 2 |
| <u>Subtotal</u> | | | <u>15</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Immobilization | 2 |
| 14 | Presence of natural (food) particles | Authors state that our animals were starved during the exposure. Natural particles added | 1 |
| 15 | Detection of effect thresholds | LC ₅₀ with 95% confidence intervals | 2 |
| 16 | Quality of dose-response relationship | 6 concentrations including control | 2 |
| 17 | Concentration range tested | Much higher than environmentally relevant | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one size | 0 |
| 20 | Exposure time | 96 h | 1 |
| <u>Total</u> | | | <u>23</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | MP PET is more toxic than kaolin | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “Energy expenditure may increase (2) in the presence of non-food particles since induced filtering activity is similar as for food particles.” | |

Paper: Gonçalves et al.⁷⁶

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average sizes given (2, 5, 6 and 10 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape mentioned (sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) given but not confirmed | 1 |
| 4 | Source MP | Sources given (Alfa-Aesar, Sigma-aldrich, Magsphere) | 2 |
| 5 | Data reporting | Data reported in particles/ml | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Checked in separate experiment with a coulter counter | 1 |
| 10 | Homogeneity of exposure | Dispersion verified with coulter counter in separate experiment | 1 |
| 11 | Exposure of assessment of organisms | Ingestion qualitatively assessed in separate experiment | 1 |
| 12 | Replication | At least 3 replicates | 2 |
| Subtotal | | | 11 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Suborganismal endpoints (oxidative stress response and histopathological effects) | 0 |
| 14 | Presence of natural (food) particles | Food added only for the 21 d histopathology exposure | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations including control | 0 |
| 17 | Concentration range tested | Authors state that concentrations are high, yet realistic. Comparison with MEC made. | 2 |
| 18 | Aging and biofouling | Exposure time long enough to allow for biofouling | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, small size range | 0 |
| 20 | Exposure time | Oxidative stress for 48 hours, histopathology for 21 days | 1 |
| Total | | | 17 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “Biochemical indicators for oxidative stress were generally irresponsive regardless of organ and time of exposure. Small foci of haemocytic infiltration in gastric epithelia were found, albeit not clearly related to MP ingestion. Globally, no evident histopathological damage was recorded in whole-body sections of exposed animals.” | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | - | |

Paper: Gorokhova et al. (2018)^{124*}

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Min/max size values given (1 - 5 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (sphere) but not shown | 1 |
| 3 | Polymer type | Not reported | 0 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | No concentration unit provided, only number of particles and volume water in aquaria mentioned separately | 0 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Verification of the contamination at the start of the experiment | 1 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Not addressed | 0 |
| 12 | Replication | 2 Replicates | 1 |
| <u>Subtotal</u> | | | <u>6</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Swimming and filtering behavior | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations including control | 0 |
| 17 | Concentration range tested | Only unrealistic concentrations used | 0 |
| 18 | Aging and biofouling | Particles coated with artificial biofilm; surface charge measured | 2 |
| 19 | Diversity of MP tested | One polymer type, one shape, small size range | 0 |
| 20 | Exposure time | 12 hours | 0 |
| <u>Total</u> | | | <u>12</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Increased swimming activity | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “The observed differences <i>imply</i> that daphnids <i>would spend more time performing the non-feeding movements when swimming in suspension with MP (2)</i> , and even more when the MP are carrying biofilm. The increased jumping activity of filtrators exposed to microparticles (both natural and anthropogenic) <i>may</i> translate into changes in energy balance and growth.” | |

* ‘by exception this non-peer reviewed study was included’

Paper: Gray and Weinstein (2017)¹⁰

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average sizes given (30, 34, 34, 35, 59, 75, 83, 93, 93, 116, 165 µm) but not measured | 1 |
| 2 | Particle shape | Shapes given (spheres, fragments, fibers) but not shown | 1 |
| 3 | Polymer type | Polymer types given (PE, PS, PP) and confirmed in previous study for the polypropylene rope | 2 |
| 4 | Source MP | Sources given (Cospheric and Phosphorex, TWO H Chem and Clemson Univ.) | 2 |
| 5 | Data reporting | Data reported in particles/L | 1 |
| 6 | Chemical purity | Other toxic substances are ruled out | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Only nominal concentration mentioned | 0 |
| 10 | Homogeneity of exposure | Particles were first suspended in ethanol. Gentle aeration for fragments and fibers | 1 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified in gut and respiratory chambers | 1 |
| 12 | Replication | No replicates | 0 |
| Subtotal | | | 10 |
| <i>Applicable for risk assessment</i> | | | |
| <i>Criterion</i> | <i>Explanation</i> | | |
| 13 | Endpoints | Survival | 2 |
| 14 | Presence of natural (food) particles | Absence of food | 0 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | Authors state they used an environmentally relevant concentration | 1 |
| 18 | Aging and biofouling | Use of weathered marine rope, all other used MP are pristine | 1 |
| 19 | Diversity of MP tested | 3 polymer types, 3 shapes, size ranges 30-165 µm used. However, they were separately tested | 0 |
| 20 | Exposure time | 3 hours | 0 |
| Total | | | 14 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Increased mortality | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “Retained fibers <i>may</i> become entangled within the intestinal tract over time, leading to a nonbiodegradable gut blockage (1) , resulting in death. It is also <i>possible</i> that toxicity related to fiber ingestion is the result of internal structures becoming damaged as the entangled fibers pass through the gut.” | |
| * | Speculated mechanism | “In any event, prolonged resident times of spheres in the gut can result in decreased food consumption because of false satiation (2) , leading to reduced fitness.” | |

Paper: Green (2016)¹⁰⁰

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Mean and Min/max size values (0.6 - 363 µm for PLA, 0.48 - 316 µm for HDPE) and confirmed with a Mastersizer | 2 |
| 2 | Particle shape | Shape not mentioned | 0 |
| 3 | Polymer type | Polymer types given (PLA and HDPE) but not confirmed | 1 |
| 4 | Source MP | Not given | 0 |
| 5 | Data reporting | Data reported in ug/L and particles/L | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Measured concentrations at the start of the experiment (SI) | 1 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Not mentioned | 0 |
| 12 | Replication | 6 replicates | 2 |
| <u>Subtotal</u> | | | <u>8</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Respiration, filtration, and growth rates of one species, diversity, abundance and biomass of the community | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations used, including control | 0 |
| 17 | Concentration range tested | All concentrations are environmentally realistic, although the higher one is rare | 2 |
| 18 | Aging and biofouling | Added to microalgae for 3 days, additionally the exposure time was long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | 2 polymer types separately tested, wide range of sizes (more than 1 order or magnitude) | 1 |
| 20 | Exposure time | 60 days | 2 |
| <u>Total</u> | | | <u>18</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Reduction in abundances and biomasses of organisms | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “When MP are ingested, effects on organisms <i>can</i> arise due to physical (blockages (1)) chemical (4) (plasticizers or persistent organic pollutants) ; Rochman <i>et al.</i> , 2013) or biological (microbial communities (7)) on the particles) factors. Reductions in abundances and biomasses of organisms found in the current study <i>may</i> have occurred directly due to ingestion of MP (causing mortality, reduced reproductive output or reduced feeding), or indirectly, due to MP altering the behavior or abundances of other organisms (thereby altering interactions between species).” | |

Paper: Green et al. (2016)¹²⁵

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Mean and Min/max size values given (1.4-707 µm for PLA, 2.5-316 µm for HDPE, 8.7-478 µm for PVC) but not confirmed | 1 |
| 2 | Particle shape | Not mentioned | 0 |
| 3 | Polymer type | Polymer types given (PLA, HDPE and PVC) but not confirmed | 1 |
| 4 | Source MP | Not mentioned | 0 |
| 5 | Data reporting | % of wet sediment weight | 1 |
| 6 | Chemical purity | They did not attempt to separate physical and chemical effects of MP | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Not assessed | 0 |
| 12 | Replication | 5 replicates | 2 |
| Subtotal | | | 5 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Biomass, feeding activity, bioturbation, metabolic rate | 2 |
| 14 | Presence of natural (food) particles | Natural sediment present | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 4 concentrations including control | 0 |
| 17 | Concentration range tested | Includes at least two environmentally realistic concentrations | 2 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | 3 polymer types separately tested, wide range of sizes (more than 1 order or magnitude) | 1 |
| 20 | Exposure time | Chronic exposure (31 days) | 2 |
| Total | | | 15 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Lowered egestion, metabolic rates increased, biomass decreased, alteration in burrowing behavior | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “After one month of exposure to MP the biological activity of lugworms as measured by oxygen consumption was altered. This <i>could</i> be a response to stress (6) induced by the MP.” “It is not uncommon for marine organisms that increase their respiration as a stress response to also reduce feeding and suffer from overall energy loss.” “Alterations in the burrowing behavior may have occurred as an indirect consequence of MP due to changes to the microphytobenthos.” “Stronger effects of PVC may be due to chemical leaching (4) of residual vinyl chloride monomers , which are toxic.” | |

Paper: Green et al. (2017)¹²⁶

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Mean and Min/max size values (0.6 - 363 µm for PLA, 0.48 - 316 µm for HDPE) but not confirmed | 1 |
| 2 | Particle shape | Shape not mentioned | 0 |
| 3 | Polymer type | Polymer types given (PLA and HDPE) but not confirmed | 1 |
| 4 | Source MP | Not given | 0 |
| 5 | Data reporting | Data reported in µg/L and particles/L | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Verification of exposure along the experiment using a hemocytometer (Table S1) | 2 |
| 10 | Homogeneity of exposure | No aggregations of microalgae and MP were observed during the experiment. | 1 |
| 11 | Exposure assessment of organisms | Not assessed | 0 |
| 12 | Replication | 5 replicates | 2 |
| Subtotal | | | 9 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Filtration of two bivalves, community abundance and biomass | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations used, including control | 0 |
| 17 | Concentration range tested | All concentrations are environmentally realistic, although the higher one is rare | 2 |
| 18 | Aging and biofouling | Added to microalgae for 3 days, additionally the exposure time was long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | 2 polymer types separately tested, wide range of sizes (more than 1 order or magnitude) | 1 |
| 20 | Exposure time | 50 days | 2 |
| Total | | | 19 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Reduced filtration by <i>M. edulis</i> and increased filtration by <i>O. edulis</i> | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | - | |

Paper: Green et al. (2019)¹²⁷

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Min/max (PLA 0.6 - 363 µm) (HDPE 0.48 - 316 µm) and average sizes given (PLA 65.6 µm) (HDPE 102.6 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape (fragment) mentioned but not confirmed | 1 |
| 3 | Polymer type | Polymer type (PLA and HDPE) given but not confirmed | 1 |
| 4 | Source MP | Not mentioned | 0 |
| 5 | Data reporting | Data reported in particles/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Deliberate pulse exposure to mimic realism, however, not homogeneous | 0 |
| 11 | Exposure of assessment of organisms | Not assessed | 0 |
| 12 | Replication | 5 replicates | 2 |
| Subtotal | | | 6 |
| <i>Applicable for risk assessment</i> | | | |
| <i>Criterion</i> | <i>Explanation</i> | | |
| 13 | Endpoints | Tenacity, number of byssal threads and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Food and sediment added | 2 |
| 15 | Detection of effect thresholds | 2 concentrations including control | 0 |
| 16 | Quality of dose-response relationship | Effect thresholds are not reported explicitly | 0 |
| 17 | Concentration range tested | Higher than environmental concentrations | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow for biofouling | 1 |
| 19 | Diversity of MP tested | 2 polymer types separately tested, various shapes, wide range of sizes | 1 |
| 20 | Exposure time | 52 days | 2 |
| Total | | | 14 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Reduced number of byssal threads produced and attachment strength. Altered haemolymph proteome and changes in protein abundances | |
| * | Demonstrated mechanism | “Exposure of mussels to either PLA or HDPE MP resulted in changes in the immunological profiles (8) of their haemolymph .” | |
| * | Speculated mechanism | “These immunological changes <i>may</i> be due to physical abrasion (1) from the MP after being ingested by the mussels.” “The reduction in the abundance of metabolic proteins <i>may</i> , therefore, be associated with reduced feeding (2) but further research is needed to establish this causal link.” | |

Paper: Hämer et al. (2014)¹²⁸

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Average size given for PS spheres (10 µm) min/max size values given for PS fragments (1 - 100 µm) and PA fibers (20 – 2.500 µm), but not measured | 1 |
| 2 | Particle shape | Shapes given (sphere, fragment, fiber) but not confirmed | 1 |
| 3 | Polymer type | Polymer types given (PS and PA) but not confirmed | 1 |
| 4 | Source MP | Sources given (Thermo Scientific; Magic Pyramide) but fragments and fibers were self-cut. Not enough info is provided for reproducibility of fibers | 2 |
| 5 | Data reporting | Data reported as particles per mg of food for spheres and fragments and µg per mg of food for fibers | 1 |
| 6 | Chemical purity | Spheres were washed with demineralized water. Fragments and fibers were not | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Verification of exposure at start of the experiment only for fragments. | 1 |
| 10 | Homogeneity of exposure | Spheres and fragments were homogenized in agarose, pictures of food preparations are shown in SI | 2 |
| 11 | Exposure assessment of organisms | Ingestion quantified, however in separate experiment | 1 |
| 12 | Replication | 24 replicates (although not clearly explained) | 2 |
| Subtotal | | | 12 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, growth, intermolt duration, ingestion rate | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not assessed with L(E)Cx, LOEC or NOEC (only 2 concentration tested including control) | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | No comparison made to MEC | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP. Mechanical grinding of plastic fragments | 1 |
| 19 | Diversity of MP tested | 3 polymer types separately tested, wide range of sizes (more than 1 order of magnitude for fragments and fibers) | 1 |
| 20 | Exposure time | 6-7 weeks | 2 |
| Total | | | 20 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects found. | |

Paper: Hankins, Duffy and Drisco (2018)¹²⁹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Min/max size values given (90-106 µm, 425-500 µm, 850-1000 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (bead) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PE) but not confirmed | 1 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | Data in mg/L and particles/L | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Use of 100% cotton clothing, no other measures taken | 1 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Description of how microbeads are kept in suspension | 1 |
| 11 | Exposure assessment of organisms | Ingestion visually assessed | 1 |
| 12 | Replication | 5 replicates | 2 |
| <u>Subtotal</u> | | | <u>12</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Calcification | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | No comparison made with MEC | 0 |
| 18 | Aging and biofouling | MP are placed in culture for six weeks to grow biofilm, not characterized | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, small size ranges | 0 |
| 20 | Exposure time | 48 hours | 1 |
| <u>Total</u> | | | <u>18</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects found | |

Paper: Imhof and Laforsch (2016)¹³⁰

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Mean and min/max size values given (4.64 - 602 µm) and confirmed with microscope | 2 |
| 2 | Particle shape | Shape given (irregular) and visible in figure S1 | 2 |
| 3 | Polymer type | Polymer types given (PA, PET, PC, PS, PVC) but not confirmed | 1 |
| 4 | Source MP | Source and description of how it is made in the lab given for all polymers, except for one, for which enough information is given for reproducibility. | 2 |
| 5 | Data reporting | Data reported as % plastic in food | 1 |
| 6 | Chemical purity | Not ruled out | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Exposure was not homogeneous, as MP were added on top of the food | 0 |
| 11 | Exposure assessment of organisms | Ingestion qualitatively assessed using pictures of faces from the same experiment | 1 |
| 12 | Replication | 7 replicates | 2 |
| <u>Subtotal</u> | | | <u>11</u> |
| <i>Applicable for risk assessment</i> | | | |
| <i>Criterion</i> | <i>Explanation</i> | | |
| 13 | Endpoints | Growth, Reproduction | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations including control | 0 |
| 17 | Concentration range tested | No environmentally realistic concentrations included/ no comparison made to MEC | 0 |
| 18 | Aging and biofouling | Plastic pellets were ground with grinder. Exposure time of 8 weeks allows for aging of MP. | 2 |
| 19 | Diversity of MP tested | Different polymer types with wide size ranges and a variety of shapes | 2 |
| 20 | Exposure time | 8 weeks | 2 |
| <u>Total</u> | | | <u>21</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects found | |

Paper: Imhof et al. (2017)¹³¹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average sizes measured and given with standard deviation (27.5 - 72.5; 22.3 - 48.5 µm) | 2 |
| 2 | Particle shape | Shape given (fragment) and shown | 2 |
| 3 | Polymer type | Polymer types given (PA, PC, PET, PVC and ABS terpolymer, PVC, POM homopolymer, and SAN) but not confirmed | 1 |
| 4 | Source MP | Sources given (Teijin Kasei America, BASF SE, Styrolution Group, DuPont, Granulat) | 2 |
| 5 | Data reporting | Data reported in particles/mL | 1 |
| 6 | Chemical purity | Effects of additives are not ruled out | 0 |
| 7 | Lab preparation | Experimental containers covered with glass plates | 1 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | MP sank to the bottom of the experimental glasses | 0 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified in preliminary experiment | 1 |
| 12 | Replication | At least 6 replicates | 2 |
| Subtotal | | | 12 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, growth, reproduction and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | Authors state they used an environmentally realistic concentration and give ref | 1 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | Two mixtures with 4 polymers, various shapes, small size ranges | 1 |
| 20 | Exposure time | Up to 22 days | 2 |
| Total | | | 21 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Ingestion of MP from plastic did not affect mortality, development or reproductive parameters of <i>Daphnia</i> . Small responses on morphological traits, alterations in gene expression | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanisms | “Downregulated gene GST indicates that MP may also interact with pathways related to oxidative stress (10).” “MP might act directly on morphological traits or leaching additives (4) might interact with the signaling pathway responsible for inducing phenotypic plastic responses in defensive traits.” | |

Paper: Jabeen et al. (2018)¹³²

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|--|--|--|--------------|
| 1 | Particle size | Reported (Fibers: 0.7 mm - 5 mm; fragments: 2.5 - 3 mm; pellets: 4.9-5 mm) but not confirmed | 1 |
| 2 | Particle shape | Shapes given but not confirmed pictures of shapes and reported | 2 |
| 3 | Polymer type | Polymer types given (EVA, PS, PA) and confirmed with Raman | 2 |
| 4 | Source MP | Collected from field (fibers) and purchased (fragments and pellets), but name of provider not given. Information given is incomplete and hence not fully reproducible. | 1 |
| 5 | Data reporting | Data reported in g (food + MP)/g ww fish and particles/fish | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | State that commercial fish food is completely free from MP contamination, no further measures taken. | 0 |
| 8 | Verification of background contamination | Fish food was said to be free of MP, but overall background not verified | 0 |
| 9 | Verification of exposure | There might have been small variations on average, but fish were dosed with a concentration of 0.96%, 1.36%, 1.94% and 3.81% (g (food þ MP)/g ww fish), which was equal to 0, 55e76, 15, 15 and 15 MP fed to each fish for control, fibers, fragments and pellets groups respectively. | 2 |
| 10 | Homogeneity of exposure | MP were mixed with food and fish feed actively on food particles. | 2 |
| 11 | Exposure of assessment of organisms | Ingestion quantitatively assessed in the same experiment with stereomicroscope | 1 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 15 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Mortality, condition factor, length, weight and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Plastic-amended food, so natural particles present | 2 |
| 15 | Detection of effect thresholds | Not applicable (only 2 concentrations tested including control) | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | Exposure time of experiment is long enough for biofilm to develop | 1 |
| 19 | Diversity of MP tested | For each shape category tested, there was a range of sizes reported | 1 |
| 20 | Exposure time | 6 weeks | 1 |
| Total | | | 22 |
| <i>Mechanisms explaining adverse effects</i> | | | |
| * | Effects | No mortality, but erosion of mucous cells and inflammation in upper jaw. | |
| * | Demonstrated mechanism | Breakage of filaments in gills and impacts on intestinal linings (1) due to fibers. Sharp edges of fragments cause abnormalities in jaws tissue , erosion of the lower jaw mucous cells in the upper jaw and detachment of superficial layer and breakage of the dermal and hypodermal layer (5) . Damage of liver tissue by fibers | |

*

Speculated mechanism

Some effects in liver *speculated* to be caused by plastic **associated toxicants (4)**. Structural changes in intestine due to food containing fibers.

Paper: Jacob et al. (2019)¹³³

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Average size (91.26 µm) given but not measured | 1 |
| 2 | Particle shape | Shape given (bead) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) but not measured | 1 |
| 4 | Source MP | Source given (Polysciences Warrington) | 2 |
| 5 | Data reporting | Data reported in MP/mL and mg/L | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Aquaria equipped with an air stone to maintain MP in suspension | 1 |
| 11 | Exposure assessment of organisms | Ingestion visually inspected with microscope | 1 |
| 12 | Replication | 6 replicates | 2 |
| Subtotal | | | 11 |
| <i>Applicable for risk assessment</i> | | | |
| | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, foraging activity and predation | 2 |
| 14 | Presence of natural (food) particles | Addition of food only once in the 8-day experiment | 1 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | Unrealistic concentration used | 0 |
| 18 | Aging and biofouling | Not mentioned | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 3,5 and 8 days | 0 |
| Total | | | 14 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects found | |

Paper: Jaikumar et al. (2018)¹³⁴

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Min/max values given (1 - 5 µm for spheres, 1 - 10 µm for irregular MP) confirmed with TEM, but Particle Size Distribution or average with standard deviation not given | 1 |
| 2 | Particle shape | Shapes given (irregular, sphere) and shown in TEM images | 2 |
| 3 | Polymer type | Polymer type given for irregular MP (PE) but not confirmed | 1 |
| 4 | Source MP | Source given (Cospheric) and preparation well described | 2 |
| 5 | Data reporting | Data reported particles/mL | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Use of Tween-80 | 1 |
| 11 | Exposure assessment of organisms | Not addressed | 0 |
| 12 | Replication | 4 replicates | 2 |
| Subtotal | | | 10 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Mortality | 2 |
| 14 | Presence of natural (food) particles | No addition of food or natural particles | 0 |
| 15 | Detection of effect thresholds | Detection of effect thresholds (LC ₅₀) without confidence intervals | 1 |
| 16 | Quality of dose-response relationship | 6 concentrations including control | 2 |
| 17 | Concentration range tested | Only unrealistic concentrations used | 0 |
| 18 | Aging and biofouling | Manufactured secondary MP and photographed. No aging | 1 |
| 19 | Diversity of MP tested | One polymer type, two shapes separately tested, small size ranges | 0 |
| 20 | Exposure time | 96 hours | 1 |
| Total | | | 17 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Effects on survival | |
| * | Demonstrated mechanism | “Associated mechanisms warrant further investigations.” | |
| * | Speculated mechanism | “Therefore, the observed effects <i>may</i> have been influenced by plastic additives or unbound monomers of particles. However, this is <i>unlikely</i> as no toxic effects of leachates from plastics have been detected for <i>D. magna</i> , even at much higher exposure concentrations than those used in the present study. The propensity of MP to form aggregates in the gut following ingestion has been previously described and <i>suggested</i> to cause internal abrasions (1) and mechanical damage .” | |

Paper: Jaikumar et al. (2019)¹³⁵

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Min/max sizes given (1-5 µm) and confirmed with TEM (but shown in Jaikumar et al. 2018). No particle distribution or indication of measurement error | 1 |
| 2 | Particle shape | Shapes given (sphere, irregular) and confirmed with TEM (but shown in paper Jaikumar et al. 2018) | 2 |
| 3 | Polymer type | Polymer type not given for PMP and given for SMP (PE) but not confirmed | 1 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | Data provided in particles/ml | 1 |
| 6 | Chemical purity | Cleaned only with milli water to remove Tween | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Verification of concentration with hemocytometer at start of experiment | 1 |
| 10 | Homogeneity of exposure | Addition of tween 80 | 1 |
| 11 | Exposure of assessment of organisms | Ingestion qualitatively assessed in separate experiment (but data not shown) | 1 |
| 12 | Replication | At least 12 replicates | 2 |
| Subtotal | | | 12 |
| <i>Applicable for risk assessment</i> | | | |
| 13 | Endpoints | Day of first brood, Size of first brood, Total # of broods, Size of first 3 broods, Cumulative number of neonates, Terminal length | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | NOEC, LOEC reported, but not with error data | 1 |
| 16 | Quality of dose-response relationship | 5 concentrations including control | 1 |
| 17 | Concentration range tested | Higher than reported in the environment | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow for biofouling | 1 |
| 19 | Diversity of MP tested | Various shapes, small size range | 1 |
| 20 | Exposure time | Up to 21 days | 2 |
| Total | | | 22 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Decline of reproductive output | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “MP can also result in reduced feeding rates (2) in organisms including <i>D. magna</i> which can impair the energy budget . The body size (length of carapax) and the mesh size of the filtering apparatus are defining the size range of particles ingested allowing <i>D. magna</i> to ingest particles between 200 nm and 90 µm and <i>C. dubia</i> to ingest particles with sizes up to 25 µm.” | |

Paper: Jemec et al. (2016)⁹⁷

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|--|--|---|------------------|
| 1 | Particle size | Size distribution given length 62-1400 µm, width 31-528 µm + confirmed with OM and SEM | 2 |
| 2 | Particle shape | Fiber shape given and confirmed with OM and SEM | 2 |
| 3 | Polymer type | Polymer type given (PET) and confirmed using FTIR | 2 |
| 4 | Source MP | It is said that it was commercial PET, but it was not stated where it was purchased. | 1 |
| 5 | Data reporting | Data reported as mg/L and MP/L | 2 |
| 6 | Chemical purity | MP rinsed with water and ethanol to remove surface contamination, and leachates chemically analyzed. Controls with medium and with leachates. | 2 |
| 7 | Lab preparation | No information provided | 0 |
| 8 | Verification of background contamination | No information provided | 0 |
| 9 | Verification of exposure | No verification of the exposure | 0 |
| 10 | Homogeneity of exposure | Not homogeneous (clumps and aggregates formed Fig. S1) | 0 |
| 11 | Exposure of assessment of organisms | Ingestion qualitatively assessed with FTIR in the same experiment | 1 |
| 12 | Replication | 2 replicates | 1 |
| <u>Subtotal</u> | | | <u>13</u> |
| <i>Applicable for risk assessment</i> | | | |
| <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> | |
| 13 | Endpoints | Survival, Growth | 2 |
| 14 | Presence of natural (food) particles | One series pre-fed with algae, but then no food was added during actual exposure | 0 |
| 15 | Detection of effect thresholds | Design fully correct to detect EC50, however the effect was not dose dependent | 2 |
| 16 | Quality of dose-response relationship | 5 concentrations including the control | 1 |
| 17 | Concentration range tested | Higher than realistic, but they may resemble hot spots | 1 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, but wide size range tested | 1 |
| 20 | Exposure time | 2 days | 0 |
| <u>Total</u> | | | <u>20</u> |
| <i>Mechanisms explaining adverse effects</i> | | | |
| * | Effects | No effects found if daphnids were fed | |

Paper: Jemec Kokalj, Kunej and Skalar (2018)²⁴

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---|--|--|------------------|
| 1 | Particle size | Mean diameter with standard error given 183.1 ± 92.46 , 102.9 ± 29.1 , 63.05 ± 24.75 , 264 ± 128.3 , 247.9 ± 123.6 , 136.8 ± 50.89 , 22.8 ± 6.11 μm and measured with DLS | 2 |
| 2 | Particle shape | Shapes given (fragment, fiber) and confirmed with SEM pictures | 2 |
| 3 | Polymer type | Polymer type given (PE, PET) and confirmed in previous study Jemec et al. (2016) | 2 |
| 4 | Source MP | Description of production in own lab, however source/manufacturer of commercial facial cleaner is not mentioned | 1 |
| 5 | Data reporting | Data reported in mg/L and number of particles/mg | 2 |
| 6 | Chemical purity | Washed with water to remove soap or NaCl, but not plastic-associated chemicals | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Static exposure, without mixing | 0 |
| 11 | Exposure of assessment of organisms | Ingestion qualitatively assessed with stereomicroscope | 1 |
| 12 | Replication | 2 replicates ISO 6341:2012 | 1 |
| <u>Subtotal</u> | | | <u>11</u> |
| <i>Applicable for risk assessment</i> | | | |
| 13 | Endpoints | Survival, Growth and Immobility | 2 |
| 14 | Presence of natural (food) particles | Feeding only prior to exposure | 0 |
| 15 | Detection of effect thresholds | Not applicable (only 2 concentrations tested including control) | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | Higher than environmentally realistic values. Therefore, the organisms are not likely to encounter such concentrations unless in the case of pollution hot spots. (no comparison with MEC hot spot) | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, wide range of sizes, however separately tested | 0 |
| 20 | Exposure time | <i>Daphnia magna</i> , 24 h; brine shrimp <i>Artemia franciscana</i> , 48 h | 0 |
| <u>Total</u> | | | <u>13</u> |
| <i>Mechanisms explaining adverse effects</i> | | | |
| * | Effects | No effects found | |

Paper: Jeong et al. (2016)¹³⁶

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average size given (6 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape mentioned (bead) but not confirmed | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Polyscience) | 2 |
| 5 | Data reporting | Data reported in µg/ml | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Ingestion qualitatively assessed using a microscope in separate experiment | 1 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 9 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Growth, fecundity, life span, reproduction time and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 5 concentrations including control | 1 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | Pristine microbeads used | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | Chronic exposure (12 days) | 1 |
| Total | | | 15 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Reduced growth rate, reduced fecundity, decreased lifespan, longer reproduction time, antioxidant-related enzymes and MAPK signaling were activated | |
| * | Demonstrated mechanism | “Taken together, our data <i>indicate</i> that exposure to MP significantly increases the level of ROS (10) in a size-dependent manner and that the activation of antioxidant-related enzymes including SOD, GST, GR, and GPx is directly related to a defense mechanism against MP-induced oxidative stress (10) .” | |
| * | Speculated mechanism | “Most <i>likely</i> explanations for these findings are linked to insufficient nutrition (2) due to ingestion of MP instead of diet” | |

Paper: Jeong et al. (2017)³⁶

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Average size given (6 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (bead) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Polyscience) | 2 |
| 5 | Data reporting | Data reported in µg/mL and particles/mL | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Ingestion qualitatively assessed in preliminary experiment | 1 |
| 12 | Replication | 3 replicates | 2 |
| <u>Subtotal</u> | | | <u>10</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Growth rate, fecundity, development | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 5 concentrations including control | 1 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 24h | 0 |
| <u>Total</u> | | | <u>15</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Increased GR, GST and SOD activity (MP 6µm) | |
| * | Demonstrated mechanism | “Activation of GSH-related antioxidant enzymes was involved in MP-induced oxidative stress (10) , implying that these processes are likely to be an important defense mechanism against MP-induced oxidative stress in <i>P. nana</i> .” | |
| * | Speculated mechanism | - | |

Paper: Jin et al. (2018)¹³⁷

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average sizes given (50 µm), confirmed with SEM, but standard deviation not given | 1 |
| 2 | Particle shape | Shape given (sphere) and shown in SEM images | 2 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Microsphere-Nanospheres) | 2 |
| 5 | Data reporting | Data reported in µg/L and particles/L | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Not addressed | 0 |
| 12 | Replication | 2 replicates | 1 |
| Subtotal | | | 9 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Abundance, richness and diversity of gut microbiota, cannot be linked to threat on the population level | 0 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations, including control | 0 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 14 days | 0 |
| Total | | | 11 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “Change in richness and diversity of microbiota in the gut” | |
| * | Demonstrated mechanism | “MP <i>influence</i> the volume of mucus and <i>induce</i> microbiota dysbiosis (microbial imbalance (7)) in the gut of adult zebrafish.” “We have <i>observed</i> that different sizes of polystyrene MP can induce microbiota dysbiosis and inflammation in the adult zebrafish gut (1) after a 14-day exposure.” | |
| * | Speculated mechanism | - | |

Paper: Jovanović et al. (2018)⁸⁵

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Average sizes measured and given (75.6, 111.7, 23.4, 51, 54.5, 87.6 µm) with SD | 2 |
| 2 | Particle shape | Shapes not described in the text but shown in pictures | 2 |
| 3 | Polymer type | Polymer types given (PVCHMW, PA, UHMWPE, PS, MDPE, PWCLMW) but not confirmed | 1 |
| 4 | Source MP | Source given (Sigma-Aldrich) and preparation well described | 2 |
| 5 | Data reporting | Data reported in g/kg of fish weight | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Description of mixed fish feed | 1 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified | 1 |
| 12 | Replication | No replication | 0 |
| <u>Subtotal</u> | | | <u>10</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Growth rate | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations, including control | 0 |
| 17 | Concentration range tested | They say its environmentally realistic, however no MEC provided | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | 6 different polymer types separately tested, small size ranges separately tested | 0 |
| 20 | Exposure time | 45 days | 1 |
| <u>Total</u> | | | <u>16</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effect on stress, growth rate, pathology, or accumulation in gastrointestinal tract | |

Paper: Kalčíková et al. (2017)¹³⁸

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Min/max and average sizes given (30-600, 71.30±34.29 µm; 40-400, 96.00±69.99 µm) with standard deviation | 2 |
| 2 | Particle shape | Shape given (irregular) and shown in SEM images | 2 |
| 3 | Polymer type | Polymer type given (PE) and measured by IR analysis | 2 |
| 4 | Source MP | Description how MP stock is made, however the source of exfoliating products is not mentioned | 1 |
| 5 | Data reporting | Data reported in mg/L | 1 |
| 6 | Chemical purity | Microbead leachate was used as control | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | MP adsorption onto root surface qualitatively assessed | 1 |
| 12 | Replication | 3 replicates | 2 |
| <u>Subtotal</u> | | | <u>12</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Growth rate, chlorophyll a and b | 2 |
| 14 | Presence of natural (food) particles | NA | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 4 concentrations, including control | 0 |
| 17 | Concentration range tested | No comparison made with MEC | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, variety of shapes, wide size range (1 order of magnitude) | 1 |
| 20 | Exposure time | 7 days | 1 |
| <u>Total</u> | | | <u>18</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Affected root growth, reduced viability of root cells | |
| * | Demonstrated mechanism | “Microbeads affected the root growth of <i>Lemna minor</i> through the adsorption onto the root surface and mechanically damaging (5) of the roots (fig. 6 for <i>evidence</i>).” Microbeads with sharp edges <i>affect</i> root cell viability. Microbeads with smooth surface do not. | |
| * | Speculated mechanism | - | |

Paper: Kaposi et al. (2014)⁹⁹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Min/max size values given (10 - 45 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape given (sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PE) but not checked | 1 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | Data reported in spheres/ml | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Daily checked using a microscope for the effect experiment | 2 |
| 10 | Homogeneity of exposure | Gentle aeration kept the microspheres in suspension | 1 |
| 11 | Exposure assessment of organisms | Ingestion and egestion visually quantified in the same experiment | 1 |
| 12 | Replication | 5 replicates | 2 |
| Subtotal | | | 12 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival and growth | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 5 concentrations, including control | 1 |
| 17 | Concentration range tested | No environmentally realistic concentration | 0 |
| 18 | Aging and biofouling | Pristine microsphere used. Aged in a separate experiment | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, small range of sizes | 0 |
| 20 | Exposure time | 5 days | 1 |
| Total | | | 18 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “Small non-dose dependent effect on larval growth” | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “The reduced body width of <i>T. gratilla</i> larvae exposed to 300 spheres/mL may be the result of reduced feeding efficiencies (2) associated with the ingestion of MP.” | |

Paper: Karami et al. (2017)¹³⁹

| Technical subset | Criterion | Explanation | Score |
|---------------------------------------|--|--|-----------|
| 1 | Particle size | Particle Size Distribution measured, D10 (4.64 µm), D50 (10.9 µm) and D90 (17.6 µm) values given but without error | 1 |
| 2 | Particle shape | Shape given (irregular) and shown in SEM images | 2 |
| 3 | Polymer type | Polymer type given (LDPE) but not confirmed | 1 |
| 4 | Source MP | Source given (Toxemerge Pty Ltd) | 2 |
| 5 | Data reporting | Data reported in µg/L and particles/L | 2 |
| 6 | Chemical purity | Solvent control used. MP analyzed for heavy metals, PCB's, phthalates and PAHs. | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Use of ethanol as solvent to minimize particle aggregation. Aeration of beakers. | 1 |
| 11 | Exposure assessment of organisms | Ingestion qualitatively assessed | 1 |
| 12 | Replication | 5 Replicates | 2 |
| Subtotal | | | 13 |
| Applicable for risk assessment | | | |
| 13 | Endpoints | Condition factor, length, weight | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 4 concentrations, including control | 0 |
| 17 | Concentration range tested | More than one realistic concentration tested | 2 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, variety of shapes, small size range | 1 |
| 20 | Exposure time | 10 and 20 days | 0 |
| Total | | | 21 |
| Mechanisms explaining effects | | | |
| * | Effects | Exposure to LDPE fragments has <i>minimal impact</i> on biomarker responses | |

Paper: Korez et al. (2019)²¹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Min/max average sizes given (10-100 µm), confirmed with microscope but size distribution or error not reported | 1 |
| 2 | Particle shape | Shape given (fragments), however not shown | 1 |
| 3 | Polymer type | Polymer type given (PMMA) but not confirmed | 1 |
| 4 | Source MP | Source given (Magic Pyramid Bruecher & Partner KG, Frechen, Germany) and description on how granulate was made | 2 |
| 5 | Data reporting | Data reported in particles/mg of food and µg/mg food | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Verification of exposure at the beginning of experiments in stock solution. | 1 |
| 10 | Homogeneity of exposure | Homogeneous dispersion of MP in food visually checked | 2 |
| 11 | Exposure of assessment | Not mentioned | 0 |
| 12 | Replication | 20 replicates | 2 |
| <u>Subtotal</u> | | | <u>12</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Feeding rate and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations, including control | 0 |
| 17 | Concentration range tested | Only unrealistic concentrations used | 0 |
| 18 | Aging and biofouling | Not mentioned | 0 |
| 19 | Diversity of MP tested | One polymer type, various shapes, wide range of sizes | 1 |
| 20 | Exposure time | 8 days | 1 |
| <u>Total</u> | | | <u>18</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Isopods, <i>I. emarginata</i> , are not affected by MP when they receive sufficient natural food of high nutritional quality. | |

Paper: Kratina et al. (2019)¹⁴⁰

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Average size given (40.2 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape given (sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PMMA) but not confirmed | 1 |
| 4 | Source MP | Source given (Spherotech) | 2 |
| 5 | Data reporting | Data reported in MP/cm ² | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Use of glass material to minimize contamination. Lids were placed on all microcosms to prevent contamination. | 1 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Verification of exposure at the beginning of exposure. | 1 |
| 10 | Homogeneity of exposure | Not throughout whole medium, but equal distribution at the bottom where food is placed. | 0 |
| 11 | Exposure of assessment | Not addressed | 0 |
| 12 | Replication | 3-7 replicates | 2 |
| Subtotal | | | 10 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Feeding rate, metabolic rate | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 11 concentrations, including control | 2 |
| 17 | Concentration range tested | More than 1 environmentally relevant concentration was used within the range tested. | 2 |
| 18 | Aging and biofouling | Not mentioned | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 24 h | 0 |
| Total | | | 18 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Metabolic rate increased with MP concentration at lowest temperature but decreased at the higher temperatures. MP did not affect feeding rates | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “It is <i>likely</i> that the physical presence of non-nutritious MP in place of food, can lead to longer gut passage times (2) and adverse biological impacts . A reduction in metabolism due to a combination of warming and high concentration of MP <i>could</i> further reduce the amount of energy assimilated for individual and population growth rates.” “Suppression of metabolic rates through exposure to MP has been described in other aquatic organisms, highlighting the <i>potential</i> for these tiny pollutants to impede physiological performance (8) .” | |

Paper: Lee et al. (2013)³

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average size given (6 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape mentioned (bead) but not confirmed | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Polyscience) | 2 |
| 5 | Data reporting | Data reported in µg/ml and particles/ml | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Test solutions were sonicated for 30 min immediately prior to use in each experiment aggregates were still found in 50nm beads tested. | 0 |
| 11 | Exposure assessment of organisms | Ingestion quantitatively assessed in separate experiment | 1 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 10 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, developmental time and fecundity | 2 |
| 14 | Presence of natural (food) particles | No feeding during acute test, feeding during chronic test | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are reported as LC50 and EC50 | 2 |
| 16 | Quality of dose-response relationship | Acute test 8 concentrations (including control), chronic test 5 concentrations (including control) | 2 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | Pristine microbeads used. No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | Acute (96 h) and Chronic (until adult females developed egg sacs, 14 days on average) | 1 |
| Total | | | 19 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “Decrease in fecundity” | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “This effect <i>can</i> be attributed to insufficient nutrition (2) or the inhibition of digestion (1) due to the large amount of MP ingested as prey.” “PS beads <i>may</i> physically inhibit (5) the fertilization of copepods.” | |

Paper: Lei et al. (2018)¹⁴¹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | PSD measured with a Zetasizer (20-180 µm for PA, 20-120 µm for PE, 40-180 µm for PP, 50-170 µm for PVC µm, 5 µm for PS) | 2 |
| 2 | Particle shape | Shapes not described in the text but shown in pictures | 2 |
| 3 | Polymer type | Polymer types given (PA, PE, PP, PVC, PS) and confirmed with FTIR | 2 |
| 4 | Source MP | Sources given (Sigma-Aldrich, Aladdin, Sinopharm Chemical) | 2 |
| 5 | Data reporting | Data reported in mg/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Not addressed | 0 |
| 12 | Replication | 3 replicates | 2 |
| <u>Subtotal</u> | | | <u>11</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, development and reproduction | 2 |
| 14 | Presence of natural (food) particles | Addition food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 5 concentrations for <i>C. elegans</i> , 6 concentrations for <i>D. rerio</i> , including control | 2 |
| 17 | Concentration range tested | More than 1 environmentally realistic concentration | 2 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | 5 different polymer types separately tested, small size ranges separately tested | 0 |
| 20 | Exposure time | 2 days for <i>C. elegans</i> (2 days considered sufficient for the nematode), 10 days for <i>D. rerio</i> | 1 |
| <u>Total</u> | | | <u>20</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “MP caused intestinal damage in <i>D. rerio</i> MP inhibited survival rates, body length and reproduction of <i>C. elegans</i> .” “Reduced calcium levels but increased expression of the glutathione S-transferase 4 enzyme in the intestine” | |
| * | Demonstrated mechanism | “Reduced calcium levels but increased expression of the glutathione S-transferase 4 enzyme in the intestine, which <i>indicates intestinal damage (1)</i> and <i>oxidative stress (10)</i> are major effects of MP exposure.” “These results <i>provide evidence of oxidative damage (10)</i> due to <i>ingestion and accumulation</i> of MP.” | |
| * | Speculated mechanism | - | |

Paper: LeMoine et al. (2018)¹⁴²

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|--|--|---|--------------|
| 1 | Particle size | Min/max size values given (10-45 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (sphere) but not checked | 1 |
| 3 | Polymer type | Polymer type given (PE) but not confirmed | 1 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | Data reported in mg/L and particles/ml | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Homogeneous concentration mentioned, not shown | 1 |
| 11 | Exposure of assessment of organisms | Ingestion qualitatively assessed in the same experiment with fluorescence microscopy | 1 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 11 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, Growth, Hatching and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Not applicable (only 3 concentrations tested, including control) | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations, including control | 0 |
| 17 | Concentration range tested | Only high concentrations | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, a small range of sizes | 0 |
| 20 | Exposure time | 14 days | 1 |
| Total | | | 16 |
| <i>Mechanisms explaining adverse effects</i> | | | |
| * | Effects | “Could not detect any detrimental effects of these particles on larval development, growth or metabolism.” “Transient changes in expression of thousands of genes within 48 h exposure, which largely disappeared by 14 days.” | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “We contend that as they occur during a critical phase of development and target essential processes (8) (e.g., brain and neural differentiation, metabolism), MP exposure <i>may</i> therefore have direct and indirect repercussions on the animals’ fitness at later life stages warranting further investigation.” | |

Paper: Leung and Chan (2018)¹⁴³

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Min/max size values given (8-12 µm and 32-38 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Spherotech, Cospheric) | 2 |
| 5 | Data reporting | Data in beads/ml | 1 |
| 6 | Chemical purity | Beads were rinsed and soaked overnight in FSW prior to use | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Quantification of beads with Coulter counter prior to addition. Quantification of beads in sediment samples under a microscope at the end of the experiment. At least 80% of nominal concentration maintained. | 2 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Not addressed | 0 |
| 12 | Replication | At least 6 replicates | 2 |
| Subtotal | | | 10 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Mortality and segment regeneration | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations, including control | 0 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, small size ranges | 0 |
| 20 | Exposure time | 4 weeks | 2 |
| Total | | | 17 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “Increased mortality and reduced the rate of posterior segment regeneration.” | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “Smaller-diameter beads appeared more detrimental, <i>possibility</i> due to their physical property, sorption of organic molecules and bacteria (7) , as well as selective feeding of the worms.” “The hydrophobic chemicals attached and the plasticizers leached (4) could disrupt cellular functions. ” “Therefore, the number of beads ingested by the worms <i>might</i> be higher in the small beads treatments and in turn slowing the regeneration as removal of these non-nutritional particles from the gut could incur energetic cost (2) and/or reduce intake of nutritious particles. ” | |

Paper: Lo and Chan (2018)¹⁴⁴

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Sizes measured but only min/max size values given without error (2-5 µm) | 1 |
| 2 | Particle shape | Shape given (sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Spherotech Inc) | 2 |
| 5 | Data reporting | Data reported in particles/mL | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | (SI) Nominal concentration verified, actual concentration for high concentrations measured after exposure. Environmentally relevant concentrations not verified due to detection limit | 2 |
| 10 | Homogeneity of exposure | (SI) Stirring efficiently created homogenous sample measured with Coulter Counter | 2 |
| 11 | Exposure assessment of organisms | Particle (Algae and MP) removal measured quantitatively, separate experiment | 1 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 13 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, growth, feeding, and percentage of males | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 and 4 concentrations, including control | 0 |
| 17 | Concentration range tested | One environmentally relevant concentration | 1 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, small size range | 0 |
| 20 | Exposure time | 14 and 65 days | 2 |
| Total | | | 21 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effect at environmentally relevant concentrations. Reduced growth at higher concentrations. | |
| | Demonstrated mechanism | - | |
| * | Speculated mechanism | “At higher concentrations, these micro-PS fed larvae consumed a similar number of algae compared to those in control but grew relatively slower than those in the control suggesting that ingestion and/or removal of MP was/were energetically costly (2).” “Selection against, ingesting or egesting MP could incur energetic cost, which in turn affect growth.” Due to limited energy larval settled earlier. Exposure to MP could also negatively affect organism chemically (4) through styrene's monomer. | |

Paper: Lu et al. (2016)¹⁴⁵

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average size given (5 µm) and confirmed with pictures | 2 |
| 2 | Particle shape | Shape given (sphere) and confirmed with pictures | 2 |
| 3 | Polymer type | Polymer type given (PS) and confirmed with FTIR | 2 |
| 4 | Source MP | Source given (Tianjin BaseLine ChromTech Research Centre) | 2 |
| 5 | Data reporting | Data reported in µg/L and particles/ml | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | tanks were continuously aerated to maintain the dispersion of the particles in water. No aggregation was observed in the tanks with aeration (Figure S3) | 2 |
| 11 | Exposure assessment of organisms | Ingestion quantitatively assessed in separate experiment | 1 |
| 12 | Replication | At least 3 replicates | 2 |
| Subtotal | | | 15 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Suborganismal endpoints cannot be unambiguously linked to a threat on the population or individual level. | 0 |
| 14 | Presence of natural (food) particles | Feeding not mentioned (only during maintenance) | 0 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 4 concentrations, including control | 0 |
| 17 | Concentration range tested | At least 1 environmentally realistic concentration | 1 |
| 18 | Aging and biofouling | Virgin PS particles. Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 3 weeks | 0 |
| Total | | | 17 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Inflammation, lipid accumulation, disturbed lipid and energy metabolism. | |
| * | Demonstrated mechanism | “MP caused inflammation and lipid accumulation in fish liver. MP also induced significantly increased activities of superoxide dismutase and catalase, <i>indicating</i> that oxidative stress (10) was induced after treatment with MP.” | |
| * | Speculated mechanism | “Our report on this phenomenon is the first, and this <i>may</i> be attributed to insufficient nutrition (2) or the inhibition of food digestion (1) because more large-sized MP were ingested by the fish.” | |

Paper: Lu et al. (2018)¹⁴⁶

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Sizes measured but only average values given (1, 20, 40, 90 µm) without error | 1 |
| 2 | Particle shape | Shape given (sphere) and shown | 2 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (MicroParticles) | 2 |
| 5 | Data reporting | Data reported in particles/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Description of pretreatment of beads with Tween 20 to prevent aggregation | 1 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified | 1 |
| 12 | Replication | 2 replicates | 1 |
| Subtotal | | | 10 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Gene expression cannot be unambiguously linked to a threat on the population level | 0 |
| 14 | Presence of natural (food) particles | No addition of food | 0 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations, including control | 0 |
| 17 | Concentration range tested | Only unrealistic concentration tested | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, small size range | 0 |
| 20 | Exposure time | 2 hours | 0 |
| Total | | | 10 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Upregulation of genes ifnγ, il1β, igm, s100a and downregulation of genes il1β, s100a, saa, il8gene. | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “The reason is at present unknown, but it can be <i>speculated</i> that minute particles are taken up by gill epithelia and associated cells where after immune gene expression is initiated.” | |

Paper: Magni et al. (2018)¹⁴⁷

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average sizes given (1 and 10 µm), but not confirmed | 1 |
| 2 | Particle shape | Shape given (bead) and shown | 2 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Sigma-Aldrich) | 2 |
| 5 | Data reporting | Data reported in mg/L and particles/L | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Illustration MP without aggregation phenomena | 2 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified | 1 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 13 |
| <i>Applicable for risk assessment</i> | | | |
| 13 | Endpoints | Cellular stress, oxidative damage and neuro- genotoxicity, cannot be linked to a threat on the population level | 0 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations, including control | 0 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, wide size range (1 order of magnitude) | 1 |
| 20 | Exposure time | 6 days | 0 |
| Total | | | 16 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effect on oxidative stress and genetic damage, with the exception of a modulation of catalase glutathione peroxidase activities. we can assert that PMs, contextually to our experimental design, <u>do not have direct adverse effects</u> on the nervous system of zebra mussel. Despite the accumulation of MP in the exposed mussels, our results highlight that MP <u>did not induce great alteration of both oxidative balance and neuro- genotoxicity</u> in zebra mussel, for the selected endpoints and exposure time. | |

Paper: Magni et al. (2019)⁷²

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Average sizes given (1 and 10 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape (sphere) mentioned and shown in pictures | 2 |
| 3 | Polymer type | Polymer type (PS) given but not confirmed | 1 |
| 4 | Source MP | Source given (Sigma Aldrich) | 2 |
| 5 | Data reporting | Data provided in particles/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Verification of the exposure in the stock solutions | 1 |
| 10 | Homogeneity of exposure | No aggregation in the working suspensions, Slow stirring to prevent sedimentation | 1 |
| 11 | Exposure of assessment of organisms | Ingestion qualitatively assessed in same experiment | 1 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 12 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Suborganismal endpoint and no link with population related endpoints provided | 0 |
| 14 | Presence of natural (food) particles | Food added | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations, including control | 0 |
| 17 | Concentration range tested | Higher than environmentally realistic | 0 |
| 18 | Aging and biofouling | Not addressed | 0 |
| 19 | Diversity of MP tested | Only 2 sizes included | 0 |
| 20 | Exposure time | 6 days | 1 |
| Total | | | 15 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “78 proteins were differentially modulated” | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “Overrun of homeostatic responses (8) that are no longer able to fight the injuries made by MP.” “This methodology suggested that M2 was able to create an imbalance in the oxidative status of gill cells which was reflected in the modulation of many proteins involved in some different cellular pathways.” | |

Paper: Malinich *et al.* (2018)

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Min/max size values given (180-212 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (bead) and shown | 2 |
| 3 | Polymer type | Polymer type given (PE) but not confirmed | 1 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | Data reported in both mg/L and particles/L | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Each tank contained an aerator to provide air, circulate food and plastics, and prevent settling of plastics over time | 1 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified | 1 |
| 12 | Replication | 6 replicates | 2 |
| <u>Subtotal</u> | | | <u>12</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Growth | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations, including control | 0 |
| 17 | Concentration range tested | Authors say the highest concentration used was high/unrealistic | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, small size range | 0 |
| 20 | Exposure time | 15 and 30 days | 2 |
| <u>Total</u> | | | <u>19</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effect on consumption or growth. Young juveniles are able to distinguish MP from food particles and are able to tolerate the presence of MP and do not have adverse effect on growth. | |

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average size given in the main text (1 µm), measured with a Zetasizer and particle size distribution provided in the SI | 2 |
| 2 | Particle shape | Shape given (bead) and shown in SEM pictures | 2 |
| 3 | Polymer type | Polymer type given (PS) but not measured | 1 |
| 4 | Source MP | Source given (Sigma-Aldrich) | 2 |
| 5 | Data reporting | Data reported in mg/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Visually shown | 1 |
| 12 | Replication | 3 replicates | 2 |
| | | Subtotal | 11 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Growth, Photosynthesis, Morphology | 2 |
| 14 | Presence of natural (food) particles | NA for algae | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 4 concentrations including control | 0 |
| 17 | Concentration range tested | Authors state that 2 environmentally realistic concentrations were used | 2 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 30 days | 2 |
| | | Total | 20 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “Reduced photosynthetic activity of <i>Chlorella pyrenoidosa</i> , unclear pyrenoids, distorted thylakoids and damaged cell membrane were observed.” However; “could reduce the adverse effects of MP jointly through cell wall thickening, algae homo-aggregation and algae-MP hetero-aggregation, hence triggering an increase of algal photosynthetic activity and its growth, and cell structures turned to normal.” | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “The mechanisms for the toxicity <i>might</i> be attributed to the physical damage (5) and oxidative stress (10) .” | |

Paper: Mateos-Cárdenas et al. (2019)¹⁵⁰

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Min/max sizes given (10-45 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape given (sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PE) but not confirmed | 1 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | Data reported in particles/ml and mg/L | 2 |
| 6 | Chemical purity | Control for Tween-20 run to account for surfactant toxicity, however not all chemical toxicity ruled out | 0 |
| 7 | Lab preparation | Test dishes were covered with soda-lime watch glass dishes with fused edges to avoid contamination | 1 |
| 8 | Verification of background contamination | No verification of background contamination | 0 |
| 9 | Verification of exposure | Number of particles in Lemna minor exposed to Gammarus were quantified (start of experiment) | 1 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Adhesion of MP to <i>L. minor</i> visually quantified and ingestion qualitatively assessed for Gammarus | 1 |
| 12 | Replication | At least 7 replicates | 2 |
| | | Subtotal | 12 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Photosynthetic efficiency and growth (<i>Lemna</i>) and mortality, moulting and fitness (<i>Gammarus</i>) | 2 |
| 14 | Presence of natural (food) particles | Addition of food <i>Gammarus</i> | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | Concentration not environmentally realistic | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, a small range of sizes | 0 |
| 20 | Exposure time | Chronic exposure for <i>Lemna</i> (7 days and 30 days), Acute exposure for <i>Gammarus</i> (48 h) | 2 |
| | | Total | 19 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects found | |

Paper: Mazurais et al. (2015)¹⁵¹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Min/max size values given (10 - 45 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape given (bead) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PE) but not checked | 1 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | Data reported in microbeads/gram | 1 |
| 6 | Chemical purity | PE microbeads were not cleaned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | The concentration of MP in the three diets was confirmed with a microscope at the start of the experiment | 1 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Ingestion qualitatively assessed in the same experiment | 1 |
| 12 | Replication | 6 replicates | 2 |
| <u>Subtotal</u> | | | <u>10</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Mortality and growth | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly with L(E)Cx, LOEC or NOEC | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations including control | 0 |
| 17 | Concentration range tested | No comparison with MEC, authors assume that the quantities of MP ingested correspond to high environmentally relevant concentrations of MP. | 0 |
| 18 | Aging and biofouling | Pristine microbeads used, Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, small range of sizes | 0 |
| 20 | Exposure time | 43 days | 2 |
| <u>Total</u> | | | <u>17</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “The highest concentration slightly impacted mortality rates.” | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “This slight impact is <i>likely</i> to be explained by the apparent high potential of microbeads egestion from the gut.” “While the diameter of the anterior intestine is around 60-80 mm at 29 dph in European sea bass, it <i>is conceivable</i> that microbeads of 45 mm or less used in the present work, when ingested in very high quantities, <i>could block the lumen (1)</i> at earlier stages of development.” “Base on material safety data sheet of PE microbeads. acidic conditions within the gut of fish larvae <i>may</i> produce hazardous decomposition by-products (4) such as oxides of sulfur.” | |

Paper: Murphy and Quinn (2018)¹⁵²

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Max size given (400 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape given (irregular) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PE) and confirmed with FTIR | 2 |
| 4 | Source MP | MP were extracted from a face wash. The information given is incomplete (product name not provided) and hence not fully reproducible. | 1 |
| 5 | Data reporting | Data reported in mg/L; I50 also reported in particles/ml | 2 |
| 6 | Chemical purity | MP were cleaned with ethanol 70% three times and then rinsed with water | 2 |
| 7 | Lab preparation | Not done | 0 |
| 8 | Verification of background contamination | Done for preliminary tests, not for actual tests | 0 |
| 9 | Verification of exposure | No verification of the exposure | 0 |
| 10 | Homogeneity of exposure | Description of the method used to obtain homogeneous exposure. | 1 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified in the same experiment | 1 |
| 12 | Replication | At least 3 replicates | 2 |
| Subtotal | | | 13 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Feeding rates, morphological scores and hydranth number | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are reported; however, no confidence interval is provided. | 1 |
| 16 | Quality of dose-response relationship | 5 concentrations including control | 1 |
| 17 | Concentration range tested | No environmentally realistic concentration used | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, irregularly shaped, wide range of sizes (more than 1 order of magnitude) | 1 |
| 20 | Exposure time | 1 hour | 0 |
| Total | | | 20 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Reduction in feeding rate and (non-lethal) changes to morphology | |
| * | Demonstrated mechanism | “Exposure to MP has the potential to reduce the health of <i>H. attenuata</i> by impacting its ability to feed and limiting the amount of prey consumed.” “Normally it takes <i>H. attenuata</i> less than 8 h to expel any waste material from their gastric cavity, but in the current study this took considerably longer, between 24 and 48 h in some individuals to egest MP. These results <i>indicate</i> that when exposed to MP <i>H. attenuata</i> are expending considerably more time and energy clearing their gastric cavity (2) then under normal conditions.” | |
| * | Speculated mechanism | “MP ingestion <i>may</i> cause internal damage (1) to the gastric cavity, a false sense of satiation (2) and impairment of appendages (5) .” | |

Paper: Ogonowski et al. (2016)²⁹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Min/max and average sizes given for spheres (1 - 5, 4.1±1.0 µm) and irregular particles (2.6 ±1.8 µm) and confirmed with a Spectrex laser particle counter | 2 |
| 2 | Particle shape | Shapes given (sphere and irregular) mentioned but not shown | 1 |
| 3 | Polymer type | Polymer type only given for one of the MP (PE), no confirmation of polymer type | 1 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | Data reported in particles/ml | 1 |
| 6 | Chemical purity | Chemical effects were not ruled out, Tween 80 added | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of the background contamination | 0 |
| 9 | Verification of exposure | Verification of the exposure at the start of the experiment using a hemocytometer | 1 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | Ingestion quantitatively assessed in separate experiment | 1 |
| 12 | Replication | At least 5 replicates | 2 |
| Subtotal | | | 11 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, reproductive output, body size, number of broods, time between broods, age at first reproduction, | 2 |
| 14 | Presence of natural (food) particles | Food and natural particles added | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are given with EC50 for number of broods (SI, Table S3) | 2 |
| 16 | Quality of dose-response relationship | 5 concentrations including control used for adults, 2 concentrations for neonates | 1 |
| 17 | Concentration range tested | No environmentally realistic concentrations included, no ref included | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, 2 shapes separately tested, small range of sizes | 0 |
| 20 | Exposure time | Acute (24 h) and chronic exposure (up to 21 days) | 2 |
| Total | | | 21 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “Exposure to SMP caused elevated mortality, increased inter-brood period and decreased reproduction” | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “Given the observed aggregative properties of SMP, it is <i>possible</i> that the formation and passage of aggregates through the gut <i>might</i> have caused some internal damage (1) and, thereby, contributed to the elevated mortality .” “This <i>suggests</i> that growth effects mainly depend on overall food availability (2) , which is in line with other studies demonstrating both positive and negative effects of natural particulates on the growth of filter feeders.” | |

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Min/max average size value given (1-5 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape given (sphere) but not confirmed | 1 |
| 3 | Polymer type | Not given | 0 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | Data reported in mg/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Verification of the exposure with hemacytometer along the experiment and evidence that at least 80% of the nominal concentration throughout the test is maintained | 2 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure of assessment of organisms | Ingestion was qualitatively confirmed by fluorescence microscopy, and assessed by difference, using mass balance | 1 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 10 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, Growth, Feeding, Reproduction | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 4 concentrations including control | 0 |
| 17 | Concentration range tested | One environmentally realistic concentration included | 1 |
| 18 | Aging and biofouling | Not addressed, not possible to grow as test medium are renewed every 2 days | 0 |
| 19 | Diversity of MP tested | Only a (small) range of sizes tested | 0 |
| 20 | Exposure time | 44 days | 2 |
| Total | | | 17 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Reproductive success strongly affected | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | - | |

Paper: Qiao et al. (2019)⁷³

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average size given (5 µm) and confirmed with TEM. no indication of measurement error | 1 |
| 2 | Particle shape | Spherical shape given and confirmed with TEM | 2 |
| 3 | Polymer type | Polymer type (PS) given and confirmed with FTIR | 2 |
| 4 | Source MP | Name of provider given (Tianjin Base-Line ChromTech Research Centre) | 2 |
| 5 | Data reporting | Data reported in µg/L and particles/L | 2 |
| 6 | Chemical purity | No plasticizers detected in FTIR | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Dispersion by aeration | 1 |
| 11 | Exposure of assessment of organisms | Ingestion qualitatively assessed with polarized light microscope in the same experiment | 1 |
| 12 | Replication | Treatments not replicated | 0 |
| Subtotal | | | 12 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Suborganismal endpoints (tissue histology, enzymatic biomarkers, gut microbiome and metabolomic responses) | 0 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations including control | 0 |
| 17 | Concentration range tested | One relevant concentration included | 1 |
| 18 | Aging and biofouling | Exposure time long enough to allow for biofouling | 1 |
| 19 | Diversity of MP tested | One polymer type, one size, one shape | 0 |
| 20 | Exposure time | 21 days | 0 |
| Total | | | 16 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “Significant histological changes and enzymatic biomarkers alterations” | |
| * | Demonstrated mechanism | “MP induced inflammation (1) and oxidative stress (10) in zebrafish.” “Gut inflammation, bowel wall thinning, villi damage and epithelial damage were <i>observed</i> in the MPs-treated gut tissues.” “ <i>indicating oxidative stress</i> was caused by MP exposure. This was <i>confirmed</i> by the increased glutathione in the gut tissues. Glutathione is an important intracellular antioxidant that prevents the uncontrolled formation of free radicals and reactive oxygen species (ROS) and inhibits their reactions with DNA, proteins and lipids.” | |

Paper: Redondo-Hasselerharm et al. (2018)²

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Min/max size values and average size given (20 - 500 µm) and particle size distribution measured and given | 2 |
| 2 | Particle shape | Shape given (fragment) and shown in pictures | 2 |
| 3 | Polymer type | Polymer type given (PS) and confirmed using FTIR | 2 |
| 4 | Source MP | Source given (Axalta Coating Systems) | 2 |
| 5 | Data reporting | Data reported in g/kg of sediment (dry weight) and particles/kg of sediment (dry weight) | 2 |
| 6 | Chemical purity | To remove additives present, if any, the MP were washed with methanol three times | 2 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Well mixed in sediment | 2 |
| 11 | Exposure assessment of organisms | Ingestion quantified in the same experiment with FTIR | 2 |
| 12 | Replication | At least 3 replicates | 2 |
| <u>Subtotal</u> | | | 18 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, Growth, Feeding activity and Reproduction | 2 |
| 14 | Presence of natural (food) particles | Food and sediment added | 2 |
| 15 | Detection of effect thresholds | Effect thresholds reported with confidence intervals | 2 |
| 16 | Quality of dose-response relationship | At least 7 concentrations including control | 2 |
| 17 | Concentration range tested | One environmentally realistic concentration included | 1 |
| 18 | Aging and biofouling | Acclimatization of particles in natural sediment for 6 weeks | 1 |
| 19 | Diversity of MP tested | One polymer type, wide range of sizes and shapes | 1 |
| 20 | Exposure time | 28 days | 2 |
| <u>Total</u> | | | 31 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “Significant reduction in growth” | |
| * | Demonstrated mechanism | “Our results <i>indicate</i> that growth reduction of <i>G. pulex</i> was a sublethal effect caused by a lower ability of these organisms to assimilate food (2) due to the ingestion of PS MP, as well as by the gut blockage (1) by these particles due to a longer excretion time needed to depurate their gut.” | |
| * | Speculated mechanism | - | |

Paper: Redondo-Hasselerharm et al. (2020)⁴⁷

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Particle size distribution measured (not shown) range 20 to 516 µm, average size 227.7 ± 6.01 µm | 1 |
| 2 | Particle shape | Shape given (irregular fragments), however not confirmed | 1 |
| 3 | Polymer type | Polymer type given (PS), however not confirmed | 1 |
| 4 | Source MP | Name provider given (Axalta Coating Systems GMBH) | 2 |
| 5 | Data reporting | % plastic in sediment dry weight | 1 |
| 6 | Chemical purity | MP thoroughly washed with methanol to remove organic chemicals associated with the MP | 2 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Measurements of actual concentration at start and after 3 months and 15 months. Evidence that at least 80% of the nominal concentration throughout the test is maintained | 2 |
| 10 | Homogeneity of exposure | MP powder added to sediment and mixed with cement drill | 2 |
| 11 | Exposure of assessment | Not addressed | 0 |
| 12 | Replication | 4 replicates | 2 |
| <u>Subtotal</u> | | | <u>14</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Population abundance and diversity, individual abundance | 2 |
| 14 | Presence of natural (food) particles | OM in sediment | 2 |
| 15 | Detection of effect thresholds | Effect thresholds reported in LOEC and NOEC | 1 |
| 16 | Quality of dose-response relationship | 5 concentrations including control | 1 |
| 17 | Concentration range tested | At least 2 environmentally relevant concentrations used | 2 |
| 18 | Aging and biofouling | Exposure time of experiment is long enough for biofilm to develop, however not characterized | 1 |
| 19 | Diversity of MP tested | One polymer type, various shapes, wide range of sizes | 1 |
| 20 | Exposure time | Up to 15 months | 2 |
| <u>Total</u> | | | <u>26</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | MP adversely affected the abundance of macroinvertebrates, which was caused by a reduction in the number of Naididae | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “A reduction in food intake due to the dilution of organic matter in the sediment, together with the uptake and longer retention of MP by the Naididae worms, <i>could have caused a depletion of energy reserves (2)</i> over time, as previously found in laboratory tests for other benthic invertebrates.” | |

Paper: Rehse, Kloas and Zarfl (2016)³⁴

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Min/max size values given (1 - 4 and 90 - 106 µm) and particle size distribution confirmed with pictures | 2 |
| 2 | Particle shape | Shape given (sphere) and shown in pictures | 2 |
| 3 | Polymer type | Polymer type given (PE) but not confirmed | 1 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | Data reported in mg/L and number of particles/L | 2 |
| 6 | Chemical purity | Effect of chemicals ruled out using provider's material safety data sheet | 1 |
| 7 | Lab preparation | Beakers were loosely covered with glass petri dishes to reduce airborne contamination, use of glass materials | 1 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Homogeneity of exposure not possible (p94 Results) | 0 |
| 11 | Exposure assessment of organisms | Ingestion qualitatively assessed in the same experiment | 1 |
| 12 | Replication | 4 replicates | 2 |
| <u>Subtotal</u> | | | 14 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Immobilization and abnormal behavior or appearance (being trapped at the water surface, malformation) | 2 |
| 14 | Presence of natural (food) particles | No food added | 0 |
| 15 | Detection of effect thresholds | Effect thresholds reported with confidence intervals | 2 |
| 16 | Quality of dose-response relationship | 7 concentrations including control | 2 |
| 17 | Concentration range tested | More than 1 environmentally relevant concentration was used within the range tested. | 2 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, small range of sizes | 0 |
| 20 | Exposure time | Up to 96 h | 0 |
| <u>Total</u> | | | 22 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Immobilization | |
| * | Demonstrated mechanism | “Since we <i>can</i> exclude chemical effects by additives and pollutants attached to the MP and adherence of particles to outer structures of the daphnids, immobilization <i>can</i> be related to physical effects by ingestion (1) .” | |
| * | Speculated mechanism | - | |

Paper: Reichert et al. (2018)¹⁵⁴

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Min/max and average sizes given in the main text (37-163 µm), measured with ImageJ and particle size distribution provided in the SI | 2 |
| 2 | Particle shape | Shape given (irregular) and shown in pictures | 2 |
| 3 | Polymer type | Polymer type given (PE) but not measured | 1 |
| 4 | Source MP | Source given (Novoplastic) | 2 |
| 5 | Data reporting | Data reported in g/L and particles/L | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Additional pump added to assure suspension of plastic particles | 1 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified | 1 |
| 12 | Replication | 3 replicates | 2 |
| <u>Subtotal</u> | | | <u>13</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Health effects (bleaching and necrosis) | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | Only environmentally unrealistic concentration used | 0 |
| 18 | Aging and biofouling | Microbial biofilm to assure more realistic exposure (section 2.1) | 1 |
| 19 | Diversity of MP tested | One polymer type, various sizes, small range of sizes | 1 |
| 20 | Exposure time | 4 weeks | 2 |
| <u>Total</u> | | | <u>21</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “Negative effects on health (i.e. bleaching and tissue necrosis)” | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “Further, the latter study showed that 5.7% of the ingested particles were retained in the corals digestive system for at least 24 h, <i>potentially</i> affecting energetics (2) , pollutant toxicity (4) and trophic transfer.” Exposure to MP increase tissue necrosis and bleaching <i>might</i> be due to the high number of attached MP (5) | |

Paper: Revel et al. (2018)⁵⁶

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Reported and measured, however only a range is given 0.4-400 µm | 1 |
| 2 | Particle shape | Not reported | 0 |
| 3 | Polymer type | Polymer types given (PE, PP) and confirmed with FTIR | 2 |
| 4 | Source MP | Production in lab described, however not the origin of MP | 1 |
| 5 | Data reporting | Data reported in µg/L for water and particles/kg for sediment | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Cotton lab coat, rinsing equipment, measures taken to prevent contamination from air | 2 |
| 8 | Verification of background contamination | Background contamination MP in sediment measured visually at the start of the experiment | 1 |
| 9 | Verification of exposure | Not addressed | 0 |
| 10 | Homogeneity of exposure | Non-homogenous exposure, aggregates formed | 0 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified | 1 |
| 12 | Replication | 9 replicates | 2 |
| <u>Subtotal</u> | | | <u>12</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Cell viability, phagocytosis activity and efficiency | 0 |
| 14 | Presence of natural (food) particles | No addition of food | 0 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations including control | 0 |
| 17 | Concentration range tested | More than 2 environmentally realistic concentrations used | 2 |
| 18 | Aging and biofouling | MP were grinded to a powder | 1 |
| 19 | Diversity of MP tested | 2 polymer types, wide size range (more than 1 order of magnitude) | 1 |
| 20 | Exposure time | 10 days | 1 |
| <u>Total</u> | | | <u>17</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Decrease in coelomocytes viability, no alteration of phagocytosis activity, phenoloxydase and acid phosphatase. The exposure to PE and PP MP only induced a slight, but significant alteration in cell viability and a tendency to a decrease in PO and AcP enzymes which play a role in the immune system. | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “PO and AcP activity which play an important role in the immune system, had a slight tendency to decrease. This indicates that exposure to relevant MP <i>could</i> alter cellular integrity (6) and increase susceptibility of worms to environmental stress.” | |

Paper: Revel et al. (2019)⁵⁵

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Particle size distribution measured with a Beckman Coulter, min/max size values and average size with standard deviation provided (0.4 - 950 µm) | 2 |
| 2 | Particle shape | Not reported | 0 |
| 3 | Polymer type | Polymer types given (PE and PP) but not confirmed | 1 |
| 4 | Source MP | Production in lab described, however not the origin of MP | 1 |
| 5 | Data reporting | Data reported in µg/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Thorough rinsing, measures taken to avoid contamination from air, Cotton lab coat | 2 |
| 8 | Verification of background contamination | Blanks performed and measured with µFT-IR | 2 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Possible formation of agglomerates | 0 |
| 11 | Exposure assessment of organisms | Ingestion quantitatively assessed with µFT-IR | 2 |
| 12 | Replication | 5 replicates | 2 |
| <u>Subtotal</u> | | | <u>13</u> |
| <i>Applicable for risk assessment</i> | | | |
| 13 | Endpoints | Condition index, Clearance rate and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 4 concentrations including control | 0 |
| 17 | Concentration range tested | More than 2 realistic concentrations used | 2 |
| 18 | Aging and biofouling | Not mentioned | 0 |
| 19 | Diversity of MP tested | 2 polymer types, wide range of sizes (more than 1 order of magnitude) | 1 |
| 20 | Exposure time | 10 days | 1 |
| <u>Total</u> | | | <u>21</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Increase in SOD, CAT and phosphatase activities. No effect on clearance rate and histopathological parameters. | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | Significant increases in SOD and CAT activities <i>could</i> be indicative of a change in the redox balance (8) . “This <i>could potentially</i> lead to induction of oxidative stress (10) in digestive glands. However, the degree of responses remained low and longer-term studies should be conducted to clarify the effect of MP on oxidative stress and the influence of MP size and aggregation status.” “Induction of oxidative stress could also have been induced by additives (4) since we used commercial plastics in order to evaluate the toxicity of environmentally relevant MP.” | |

Paper: Revel et al. (2020)¹⁵⁵

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Particle size distribution measured with a Beckman Coulter, min/max size values (0.4-500 µm) and average size with standard deviation provided. | 1 |
| 2 | Particle shape | Shape given (fragment) but not shown | 1 |
| 3 | Polymer type | Polymer types given (PE and PP) but not confirmed | 1 |
| 4 | Source MP | Production of MP in own lab, however the origin of MP not provided | 1 |
| 5 | Data reporting | Data reported in mg/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Thorough rinsing, measures taken to avoid contamination from air, Cotton lab coat | 2 |
| 8 | Verification of background contamination | Not performed | 0 |
| 9 | Verification of exposure | Microscopic counting only in stock solutions | 1 |
| 10 | Homogeneity of exposure | Possible formation of agglomerates | 0 |
| 11 | Exposure of assessment | Ingestion quantitatively assessed with µFT-IR | 2 |
| 12 | Replication | 3 replicates | 2 |
| <u>Subtotal</u> | | | <u>12</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Clearance rate, tissue alteration, antioxidant defense, immune alteration and DNA damage | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 4 concentrations including control | 0 |
| 17 | Concentration range tested | Authors state that environmentally realistic concentrations were used and compare to MEC | 2 |
| 18 | Aging and biofouling | Not mentioned | 0 |
| 19 | Diversity of MP tested | 2 types of MP, various shapes, wide size range | 1 |
| 20 | Exposure time | 10 days | 1 |
| <u>Total</u> | | | <u>20</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects were found | |

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Average size given (20 µm) and confirmed with optical microscope (OM) and DLS | 2 |
| 2 | Particle shape | Shape (sphere) given and shown in pictures under OM | 2 |
| 3 | Polymer type | Polymer type given (PS) and identified using a FTIR | 2 |
| 4 | Source MP | Source given (Sigma-Aldrich) | 2 |
| 5 | Data reporting | Data reported in mg/L and particles/ml | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Avoidance of plastic mentioned | 1 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | No homogenization reached; particles ended at the bottom. | 0 |
| 11 | Exposure assessment of organisms | Ingestion qualitatively assessed in the same experiment | 1 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 14 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Condition index and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | No addition of natural food or sediment | 0 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | One environmentally relevant concentration tested. Comparison to MEC is hampered by the lack of info on MP <50 µm, however authors reason that one order of magnitude higher is a good compromise. Reference is given | 1 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 14 days | 1 |
| Total | | | 18 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | “MP induce effects on antioxidant capacity, DNA damage, neurotoxicity and oxidative damage.” | |
| * | Demonstrated mechanism | “Tissue-specific sensibility is involved in the clam’s response to PS exposure by <i>inducing oxidative stress (10)</i> , with the gills providing a more effective response than digestive gland.” | |
| * | Speculated mechanisms | “The present results indicated that after a week of depuration, MP were still present in both tissues.” “The presence of small aggregates of MP in the haemolymph indicates that PS MP were transported into the circulatory system, indicating a possible translocation, where they can be retained for several weeks and then transported to several tissues (1) where <i>they can cause harm</i> .” “ <i>The mechanism of genotoxicity of PS MP remains unknown</i> , but it is suggested that it can be related to ROS production and oxidative stress , not handled by the antioxidant defense mechanism, as occurs with nanoparticles.” “The accumulation of MP in this tissue might impair the digestive system (2) with a consequent decrease of feeding behavior .” | |

Paper: Rist, Baun and Hartmann (2017)⁷⁰

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Average size measured and given (6 µm) with standard deviation | 2 |
| 2 | Particle shape | Shape given (sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Phosphorex) | 2 |
| 5 | Data reporting | Data reported in mg/L and particles/mL | 2 |
| 6 | Chemical purity | No measures taken for Tween 20 and sodium azide | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Not mentioned, only stock suspension vortexed | 0 |
| 11 | Exposure assessment of organisms | Ingestion quantified in separate experiment | 1 |
| 12 | Replication | 10 replicates | 2 |
| <u>Subtotal</u> | | | <u>11</u> |
| <i>Applicable for risk assessment</i> | | | |
| 13 | Endpoints | Mortality, reproduction, growth, feeding rate | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 4 concentrations including control | 0 |
| 17 | Concentration range tested | Only unrealistic concentrations used | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 21 days | 2 |
| <u>Total</u> | | | <u>18</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects found | |

Paper: Romano et al. (2018)¹⁵⁷

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|--|--|--|------------------|
| 1 | Particle size | Size distribution measured with a Mastersizer but only D10 (40 µm), D50 (140 µm) and D90 (310 µm) reported without error | 1 |
| 2 | Particle shape | Shape given (fragment) and shown in pictures | 2 |
| 3 | Polymer type | Polymer type given (PVC) but not confirmed | 1 |
| 4 | Source MP | Source given (Toxemerger) | 2 |
| 5 | Data reporting | Data reported in mg/L | 1 |
| 6 | Chemical purity | PVC fragments analyzed for PAHs, PCBs, phthalates, arsenic, lead, cadmium and mercury and below detection limit, additionally a solvent control was used | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Ethanol was used to help evenly distribute the fragments in water, however not measured/characterized | 1 |
| 11 | Exposure assessment of organisms | Ingestion qualitatively assessed | 1 |
| 12 | Replication | 3 replicates | 2 |
| <u>Subtotal</u> | | | <u>12</u> |
| <i>Applicable for risk assessment</i> | | | |
| 13 | Endpoints | Suborganismal endpoints | 0 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 4 concentrations, including control | 0 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | Not mentioned | 0 |
| 19 | Diversity of MP tested | One polymer type, various shapes, small range of sizes | 1 |
| 20 | Exposure time | 96 hours | 0 |
| <u>Total</u> | | | <u>14</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Increase in whole body trypsin and chymotrypsin activities. Localized thickening of the mucosal epithelium. No tissue damage. | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “The increased whole-body trypsin and chymotrypsin activities <i>may</i> indicate an attempt to enhance digestion (2) to compensate for epithelial thickening of the intestine and/or to digest the plastics.” “Other potential explanation <i>may</i> have been a compensatory secretory response to improve digestion and absorption. Beside intestinal thickening, the trypsin and chymotrypsin activities were higher in the fish exposed to MP. This seems to support an attempt by the intestine to digest the consumed fragments.” | |

Paper: Seoane et al. (2019)⁷⁹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Nominal size (2 µm) given and confirmed with DLS (2274.0 ± 100.9 nm). | 2 |
| 2 | Particle shape | Spherical shape given and shown in pictures | 2 |
| 3 | Polymer type | Polymer type (PS) given but not confirmed | 1 |
| 4 | Source MP | Name of provider given (Micromod) | 2 |
| 5 | Data reporting | Data reported in µg/ml and particles/ml | 2 |
| 6 | Chemical purity | Presence of chemicals not ruled out | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of background contamination | 0 |
| 9 | Verification of exposure | Monitored using SEM pictures every 24 h, however no evidence that at least 80% of the nominal concentration is maintained. | 1 |
| 10 | Homogeneity of exposure | Not mentioned, aggregates formed | 0 |
| 11 | Exposure of assessment of organisms | Hetero-aggregates between MP and bacteria shown in SEM pictures | 2 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 14 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Growth and photosynthetic efficiency and suborganismal endpoints (cell morphology, autofluorescence, esterase activity, reactive oxygen species (ROS) levels, cytoplasmic membrane potential and neutral lipid content) | 2 |
| 14 | Presence of natural (food) particles | NA | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | Close to worst case scenario concentrations | 0 |
| 18 | Aging and biofouling | Not addressed | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size per experiment | 0 |
| 20 | Exposure time | 72 h | 2 |
| Total | | | 20 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Slight decrease in growth rate. Significant decrease in the esterase activity and the lipid reserves of MP-exposed cells. | |
| * | Demonstrated mechanism | “MP-exposed cells modulate their energy metabolism (9) to properly acclimate to the stress conditions. ” Contact, even temporal <i>could</i> be detected and considered as a stress (6) by the cells and may be translated into biochemical signals, triggering a response to deal with. It <i>could</i> be described as the “billiard ball effect”. As documented in humans, cells <i>may</i> sense mechanical cues, although the details underlying how cells respond to mechanical forces are <i>not well understood yet</i> . The potential release of chemicals (4) from MP could be another explanation to the indirect toxicity detected.” “In the present study, oxidative stress was not detected in MP-exposed cells; therefore, the decrease in lipids observed cannot be associated with its oxidation, but to modulation of energy metabolism to properly acclimate to the stress conditions, maintaining a healthy status.” | |
| * | Speculated mechanism | | |

Paper: Silva et al. (2019)⁵⁴

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Min/max and average sizes values given (32–63 µm, 63–250 µm and 125–500 µm) and PSD done by sieving. < 10 bins | 1 |
| 2 | Particle shape | powder/Irregularly shape given and shown in pictures with OM | 2 |
| 3 | Polymer type | Polymer type (PE) given but not confirmed | 1 |
| 4 | Source MP | Name of provider given (Sigma-Aldrich and Goodfellow) | 2 |
| 5 | Data reporting | Data reported in g/kg | 1 |
| 6 | Chemical purity | Acid- and milli-Q washed, but not with organic solvents | 0 |
| 7 | Lab preparation | Equipment washed, rinsed. Samples & filters covered. Plastic avoided. | 2 |
| 8 | Verification of background contamination | Blanks and controls were applied and subtracted from samples. MP measured visually | 1 |
| 9 | Verification of exposure | Checked in water and sediment in two doses at the start of experiment | 1 |
| 10 | Homogeneity of exposure | Directly mixed in sediment (which is sufficient as in bed sediment, no settling occurs) | 2 |
| 11 | Exposure of assessment of organisms | Ingestion quantitatively assessed with stereoscope | 1 |
| 12 | Replication | 13 replicates | 2 |
| Subtotal | | | 16 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Growth, Emergence | 2 |
| 14 | Presence of natural (food) particles | Food and sediment (without OM) added | 2 |
| 15 | Detection of effect thresholds | LOEC provided, and EmT50 with error | 2 |
| 16 | Quality of dose-response relationship | 6 concentrations including control | 2 |
| 17 | Concentration range tested | Realistic based on literature | 2 |
| 18 | Aging and biofouling | Exposure time long enough to allow for biofouling | 1 |
| 19 | Diversity of MP tested | Smaller range of sizes, wide range of shapes | 1 |
| 20 | Exposure time | 28-days partial life cycle assays | 2 |
| Total | | | 30 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Reduction larva's growth and delay on imagoes emergence | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “Present study points that the ingestion and persistence of smaller-sized PE particles in the gut could <i>likely</i> cause a significant reduction in the ingestion of organic items and interfering in food processing (2) .” | |

Paper: Sjollema et al. (2016)³⁵

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Average size given (6 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape given (sphere) but not confirmed | 1 |
| 3 | Polymer type | Polymer type given (PS) but not checked | 1 |
| 4 | Source MP | Source given (Polyscience) | 2 |
| 5 | Data reporting | Data reported in mg/L and particles/L | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Actual concentration measured with a flow cytometer but only for the large MP at the end of the experiment | 1 |
| 10 | Homogeneity of exposure | Test vials were gently mixed on a shaker achieving a homogenous microbead distribution. Adequate homogenization was confirmed previously using fluorescent microbeads in the same size range. | 1 |
| 11 | Exposure assessment of organisms | Not assessed | 0 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 11 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Photosynthetic efficiency and growth | 2 |
| 14 | Presence of natural (food) particles | Not applicable for algae | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations including control | 0 |
| 17 | Concentration range tested | No environmentally realistic concentration tested | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 72 h | 1 |
| Total | | | 16 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effect on the photosynthetic efficiency or growth | |

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Min/max size values given (32-63 m for PMMA and PHB), and confirmed with pictures | 2 |
| 2 | Particle shape | Shape not mentioned in the text but shown in pictures | 2 |
| 3 | Polymer type | Polymer types given (PMMA, PHB) but not confirmed | 1 |
| 4 | Source MP | Name of provider not given in the main text | 0 |
| 5 | Data reporting | Data reported as MP per individual | 0 |
| 6 | Chemical purity | Not mentioned, addition of a SpanTween-surfactant | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Equal distribution onto the bottom of the beaker (not in the whole system) | 0 |
| 11 | Exposure assessment of organisms | Ingestion quantified in separate experiment | 1 |
| 12 | Replication | 24 replicates | 2 |
| Subtotal | | | 8 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Feeding rate, assimilation efficiency, and wet weight change | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | No environmentally realistic concentration tested | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, small range of sizes | 1 |
| 20 | Exposure time | 28 days | 2 |
| Total | | | 16 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Decrease of the assimilation efficiency, lower wet weight gain | |
| * | Speculated mechanism | “The physical presence of the MP in the digestive tract generally leads to a reduced uptake of food (2) , which in turn lowers energy intake with subsequent physiological effects.” “A longer gut retention time, <i>possibly</i> leading to gut obstruction (1) , <i>might</i> explain the negative effects on weight gain relative to the particle-free control.” “The presence of MP in the gut, however, had more effects on WWch than the presence of natural silica particles, if we assume the latter were ingested.” | |
| * | Demonstrated mechanism | - | |

Paper: Sun et al. (2018)¹⁵⁹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Average diameter size given (1 µm) and measured with DLS (1626 ± 362 nm) | 2 |
| 2 | Particle shape | Shape (beads) given and confirmed with TEM images Fig.S1 | 2 |
| 3 | Polymer type | Polymer type given (PS) and shown in the bacteria | 2 |
| 4 | Source MP | Source given (Sigma Aldrich) | 2 |
| 5 | Data reporting | Data provided in mg/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of background contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Homogeneity of stock solutions using sonication, not shown | 0 |
| 11 | Exposure of assessment of organisms | Interaction bacteria and MP shown in separate experiment | 1 |
| 12 | Replication | 3 replicates | 2 |
| <u>Subtotal</u> | | | <u>12</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Cell viability and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | No natural particles present | 0 |
| 15 | Detection of effect thresholds | Effect thresholds not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 6 concentrations including control for cell viability, 2 concentrations including control for intracellular ROS generation | 2 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 2 h | 0 |
| <u>Total</u> | | | <u>16</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Inhibition of growth | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “The growth promotion of the bacteria cells <i>might</i> be attributed to hermetic responses, which was due to the defensive and adaptive responses of cells to stress (6). ” | |

Paper: Sussarellu et al. (2016)²⁸

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Average sizes given (2 and 6 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape given (sphere) but not confirmed | 1 |
| 3 | Polymer type | Polymer type given (PS) but not checked | 1 |
| 4 | Source MP | Source given (Polyscience) | 2 |
| 5 | Data reporting | Data reported in particles/ml and mg/L | 2 |
| 6 | Chemical purity | Full analysis of chemicals in MP done. Effect by chemicals ruled out and control used for Tween-20 | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Daily verification of the exposure with a flow cytometer | 2 |
| 10 | Homogeneity of exposure | Water inflow was pressurized, and air bubbling was used. MP were supplied to tanks with Tween-20 | 1 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified in the same experiment | 1 |
| 12 | Replication | 3 replicates | 2 |
| | | <u>Subtotal</u> | <u>14</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Feeding, Fecundity, Offspring development and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | One environmentally relevant concentration tested | 1 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, small range of sizes | 0 |
| 20 | Exposure time | Up to 8 weeks | 2 |
| | | <u>Total</u> | <u>22</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Increased consumption and absorption efficiency, disturbance in energetics | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “Consumption of microalgae and absorption efficiency appeared significantly higher in exposed oysters, suggesting a compensatory effect on food intake (2) and absorption efficiency and an enhancement of mechanical digestion.” “A disturbance in individual energetics revealed by DEB modeling <i>suggested</i> that micro-PS particles have threatened the physiological integrity (8) of oysters and consequently increased the maintenance costs , as described in response to various stresses and species (45–47). MP <i>may potentially</i> act as endocrine disruptors (4) . The chemical analyses of virgin micro-PS only revealed bibenzyl and 1(2H) naphthalenone,3,4, dihydro4phenyl in destructive conditions after dichloromethane extraction. Bibenzyl-diol core molecules may have endocrine disruption properties, as established in mammal cells, because they are structural analogs of estrogens.” | |

Paper: Tang et al. (2018)⁷⁴

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average size given (1 µm) but not measured | 1 |
| 2 | Particle shape | Not reported | 0 |
| 3 | Polymer type | Polymer type given (PS) but not measured | 1 |
| 4 | Source MP | Source given (Saierqun) | 2 |
| 5 | Data reporting | Data reported in mg/L and particles/L | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Description of method to obtain homogenous exposure | 1 |
| 11 | Exposure assessment of organisms | Not addressed | 0 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 9 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Symbiont density and chlorophyll content | 2 |
| 14 | Presence of natural (food) particles | No addition of food | 0 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | Only unrealistic concentration used | 0 |
| 18 | Aging and biofouling | Not mentioned | 0 |
| 19 | Diversity of MP tested | One polymer type, one size | 0 |
| 20 | Exposure time | Up to 24 h | 0 |
| Total | | | 11 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effect on symbiotic relationship. Chlorophyll content increased. Increase in antioxidant enzymes. Decrease in detoxifying and immune enzyme. Up-regulated coral genes related to stress response. Down-regulated genes involved in sterol transport and EGF-ERK1/2 pathway. | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “These results <i>suggest</i> that acute exposure of MP can activate the stress response (6) of the scleractinian coral <i>P. damicornis</i> and repress its detoxification and immune system (8) through the JNK and ERK signal pathways.” | |

Paper: Van Cauwenberghe, Claessens, et al. (2015)¹⁶⁰

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Average size values of the 3 MP present in the mixture given (10, 30, 90 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape given (sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Coulter Standard latex beads, Analis) | 2 |
| 5 | Data reporting | Data reported in particles/ml | 1 |
| 6 | Chemical purity | Not assessed | 0 |
| 7 | Lab preparation | Not mentioned for effect assessment | 0 |
| 8 | Verification of background contamination | No verification of background contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Use of a magnetic stirrer | 1 |
| 11 | Exposure assessment of organisms | Quantitatively assessed in separate experiment | 1 |
| 12 | Replication | At least 5 replicates | 2 |
| <u>Subtotal</u> | | | <u>10</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Only suborganismal endpoints assessed cannot unambiguously be linked to higher level | 0 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | No environmentally realistic concentration used | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, small range of sizes | 0 |
| 20 | Exposure time | 14 days | 1 |
| <u>Total</u> | | | <u>13</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects found | |

Paper: Von Moos, Burkhardt-Holm and Köhler (2012)¹¹⁴

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Min/max sizes given (0 - 80 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape mentioned (nonuniform) but not shown | 1 |
| 3 | Polymer type | Polymer type given (HDPE) but not confirmed | 1 |
| 4 | Source MP | Source given (Abifor Zürich) | 2 |
| 5 | Data reporting | Data reported in g/L | 1 |
| 6 | Chemical purity | Presence of chemical additives ruled out by referencing the material data sheet from provider | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of background contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Exposure beakers were equipped with aeration stones to attempt an even distribution of MP in suspension | 1 |
| 11 | Exposure assessment of organisms | Ingestion quantified in the same experiment | 2 |
| 12 | Replication | 3 replicates | 2 |
| | | Subtotal | 12 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Condition index and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Food added (half a drop of LiquifryMarine per beaker) only after 48 h | 1 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations tested including control | 0 |
| 17 | Concentration range tested | No environmentally realistic concentration used | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, wide range of sizes (more than 1 order of magnitude) | 1 |
| 20 | Exposure time | Up to 96 h | 0 |
| | | Total | 16 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Histological changes and strong inflammatory response | |
| * | Demonstrated mechanism | “In the digestive gland (1) MP caused the formation of granulocytomas (after 3 h) and a steady decrease in lysosomal stability (after 6 h).” “The formation of granulocytomas is a non-neoplastic inflammatory cellular response (1) .” “We provide proof of principle that MP are taken up into cells and cause significant effects on the tissue and cellular level .” | |

Paper: Vroom et al. (2017)¹⁰⁹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average size given for beads (15 µm) but not confirmed. Size distribution of PS fragments (<30µm) given and confirmed with photograph and LAS software, however not used for effect study (figure S1 SI) | 1 |
| 2 | Particle shape | Shape given (bead) and shown in pictures | 2 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Phosphorex) | 2 |
| 5 | Data reporting | Data reported in particles/ml | 1 |
| 6 | Chemical purity | Effects of solvent and styrene monomers ruled out using references. | 1 |
| 7 | Lab preparation | Filtered seawater with 1 µm filter | 1 |
| 8 | Verification of background contamination | No verification of background contamination | 0 |
| 9 | Verification of exposure | MP were quantified using stereoscope software LAS | 1 |
| 10 | Homogeneity of exposure | Not assessed | 0 |
| 11 | Exposure assessment of organisms | Ingestion qualitatively assessed in the same experiment; Ingestion quantified in separate experiment | 1 |
| 12 | Replication | At least 53 replicates | 2 |
| Subtotal | | | 13 |
| <i>Applicable for risk assessment</i> | | | |
| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
| 13 | Endpoints | Survival | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations including control | 0 |
| 17 | Concentration range tested | More than 1 environmentally realistic concentration included | 2 |
| 18 | Aging and biofouling | Not clear if aged MP were used in the effect assessment. PS granules were pulverized to PS fragments, however not used in effect study | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 11 days | 1 |
| Total | | | 20 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects found | |

Paper: Wan et al. (2019)¹⁶¹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|------------------|
| 1 | Particle size | Average size given (5 and 50 µm) and confirmed with SEM, no indication of error measurement | 1 |
| 2 | Particle shape | Spherical shape given and confirmed with SEM | 2 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Sources given (Microsphere-Nanospheres and Phosphorex) | 2 |
| 5 | Data reporting | Data reported in µg/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Not explicitly mentioned, Sonication prior to exposure | 1 |
| 11 | Exposure of assessment of organisms | Ingestion qualitatively assessed with laser scanning microscope in separate experiment | 1 |
| 12 | Replication | 5 replicates | 2 |
| | | <u>Subtotal</u> | <u>11</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Suborganismal endpoints that cannot be unambiguously linked to a threat at the individual or population level | 0 |
| 14 | Presence of natural (food) particles | No feeding during exposure and no additional particles used | 0 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations including control | 0 |
| 17 | Concentration range tested | Not compared with MEC | 0 |
| 18 | Aging and biofouling | Not addressed | 0 |
| 19 | Diversity of MP tested | One polymer type, two sizes, but separate experiment, one shape | 0 |
| 20 | Exposure time | 7 days | 1 |
| | | <u>Total</u> | <u>12</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Alterations in the microbiome at the phylum and genus levels in larval zebrafish, including changes in abundance and diversity of the microbiome. Changes in glycolysis-related genes and lipid metabolism-related genes, confirming that polystyrene MP disturbed glycolipid and energy metabolism. | |
| * | Demonstrated mechanism | “ Alterations in the microbiome (7) at the phylum and genus levels in larval zebrafish, confirming that PS MP disturbed glycolipid and energy metabolism (8) .” | |
| * | Speculated mechanism | - | |

Paper: Wang et al. (2019)³²

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Particle size distribution (S3,4) given (10-11 µm) and confirmed with coulter counter | 2 |
| 2 | Particle shape | Spherical shape given and confirmed with images of by microscope software. see SI | 2 |
| 3 | Polymer type | Polymer type given and confirmed with FTIR | 2 |
| 4 | Source MP | Name of provider given (Thermo Fisher Scientific Corporation, USA) | 2 |
| 5 | Data reporting | Data reported in particles/ml and µg/L | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | In exposure solutions, possible changes in the concentration of prepared solutions were measured over four time points. Evidence that at least 80% of the nominal concentration is maintained. See SI. | 2 |
| 10 | Homogeneity of exposure | Sonication before use, however not during exposure | 1 |
| 11 | Exposure of assessment of organisms | Ingestion assessed qualitatively | 1 |
| 12 | Replication | 4 replicates | 2 |
| Subtotal | | | 16 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, growth, development and epithelial cells lining the digestive tract | 2 |
| 14 | Presence of natural (food) particles | Food added in the chronic tests (14 d), not in the acute tests (24 h) | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | At least 6 concentrations including control | 2 |
| 17 | Concentration range tested | More than one environmentally realistic concentration used | 2 |
| 18 | Aging and biofouling | Not addressed | 0 |
| 19 | Diversity of MP tested | One polymer type, one size, one shape | 0 |
| 20 | Exposure time | 24 h epithelial cells lining the digestive tract, and 14 d survival growth, development and survival | 1 |
| Total | | | 25 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects on survival, growth and development. Abnormal ultrastructures of intestinal epithelial cells were observed including fewer and disordered microvilli, an increased number of mitochondrion and the appearance of autophagosome | |
| * | Demonstrated mechanism | “ Abnormal ultrastructures of intestinal epithelial cells (1) were observed including fewer and disordered microvilli (1) , an increased number of mitochondrion and the appearance of autophagosome.” | |
| * | Speculated mechanism | “These phenomena <i>could affect nutrition absorption (2) and energy metabolism.</i> ” | |

Paper: Watts et al. (2016)¹⁶²

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Average size given (8 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape given (sphere) but not confirmed | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | Source given (Spherotec) | 2 |
| 5 | Data reporting | Data reported in particles/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of background contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Not assessed | 0 |
| 11 | Exposure assessment of organisms | Ingestion qualitatively assessed in a separate experiment | 1 |
| 12 | Replication | 10 replicates | 2 |
| Subtotal | | | 9 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Mortality (not directly assessed) and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Absence of food | 0 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations including control | 0 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | Up to 24 h | 0 |
| Total | | | 11 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Small effect on oxygen consumption, decrease in hemolymph sodium ions and increase in calcium ions | |
| * | Demonstrated mechanism | “The physiological consequences to the crabs, under short-term exposure, were minimal .” “Evidently, crabs <i>are able</i> to overcome these minor effects on ion exchange induced by exposure to the polystyrene microspheres used here by minor physiological regulation (9) .” | |
| * | Speculated mechanism | - | |

Paper: Weber et al. (2018)²³

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Min/max size values given (10-150 µm) and particle size distribution measured in the stock suspension and provided | 2 |
| 2 | Particle shape | Shape mentioned (fragment) and shown in SEM pictures | 2 |
| 3 | Polymer type | Polymer type given (PET) and confirmed using FTIR | 2 |
| 4 | Source MP | Self-made MP, source of soft drink bottle not mentioned. Not enough information for reproducibility | 1 |
| 5 | Data reporting | Data reported in particles/ml | 1 |
| 6 | Chemical purity | Used solvent control, however plastics were not checked | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Verification of the exposure done at the start of the experiment using different methods depending on concentration | 1 |
| 10 | Homogeneity of exposure | Homogenization of the exposure reached with the use of cetyl alcohol | 1 |
| 11 | Exposure assessment of organisms | Ingestion quantitatively assessed in separate experiment | 1 |
| 12 | Replication | 10 replicates | 2 |
| Subtotal | | | 13 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, development (molting), feeding activity and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 6 concentrations including control | 2 |
| 17 | Concentration range tested | One environmentally realistic concentration included | 1 |
| 18 | Aging, weathering and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, variety of shapes, wide size range (more than 1 order of magnitude) | 1 |
| 20 | Exposure time | 48 days | 2 |
| Total | | | 24 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | No effects found. | |

Paper: Welden and Cowie (2016)⁵⁷

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Min/max size values given (3-5 mm) but not measured | 1 |
| 2 | Particle shape | Shape given (fiber) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PP) but not confirmed | 1 |
| 4 | Source MP | Source given (Gaelforce) | 2 |
| 5 | Data reporting | Data reported in number of fibers, but no concentration given | 0 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | A closed 68 L header tank was used to circulate filtered seawater to prevent introduction of foreign MP. | 1 |
| 8 | Verification of background contamination | Contamination checked during the exposure assessment | 1 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure assessment of organisms | MP retention visually quantified | 1 |
| 12 | Replication | No replicates | 0 |
| <u>Subtotal</u> | | | <u>8</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Feeding rate and body mass | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | No comparison made with MEC | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, small size range | 0 |
| 20 | Exposure time | 8 months | 2 |
| <u>Total</u> | | | <u>15</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Reduction in mean body mass | |
| * | Demonstrated mechanism | “MP aggregations reduce the nutritional health (2) of <i>N. norvegicus</i> . The reduction in mean body mass of the MP-fed individuals indicates that retention of MP results in lower growth rates.” | |
| * | Speculated mechanism | - | |

Paper: Wen et al. (2018)¹⁶³

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Min/max size values given (70-88 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PE) but not measured | 1 |
| 4 | Source MP | Source given (BaseLine ChromTech Research Centre) | 2 |
| 5 | Data reporting | Data reported in µg/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | No significant aggregation observed with DLS. | 2 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified | 1 |
| 12 | Replication | 3 replicates | 2 |
| | | Subtotal | 11 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, growth, predatory performance and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, small range of sizes | 0 |
| 20 | Exposure time | 30 days | 1 |
| | | Total | 17 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Decreased in AChE and ALP activity and increase in LDH and COX activity. No effects on survival or growth. Decrease in post-exposure predatory performance. Decrease in AChE activity indicating adverse effects in nervous and neuromuscular function. Decrease in Trypsin activity. Decrease in ALP activity. Nevertheless, juvenile survival and growth were minimally impacted, and thus, <i>S. aequifasciatus</i> could cope with near-future temperature increases and MP exposure. | |
| * | Demonstrated mechanism | “Results indicate deficits in the digestive capabilities (2) of early-stage <i>S. aequifasciatus</i> .” | |
| * | Speculated mechanism | - | |

Paper: Wright et al. (2013)¹²

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Average size given (130 µm) and confirmed with SEM pictures and a Mastersizer but error not provided | 2 |
| 2 | Particle shape | Shape mentioned in the SI (irregular) and shown in SEM pictures | 2 |
| 3 | Polymer type | Polymer type given (UPVC) but not confirmed | 1 |
| 4 | Source MP | Source given (Goodfellow Cambridge) | 2 |
| 5 | Data reporting | Data reported in % plastic in sediment (dry weight) | 1 |
| 6 | Chemical purity | Analysis of chemicals done | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | All sediment mixtures were homogenized for five minutes using an electric paint mixer | 2 |
| 11 | Exposure assessment of organisms | Not assessed | 0 |
| 12 | Replication | 11 replicates | 2 |
| Subtotal | | | 13 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Feeding activity, immunity, energy reserves and gut residence time | 2 |
| 14 | Presence of natural (food) particles | Addition of food (natural sediments with OM) | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 4 concentrations including control | 0 |
| 17 | Concentration range tested | MEC mentioned of 3% by weight, included 2 environmental concentrations | 2 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, irregular shapes, a small range of sizes | 1 |
| 20 | Exposure time | Acute (24 h) and chronic exposure (28 days) | 2 |
| Total | | | 23 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Depleted energy reserves | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | <p>“Our results <i>suggest</i> that depleted energy reserves arise from a combination of reduced feeding activity (2), longer gut residence times of ingested material and inflammation (1).” “In our experiments, depleted energy reserves, which closely followed the trend for lipid reserves, <i>could compromise</i> somatic maintenance and growth, maturity and reproduction.” “Prolonged gut residence times <i>imply</i> that MP, which are of low nutritional value (2), are being retained and subjected to extensive digestion (2), at an energetic cost.”</p> | |

Paper: Wu et al. (2019)⁹⁵

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | D ₁₀ , D ₅₀ , D ₉₀ (111, 157, 216 µm PVC) (64, 172, 236 µm PP) values reported and PSD measured with DLS. Less than 10 bins | 1 |
| 2 | Particle shape | Not given, but they say they measured it with laser particle size analyzer. Unclear what shape is tested | 0 |
| 3 | Polymer type | Polymer type (PP and PVC) given but not confirmed | 1 |
| 4 | Source MP | Name of provider given (Aladdin Industrial Corporation) | 2 |
| 5 | Data reporting | Dara reported in mg/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure of assessment of organisms | Not addressed | 0 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 7 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Chlorophyll a content and photosynthetic activity | 2 |
| 14 | Presence of natural (food) particles | Food (nutrients) added | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 7 concentrations including control | 2 |
| 17 | Concentration range tested | Authors state that they covered and exceeded actual environmental concentrations. No comparison with MEC. | 0 |
| 18 | Aging and biofouling | Not addressed | 0 |
| 19 | Diversity of MP tested | Only a small range of sizes used, two types of MP in separate experiments | 0 |
| 20 | Exposure time | 7 to 11 days | 2 |
| Total | | | 15 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Reduced chlorophyll a content. Decrease and subsequently an increase of photosynthetic activity | |
| * | Demonstrated mechanism | “Thus, the algae <i>were</i> in an oxidative stress (10) state.” | |
| * | Speculated mechanism | “There <i>may</i> be two reasons for the decrease in chlorophyll a content: first, the accumulation of intracellular reactive oxygen species damages the cell structure and blocks chlorophyll synthesis; second, MP have a large surface area and strong adsorption ability, and the microalgae and MP would form hetero-aggregates of different sizes, and make the microalgae inactive (5) .” “Fv/Fm and Fv/F0 ratios were relatively stable under normal conditions and were not susceptible to species and growth conditions, but when their values decrease, they indicate that plants are inhibited by light [49]. This <i>may</i> be ascribed to MP that interrupted the photosynthetic electron transport (5) between q _a and q _b , thus forming more q _b -non-reducing PSII reaction centers, which resulted in the reduction of PSII reaction center oxygen evolution.” | |

| Paper: Yin et al. (2018)¹⁶⁴ | | | |
|---|--|--|--------------|
| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
| 1 | Particle size | PSD measured but average size provided (15 µm) without error | 1 |
| 2 | Particle shape | Shape given (bead) and shown in pictures | 2 |
| 3 | Polymer type | Polymer type given (PS) but not measured | 1 |
| 4 | Source MP | Source given (Tianjin Unibead Scientific Co) | 2 |
| 5 | Data reporting | Data reported in microspheres/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | MP suspended in water were not homogenous | 0 |
| 11 | Exposure assessment of organisms | Ingestion qualitatively assessed | 1 |
| 12 | Replication | 5 replicates | 2 |
| | | Subtotal | 10 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Feeding activity, Growth, Swimming speed and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentration including control | 0 |
| 17 | Concentration range tested | MEC given in wrong unit, not well motivated | 0 |
| 18 | Aging and biofouling | Not mentioned | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 14 days | 0 |
| | | Total | 14 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Increased foraging time and reduced swimming speed. Lower growth, protein and lipid content. Injury to bile and liver. | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | <p>“The accumulation of MP in the gastrointestinal tract stimulated the intestinal tract and affected digestive function, reducing food (2) intake through changes in behavior and appetite. Respiratory stress and lesions caused by MP may be responsible for changes in behavior. MP adhered to gills and skin may cause a small, but transient change in oxygen consumption and ion regulation. Oxidative stress (10) in fish was induced by PS-MP.” “Liver in PS MP group did appear obvious hyperaemia in the present work, <i>indicating</i> that MP induced stress (6) in liver, thereby <i>possibly</i> impacting the function of liver. MP were stored in the intestine, <i>potentially irritating</i> to the gastrointestinal lining (1). Such changes in the morphology of the intestine <i>may affect absorption</i> of nutrients and further impair fish growth.”</p> | |

Paper: Yin et al. (2019)¹⁶⁵

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average size given (15 µm) and confirmed with SEM. No indication of measurement error given | 1 |
| 2 | Particle shape | Spherical shape (bead) given and confirmed with SEM | 2 |
| 3 | Polymer type | Polymer (PS) type given and confirmed with FTIR | 2 |
| 4 | Source MP | Name of provider given (Tianjin Unibead Scientific Co. Ltd) | 2 |
| 5 | Data reporting | Data provided in µg/L and particles/L | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | No verification of exposure | 0 |
| 10 | Homogeneity of exposure | Not mentioned | 0 |
| 11 | Exposure of assessment of organisms | Not assessed | 0 |
| 12 | Replication | 5 replicates | 2 |
| Subtotal | | | 11 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, growth, condition factor, behavior, oxygen consumption, ammonia excretion and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Fish were fed | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | Authors state that they use an environmentally realistic concentration and compare to MEC | 1 |
| 18 | Aging and biofouling | Not addressed | 0 |
| 19 | Diversity of MP tested | One polymer type, one size, one shape | 0 |
| 20 | Exposure time | 14 days | 0 |
| Total | | | 16 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Changes in behavior, increased oxygen consumption and ammonia excretion, lower protein and lipid contents | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “ Suggesting respiration and metabolism stress (6) . Led to significantly lower protein and lipid contents, <i>suggesting energy reserve and nutrition quality reduction (2)</i> of fish.” | |

Paper: Yu et al. (2018)¹⁶⁶

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Average size given (5 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (sphere) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) and measured with FTIR in previous experiment | 2 |
| 4 | Source MP | Source given (Tianjin BaseLine ChromTech Research Centre) | 2 |
| 5 | Data reporting | Data reported in µg/L and particles/mL | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Aquaria were continuously aerated to avoid agglomeration of MP. | 1 |
| 11 | Exposure assessment of organisms | Ingestion qualitatively assessed | 1 |
| 12 | Replication | 4 replicates | 2 |
| <u>Subtotal</u> | | | <u>12</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, Growth and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 5 concentrations including control | 1 |
| 17 | Concentration range tested | According to cited reference in paper only unrealistic concentrations tested | 0 |
| 18 | Aging and biofouling | Exposure time long enough to allow biofouling of MP | 1 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 21 days | 1 |
| <u>Total</u> | | | <u>19</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Inhibition growth. Reduced activities of AChE and GPT. Decreased activity of GOT, SOD, GSH and PGx at high concentrations. Increase expression gene encoding p38 in the MAPK signaling pathway and decreased expression of genes encoding ERK, AKT and MEK. | |
| * | Demonstrated mechanism | “In addition, exposure to MP causes damage and induces oxidative stress (10) in the hepatopancreas of <i>E. sinensis</i> .” | |
| * | Speculated mechanism | - | |

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Min/max size values given (63-250 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (granule) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PS) but not confirmed | 1 |
| 4 | Source MP | No information on source | 0 |
| 5 | Data reporting | Data reported in items/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Tanks were equipped with air supply to ensure MP were well mixed | 1 |
| 11 | Exposure assessment of organisms | Ingestion visually quantified in faeces and pseudofaeces | 1 |
| 12 | Replication | Not clearly stated | 0 |
| Subtotal | | | 6 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Clearance rate, absorption efficiency and respiration rate | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations including control | 0 |
| 17 | Concentration range tested | Concentrations not compared to MEC | 0 |
| 18 | Aging and biofouling | MP were soaked in artificial seawater for a week | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, small size range | 0 |
| 20 | Exposure time | 1, 5 and 10 days | 1 |
| Total | | | 11 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Decreased clearance rate | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | <p>“The mechanism for reducing the clearance rate in <i>A. striata</i> by MP is <u>unknown</u>.” “In view of the absorption efficiency and respiration being unaffected in exposed <i>A. striata</i>, the reduction in clearance rate <i>implied</i> that energy uptake should be reduced and hence affect energy allocation to growth and reproduction, not to mention that the possible digestive interference (2) caused by the MP.”</p> | |

Paper: Zhang et al. (2017)⁸⁰

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Average size given (1 µm) and confirmed with SEM pictures | 2 |
| 2 | Particle shape | Spherical and block shapes mentioned and shown in SEM pictures | 2 |
| 3 | Polymer type | Polymer type given (PVC) but not confirmed | 1 |
| 4 | Source MP | Source given (Shanghai Youngling Electromechanical Technology Co.) | 2 |
| 5 | Data reporting | Data reported in mg/L | 1 |
| 6 | Chemical purity | Absence of additives mentioned | 1 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of the contamination | 0 |
| 9 | Verification of exposure | No verification of the exposure | 0 |
| 10 | Homogeneity of exposure | mPVC uniformly distributed, bPVC not | 1 |
| 11 | Exposure assessment of organisms | Exposure assessment of organisms (adsorption) measured qualitatively with SEM | 1 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 13 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Growth inhibition, chlorophyll and photosynthetic efficiency | 2 |
| 14 | Presence of natural (food) particles | No natural particles added | 0 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 5 concentrations including control for growth inhibition, 3 concentrations including control for other endpoints | 1 |
| 17 | Concentration range tested | No comparison with MEC | 0 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | Up to 96 h | 2 |
| Total | | | 18 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | <u>High concentrations</u> (50mg/L) of mPVC had negative effects on algal photosynthesis since both chlorophyll content and photosynthetic efficiency decreased. | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “Interactions between MP and algae and physical damage (5) may be the <i>probable</i> reason for toxic effects of MP on algae. Algae could adsorb mPVC on the surface of cells and these mPVC could wrap up caveolae on the surface.” “It could limit the transfer of energy and substance between cells and environment and lead to decrease of nutrition (2), light, CO₂ and O₂ from medium into cells. The harmful metabolite of algae also had the potential to be locked in the cell to disturb the algal growth.” | |

Paper: Zhang et al. (2019)¹¹⁵

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Particle distribution was analyzed using Beckman Coulter (5.846 ± 0.329 µm). | 2 |
| 2 | Particle shape | Shape (bead) mentioned but not confirmed | 1 |
| 3 | Polymer type | Polymer type given (PS) and confirmed with Raman spectroscopy | 2 |
| 4 | Source MP | Name of provided given (Polyscience) | 2 |
| 5 | Data reporting | Data reported in mg/L and particles/ml | 2 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of background contamination | 0 |
| 9 | Verification of exposure | concentration checked in ingestion experiment (3 days), however not in main experiment | 0 |
| 10 | Homogeneity of exposure | Sonication prior to exposure, no other measures taken | 0 |
| 11 | Exposure of assessment of organisms | Ingestion quantitatively assessed with fluorescent microscope. With digestion step | 1 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 12 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, sex ratio, developmental time, number of clutches, number of nauplii/clutch, fecundity, and suborganismal endpoints | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 3 concentrations including control | 0 |
| 17 | Concentration range tested | 2 environmentally realistic concentrations used | 2 |
| 18 | Aging and biofouling | Not addressed | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | Two generations. Short in terms of days, but effects were demonstrated. | 2 |
| Total | | | 20 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Significant reduction in survival rate, number of nauplii/clutch, and fecundity. Restored in the recovery generation | |
| * | Demonstrated mechanism | “Proteomic analysis <i>demonstrated</i> that MP exposure increased several cellular biosynthesis processes and, in turn, reduced energy storage (2) due to the trade-off, hence compromising survival and reproduction of the treated copepods.” | |
| * | Speculated mechanism | - | |

Paper: Zhao et al. (2019)¹⁶⁸

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|--|--------------|
| 1 | Particle size | Average size given (1 µm) but not measured | 1 |
| 2 | Particle shape | Shape given (sphere) and shown in pictures | 2 |
| 3 | Polymer type | Polymer type given (PVC) but not confirmed | 1 |
| 4 | Source MP | Source given (Shangai Youngling Electromechanical Technology) | 2 |
| 5 | Data reporting | Data reported in mg/L | 1 |
| 6 | Chemical purity | Not mentioned | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | Not mentioned | 0 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Forming of homo-aggregates | 0 |
| 11 | Exposure assessment of organisms | Visually shown with SEM pictures | 1 |
| 12 | Replication | 3 replicates | 2 |
| Subtotal | | | 10 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Growth, chlorophyll content and photosynthetic efficiency | 2 |
| 14 | Presence of natural (food) particles | NA for algae | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 6 concentrations including control | 2 |
| 17 | Concentration range tested | Only unrealistic concentrations used | 0 |
| 18 | Aging and biofouling | Not mentioned | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, one size | 0 |
| 20 | Exposure time | 0, 24, 48, 72, 96 h | 1 |
| Total | | | 17 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Reduced algal growth, chlorophyll content and photosynthetic efficiency | |
| * | Demonstrated mechanism | “The SEM images provided an intuitive visual method to observe the behaviors and interactions between MP and microalgae. It was found from the SEM images that microalgae were wrapped by MP beads. The physical blockage (5) and aggregation were also responsible for the cytotoxicity of <i>K. mikimotoi</i> .” | |
| * | Speculated mechanism | - | |

Paper: Ziajahromi et al. (2017)⁵⁸

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|--------------|
| 1 | Particle size | Min/max size values given for fibers (0 - 1000 µm) and spheres (1- 4 µm), but PSD only given for fibers | 2 |
| 2 | Particle shape | Shapes given (fiber, spheres) and shown only for fibers | 2 |
| 3 | Polymer type | Polymer type given for fibers (polyester) and confirmed with FTIR | 2 |
| 4 | Source MP | Source given for spherical MP (Cospheric) | 2 |
| 5 | Data reporting | Data reported in mg/L and particles/L | 2 |
| 6 | Chemical purity | Fibers cleaned with ethanol and solvent control used | 2 |
| 7 | Lab preparation | Beakers washed with ultrapure water, converting with cling wrap | 1 |
| 8 | Verification of background contamination | Controls checked visually for contamination in acute experiment | 1 |
| 9 | Verification of exposure | Not mentioned | 0 |
| 10 | Homogeneity of exposure | Description of method used to obtain well-dispersed suspension | 1 |
| 11 | Exposure assessment of organisms | Ingestion qualitatively assessed | 1 |
| 12 | Replication | At least 4 replicates | 2 |
| | | Subtotal | 18 |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Mortality, growth and reproduction | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Detection of effect thresholds (LC ₅₀ , EC ₅₀) with confidence intervals | 2 |
| 16 | Quality of dose-response relationship | 7 doses, including control | 2 |
| 17 | Concentration range tested | 1 dose is environmentally realistic | 1 |
| 18 | Aging and biofouling | No aging | 0 |
| 19 | Diversity of MP tested | 2 polymer types, 2 shapes, wide size range of fibers (more than one order of magnitude) | 2 |
| 20 | Exposure time | 8 days | 1 |
| | | Total | 30 |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Decrease in survival (acute high exposure), decrease in growth and reproduction | |
| * | Demonstrated mechanism | “Unlike previous studies, we did not observe any ingested fibers in <i>C. dubia</i> . However, malformations were <i>observed</i> in the carapace of organisms exposed to polyester fibers . This <i>demonstrates</i> that the adverse impact of MP fibers on exposed aquatic organisms is not solely due to ingestion but also external physical damage (5) , and that the latter can significantly affect survival, growth and fecundity of <i>C. dubia</i> .” | |
| * | Speculated mechanism | - | |

Paper: Ziajahromi et al. (2018)³¹

| <i>Technical subset</i> | <i>Criterion</i> | <i>Explanation</i> | <i>Score</i> |
|---------------------------------------|--|---|------------------|
| 1 | Particle size | Min/max sizes given (1 - 4, 10 - 27, 43 - 54, 100 -126 µm) but not confirmed | 1 |
| 2 | Particle shape | Shape given (bead) but not shown | 1 |
| 3 | Polymer type | Polymer type given (PE) but not confirmed | 1 |
| 4 | Source MP | Source given (Cospheric) | 2 |
| 5 | Data reporting | Data reported in particles/kg of sediment | 1 |
| 6 | Chemical purity | Not mentioned, surfactant Tween-20 added | 0 |
| 7 | Lab preparation | Not mentioned | 0 |
| 8 | Verification of background contamination | No verification of contamination | 0 |
| 9 | Verification of exposure | Verification of the exposure at the start of the experiment | 1 |
| 10 | Homogeneity of exposure | MP were homogeneously distributed throughout the sediment | 2 |
| 11 | Exposure assessment of organisms | Ingestion qualitatively assessed | 1 |
| 12 | Replication | 4 replicates | 2 |
| <u>Subtotal</u> | | | <u>12</u> |
| <i>Applicable for risk assessment</i> | <i>Criterion</i> | <i>Explanation</i> | |
| 13 | Endpoints | Survival, growth and emergence | 2 |
| 14 | Presence of natural (food) particles | Addition of food | 2 |
| 15 | Detection of effect thresholds | Effect thresholds are not reported explicitly | 0 |
| 16 | Quality of dose-response relationship | 2 concentrations including control | 0 |
| 17 | Concentration range tested | One environmentally relevant concentration | 1 |
| 18 | Aging and biofouling | No aging, pristine particles used | 0 |
| 19 | Diversity of MP tested | One polymer type, one shape, different size ranges separately tested | 0 |
| 20 | Exposure time | Up to 10 days | 1 |
| <u>Total</u> | | | <u>18</u> |
| <i>Mechanisms explaining effects</i> | | | |
| * | Effects | Our results suggest that <u>environmentally realistic concentrations</u> of MP in sediment can adversely affect the survival, growth and emergence of sediment-dwelling organism <i>C. tepperi</i> . | |
| * | Demonstrated mechanism | - | |
| * | Speculated mechanism | “This (survival and development) <i>can be</i> attributed to the longer residence time of MP in the gut of <i>C. tepperi</i> compared to sediment particles due to the <i>likelihood</i> of MP to agglomerate in the gut of <i>C. tepperi</i> , in particular for the smaller PE MP, which showed a very strong tendency to aggregate in the water phase. This could potentially result in blockage of the digestive tract (1) and consequently inhibit food uptake (2) . The delayed emergence in the presence of 100-126 µm (largest) MP <i>may be</i> explained by changing feeding capacity (2) during larvae development in the emergence assay, potentially affecting the size preference of ingested MP.” | |

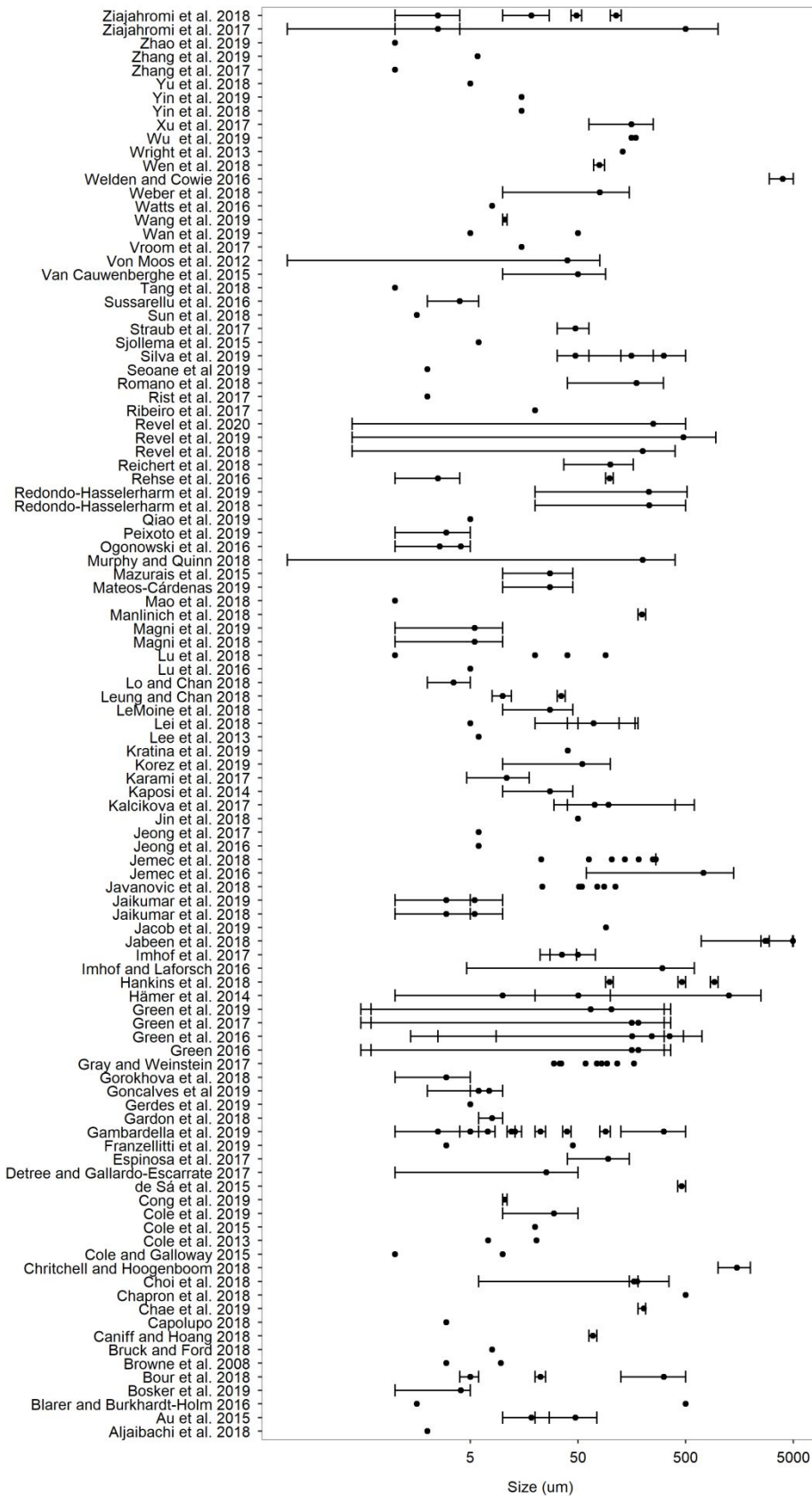


Figure S1. Size ranges used in the scored studies. Lines represent the size range reported and data points represent the reported or calculated average size.

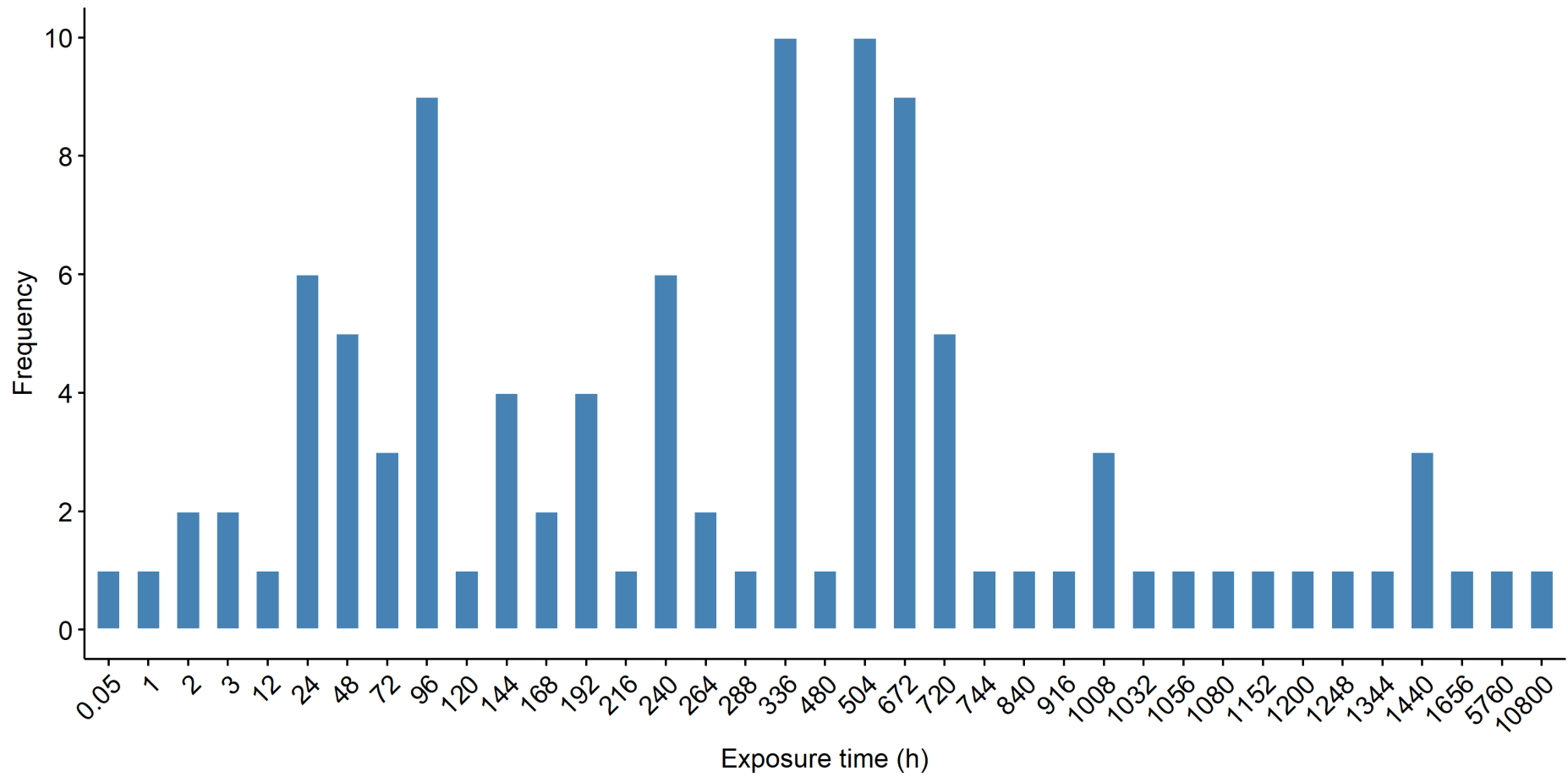


Figure S2. Exposure duration in hours for $n=105$ studies.

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